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Functional Proteomics of *Arion vulgaris* and
Drosophila melanogaster by Mass
Spectrometry Approach

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Primena masene spektrometrije u oblasti
funkcionalne proteomike na primeru *Arion
vulgaris* i *Drosophila melanogaster*

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Abstract

Functional proteomics represent an approach which could provide answers to two major questions: the revelation of biological functions of unknown proteins and the description of cellular mechanisms at the protein level. The Spanish slug, *Arion vulgaris*, is one of the hundred most invasive species in Central Europe. No transcriptomic or proteomic study on *A. vulgaris* has been carried out so far. In this thesis, the first transcriptomic database from adult specimens of *A. vulgaris* is reported. To facilitate and enable proteomics in this non-model organism, a mRNA-derived protein database was constructed for protein identification. A gel-based proteomic approach was used to obtain the first generation of a comprehensive slug mantle proteome. A total of 2128 proteins were unambiguously identified from which 48 proteins represent novel proteins with no significant homology in NCBI non-redundant database. Combined transcriptomic and proteomic analysis revealed an extensive repertoire of novel proteins with a role in innate immunity, including many associated pattern recognition proteins, effector proteins and cytokine-like proteins. The immense and very successful adaptation and spreading of *A. vulgaris* suggests that it developed highly effective mechanisms to deal with infections and natural predators. The number and diversity in gene families encoding lectins point to a complex defense system, probably as a result of adaptation to a pathogen-rich environment. These results are providing a fundamental and important resource for subsequent studies on molluscs, as well as for putative antimicrobial compounds for drug discovery and biomedical applications.

The second part of the thesis is based on a bottom-up approach to analyse an enriched membrane fraction from *Drosophila melanogaster* heads using

multidimensional liquid chromatography (LC) coupled with tandem-mass spectrometry (MS/MS) that relies on complete solubilization and digestion of proteins. Membrane proteins play key roles in several fundamental biological processes such as cell signaling, energy metabolism and transport. Despite the significance, these still remain an under-represented group in proteomics datasets. An enriched membrane fraction was prepared using equilibrium density centrifugation on a discontinuous sucrose gradient, followed by solubilization using the filter-aided sample preparation (FASP), tryptic and sequential chymotrypsin digestion of proteins. Peptides were separated by reversed-phase (RP) LC at high pH in the first dimension and acidic RP-LC in the second dimension coupled directly to an Orbitrap Velos Pro mass spectrometer. A total number of 4812 proteins from 114865 redundant and 38179 distinct peptides corresponding to 4559 genes were identified in the enriched membrane fraction from fly heads. These included brain receptors, transporters and channels that are most important elements as drug targets or are linked to disease.

Apstrakt

Funkcionalna proteomika danas predstavlja metodu koja može da ukaže na odgovor na dva pitanja: koja je biološka funkcija novootkrivenih proteina i da okarakteriše ćelijske procese na proteinskom nivou. *Arion vulgaris* predstavlja jednu od sto najinvazivijih vrsta u Evropi. Do sada u naučnoj literaturi nisu zabeleženi podaci o iRNK i proteinskom profilu ove vrste. U prvom delu teze obrađena je data problematika gde su prikazani rezultati analize iRNA na osnovu čega je napravljena baza podataka koja je korišćena za identifikaciju proteina. Ukupan broj identifikovanih proteina, dobijenih tripsinskom digestijom u gelu i analizom na masenom spektrometru, iznosi 2128. Od ukupnog broja indentifikovanih proteina, 48 nema homologe u nacionalnom centru za biotehnoške informacije (NCBI) i predstavljaju do sada neopisane proteine. Kombinacijom transkriptoma i proteoma opisan je čitav repertoar kako novih, tako i već poznatih proteina koji imaju ulogu u urođenom imunom sistemu, kao na primer „*associated pattern recognition*“, efektorski i citokinu-slični proteini. Dobijeni rezultati ukazuju na mehanizam borbe protiv infekcija i prirodnih neprijatelja, koji verovatno omogućavaju uspešnu adaptaciju i široku rasprostranjenost ove vrste. Velika raznovrsnost gena koji kodiraju lektine ukazuje na odbrambeni sistem vrste *Arion vulgaris* u sredini koji je bogata patogenima. Rezultati koji su proizašli iz ove teze pružaju fundamentalno znanje o proteomskom profilu ove vrste mekušaca, uključujući i 48 novih proteina.

Drugi deo teze je fokusiran na „*gel-free bottom-up*“ analizu proteina iz membranskih frakcija glave *Drosophila melanogaster*. Iako je genom *D. melanogaster* sekvencioniran u potpunosti, mali broj membranskih i transmembranskih proteina je

detektovan na proteinskom nivou. Oni predstavljaju značajnu grupu proteina, jer su potencijalna meta za većinu medikamenata, s obzirom da učestvuju u procesima energetskog metabolizma, ćelijske signalizacije i transporta brojnih biomolekula. U svrhu identifikacije membranskih i transmembranskih proteina korišćena je dvodimenzionalna tečna hromatografija u tandemu sa masenom spektrometrijom. Rađena je tripsinska/himotripsinska digestija proteina na membrani (FASP - filter-aided sample preparation). Peptidi su prvo razdvojeni u više frakcija pomoću tečne hromatografije na reverznim fazama pri visokoj vrednosti pH, a zatim je svaka frakcija dodatno razdvojena pomoću tečne hromatografije na reverznim fazama pri niskoj vrednosti pH i direktno analizirana na masenom spektrometru tipa Orbitrap Velos Pro. Broj identifikovanih peptida čija se sekvenca ponavlja u više proteinskih grupa („*redundant peptides*“) iznosi 114865, dok broj identifikovanih peptida koji su prisutni samo u jednoj grupi proteina („*distinct peptides*“) iznosi 38179. Ukupan broj identifikovanih proteina iznosi 4812, među kojima je identifikovan veliki broj membranskih i transmembranskih proteina, uključujući receptore, transportere i jonske kanale.

LIST OF ABBREVIATIONS

1D - one dimensional

SDS - sodium dodecyl sulphate

PAGE - polyacrylamide gel electrophoresis

MS - mass spectrometry

2D - two dimensional electrophoresis

Mud PIT - Multidimensional Protein Identification Technology

IEF - isoelectric focusing

FASP - filter aided sample preparation

HPLC - high-performance liquid chromatography

ETD - enhanced trace detection

CID - collision-induced dissociation

HCD - higher-energy collisional dissociation

RNA - ribonucleic acid

mRNA - messenger ribonucleic acid

cDNA - complementary deoxyribonucleic acid

USER - Uracil-Specific Excision Reagent

PCR - Polymerase chain reaction

TABLE OF CONTENTS

1. INTRODUCTION.....	1
1.1. Proteomics.....	1
1.1.1. Definition	1
1.1.2. Sample preparation, protein separation and enrichment	2
1.1.3. MS based proteomics	8
1.1.4. HPLC and MS	13
1.2. Transcriptomics.....	20
1.2.1. Definition, problems and perspectives	20
1.2.2. Transcriptomics analysis of model organisms and humans	22
1.2.3. Transcriptomics analysis of non-model organisms	22
1.3. <i>Arion vulgaris</i>.....	23
1.3.1. Morphology and phylogeny	23
1.3.2. Ecology and epidemiology	24
1.4. <i>Drosophila melanogaster</i>	25
2. AIMS OF THE STUDY	27
3. PUBLISHED PAPERS	27
4. RESULTS AND DISCUSSION.....	28
4.1. <i>Arion vulgaris</i> - whole body transcriptome	28
4.2. <i>Arion vulgaris</i> mantle proteome	35
4.3. <i>Drosophila melanogaster</i> membrane associated and transmembrane proteins	53
5. CONCLUSIONS	68
5.1. Whole body transcriptome of <i>Arion vulgaris</i>	68
5.2. <i>Arion vulgaris</i> mantle proteome	69
5.3. <i>Drosophila melanogaster</i> membrane associated and transmembrane proteins	69
6. EXPERIMENTAL PART	71
6.1. Chemicals, solutions and buffers	71

6.2.	Apparatus and procedures	77
6.2.1.	Protein isolation and purification	77
6.2.2.	Gel electrophoresis and staining.....	79
6.2.3.	In gel digestion	79
6.2.4.	Filter aided sample preparation (FASP).....	80
6.2.5.	Solid phase extraction	81
6.2.6.	Off-line fractionation.....	81
6.2.7.	LC-MS/MS analysis.....	81
6.3.	Transcriptome sequencing and assembly.....	83
6.3.1.	Isolation of RNA	83
6.3.2.	mRNA isolation, fragmentation and priming.....	83
6.3.3.	First strand and second cDNA synthesis	84
6.3.4.	Purifying the double-stranded cDNA.....	85
6.3.5.	End repair/dA-tail of cDNA library and adaptor ligation	85
6.3.6.	USER excision, PCR library enrichment and purifying.....	85
6.3.7.	Selection of Adaptor-ligated DNA and sequencing	86
6.4.	Bioinformatics and statistics	88
7.	REFERENCES	89
8.	APPENDIX	104
8.1.	Published papers	104
8.2.	Table 1. List of proteins identified in 2-DE proteomics study.....	127
8.3.	Table 2. The list of proteins obtained from 1D-SDS-PAGE of <i>A. vulgaris</i> mantel	148
8.4.	Table 3. Lectin-like proteins detected in transcriptomics level.....	195
8.5.	Figure 1. Alignment of amino acid sequences of H-type lectins from <i>A. vulgaris</i> with <i>Helix pomatia</i> and <i>Cepaea hortensis</i>.....	202

LIST OF FIGURES

Figure 1. Types of proteomics in biology.....	1
Figure 2. Schematic representation of isoelectric focusing of proteins according to pH. 5	
Figure 3. Fluid mosaic model of cell membrane	7
Figure 4. Trypsin active site	9
Figure 5. Schematic presentation of in-gel digestion procedure	11
Figure 6. Schematic presentation of Filter-aided sample preparation (91)	13
Figure 7. Schematic presentation of reversed-phase chromatography (93)	14
Figure 8. Schematic presentation of ionized and neutral peptide state	15
Figure 9. Illustration of main processes of ESI. TDC-total droplet current (I).	17
Figure 10. Nomenclature of ions species according to P. Roepstorff and J. Fohlman... 20	
Figure 11. Application of different sequencing techniques depending on requirements of research field	23
Figure 12. <i>Arion vulgaris</i> (<i>Arion rufus</i> var. <i>vulgaris</i> Moquin-Tandon, 1855, <i>Arion lusitanicus</i> auct. non Mabille)	24
Figure 13. <i>Drosophila melanogaster</i> , anatomy of an adult CNS) and external appearance.....	26
Figure 14. Distribution of 53,523 protein-coding sequences based on length	28
Figure 15. Distribution of number of CDSs with hits in nr database (A) and CDSs without hits in nr database.....	29
Figure 16. Specification of final assembly of <i>A. vulgaris</i> transcripts in comparison with the non-redundant NCBI database (A) and distribution of proteins in comparison with nr NCBI database (B).....	30
Figure 17. List of annotated biological function according to Gene Ontology (GO) analyses of the <i>A. vulgaris</i> transcriptome.....	31
Figure 18. List of annotated molecular function and cellular component according to Gene Ontology (GO) analyses of the <i>A. vulgaris</i> transcriptome.....	32
Figure 20. Results of blastX hits against the nr database in comparison with other species	35
Figure 21. Representative 2-DE gel from <i>A. vulgaris</i> mantel proteome study. Numbers represent spots selected for MS analysis.....	37
Figure 22. Alignment of amino acid sequences of incilarin-like proteins from <i>A. vulgaris</i>	43
Figure 23. Domain architecture of Lectin_C-like proteins from <i>A. vulgaris</i>	44
Figure 24. Domain construction of calnexin-like proteins from <i>A. vulgaris</i>	45
Figure 25. Domain architecture of three H-type lectins of <i>A. vulgaris</i>	46
Figure 26. Tandem-repeat structure of galectin like proteins identified in <i>A. vulgaris</i> ..	46
Figure 27. Domain architecture of two L-type lectins of <i>A. vulgaris</i>	47
Figure 28. Structure of R-type-like lectin protein from <i>A. vulgaris</i>	48
Figure 30. Alignment of amino acid sequences of toxin-like proteins from <i>A. vulgaris</i> with U8-agatoxine–Aol1a from <i>Agelena orientalis</i>	52

Figure 31. Schematic presentation of the three types of cystine's knots.....	52
Figure 32. Experimentally identified proteins from membrane fraction of <i>Drosophila melanogaster</i> , with respect to the number of distinct peptides	53
Figure 33. Distributions of length (A) and pI (B) of 4812 experimentally identified proteins in comparison to all proteins (20,328). Under-represented areas are shown in pink, over-represented areas in green.....	55
Figure 34. Transmembrane domain distribution among identified proteins	56
Figure 35. Under-represented "biological process" according to AmiGO2 bioinformatics tool	57
Figure 36. Under-represented "molecular function" according to AmiGO2 bioinformatics tool	58
Figure 37. Under-represented "cellular compartment" according to AmiGO2 bioinformatics tool	59

List of tables

Table 1. The most common enzymes used in proteomic analysis (75).....	9
Table 2. Types of mass analysers	18
Table 3. Detailed information on 19 proteins identified from 2-DE proteomics study without significant hit on NCBI non-redundant database database	38
Table 4. Detailed information on 44 proteins identified from 1-DE proteomics study without significant hit on NCBI non-redundant database	39
Table 5. The list of lectin like proteins detected at the transcriptome (T) and protein level (P) with status- complete or incomplete as the entire CDS from the initial ATG to the STOP codon was represented.	42
Table 6. List of ionotropic receptors detected from gel-free MS analysis of <i>Drosophila melanogaster</i> heads	60
Table 7. List of metabotropic receptors detected from gel-free MS analysis of <i>Drosophila melanogaster</i> heads.....	61
Table 8. List of channels detected from gel-free MS analysis of <i>Drosophila melanogaster</i> heads	66
Table 9. . List of transporters detected from gel-free MS analysis of <i>Drosophila melanogaster</i> heads	67
Table 10. List of used chemicals, solutions and commercial kits	71

1. INTRODUCTION

1.1. Proteomics

1.1.1. Definition

The word “proteome” is mentioned for the first time in 1995 by Wilkins et al. (1). They have postulated the concept of proteome - which represents the entire set of proteins in cells/organism at a specific time point. Besides the great technological progress of DNA and RNA sequencing, those data are not providing representable bottom line of biological samples. It is known that mRNA is not translated directly, that a journey from gene to protein is not that simple. One gene can encode more than one protein, and those proteins are the ones which define the active life of the cell (2). Hence, it is essential to determine protein expression levels at specific time points or conditions. Proteomics is a field of science whose main interest is to use quantitative protein-level measurement and/or identification of the proteins in order to characterize biological processes and functions (3). Today, we can recognize different fields of proteomic and their application (Figure 1.).

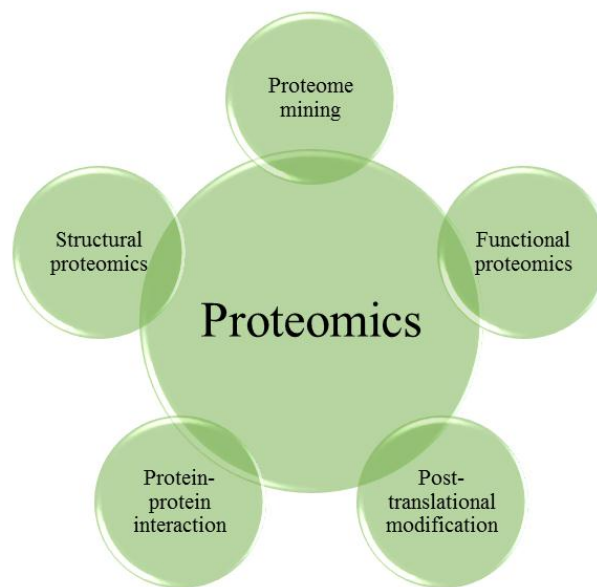


Figure 1. Types of proteomics in biology

The two major areas in the current proteomics investigations are expression and functional proteomics. Expression proteomics aims to measure protein levels (up- and down-regulation, protein-protein interaction), while functional proteomics characterizes protein activities, function, signalling pathways, etc. (4-6). Functional proteomics is “focused to monitor and analyse the spatial and temporal properties of the molecular networks and changes involved in living cells” (7). Also, the goal of this approach is to elucidate the biological function of unknown proteins, and it combines the analysis of mRNA coding and proteomics (8).

In combination with the elegant tools which have been designed to study proteins, such as high quality separation (1D and 2D electrophoresis, chromatography) and characterization by mass spectrometry, proteomics provide insight into many fields – the causes of disorders, identifying early markers of a disease process, isoforms of protein, post-translational modifications, etc. (9)

1.1.2. Sample preparation, protein separation and enrichment

Different types of biological samples are used in proteomic research: cells, tissues and body fluids (blood plasma, cerebrospinal fluid). The sample preparation is the crucial step in protein analysis and it is specific for each type of sample. It consists of solubilization, denaturation and reduction, in order to completely break up the interactions between the proteins, and removal of all chemicals which can have a negative effect on the sample separation (10).

Cell culture samples are less challenging for preparation than tissue samples, due to its simple structure. In case of tissue, composition of samples should be considered, based on its origin (lipids in brain, connective tissue in skin, etc.). In case of skin, the best way is to crush the sample in liquid nitrogen, followed by homogenisation in lysis buffer with detergents, reductants and protease/phosphatase inhibitors (11). Mechanical homogenisation (rotor-stator homogenizers, open blade mills) can be combined with ultrasonic homogenisation. Advantage of ultrasonic homogenization is that besides homogenization, it also performs emulsifying, dispersing and suspending of the mixtures(12-14).

During work on this thesis, special attention was given to presence of polysaccharides and lipids in samples for proteomic analysis. Interactions between polysaccharides and sample buffer can increase viscosity of the sample itself, which will make problems during electrophoresis (obstruction of the pores in polyacrylamide gels). If samples are analysed in 2D gels, polysaccharides will interact with ampholytes and cause prolongation of focusing time, as well as streaking visible on 2D gels(11). Moreover, since some polysaccharides are negatively charged, they may form complexes with proteins by electrostatic interaction (15). Removing sugars from samples can be done in many ways: precipitation with trichloroacetic acid (TCA), acetone, ammonium sulphate or commercially available kits.

In case of lipids, especially in the membrane fraction from the brain, they may influence protein solubility, as well as *pI* and molecular weight. Depending on the method for proteomic analysis, removing of lipids can be performed by TCA or acetone precipitation, or by centrifugation of samples with a filter device in buffer which include CHAPS or SDS (11). In case of 2D analysis of membrane proteins, heating in combination with SDS can not be used, since it will influence isoelectrical focusing IEF. The most common way of removing lipids from samples is the combination of Urea/Thiourea/CHAPS with centrifugation cut-off filters (from 3 to 100 kD). Studies have shown that this method not only removes lipids, but also salts. The yield of solubilized proteins is estimated over 90% (11, 16). The method of the sample preparation and separation depends on the ultimate goal and the type of analysis needed.

After isolation of the proteins, by native structure, usually they will be insoluble. In order to enable protein solubilization and expose cleavage places to enzymes, interactions in proteins (disulphide bonds, ionic and hydrophobic interaction) should be broken (17, 18). Depending on the method of separation, different chemicals can be used for this purpose (SDS, Urea/Thiourea, DTT, IAA, etc.).

1.1.2.1. 1D-SDS-PAGE

Polyacrylamide gel electrophoresis (19) is a frequently used technique in biochemistry, molecular biology and biotechnology, in order to separate

macromolecules (proteins or nucleic acid) based on their electrophoretic mobility. The mobility of biomolecules depends on their molecular weight. Sodium dodecyl sulphate (SDS), as an anionic detergent, is used for linearization of proteins in the sample. Also, its function is to impart a negative charge on proteins (19, 20). One-dimensional separation is used as a way of prefractionation technique for MS analysis (11). Mixture of the proteins is separated due to different molecular weights. After staining, the whole protein lane is cut into approximately 20 gel slices, and then every slice (one sample) is additionally cut into 3-5 peaces in order to enlarge the reaction surface between enzymes and proteins in gel. It is a very useful and quick method for samples with wide dynamic range, since many projects are limited by the amount of samples and cost/time of procedure and instruments (21). Choosing percentage of gel, better separation can be accomplished for low or high molecular weight. Also, separation of both high and low molecular weight can be accomplished by using gradient gels (21). After digestion and extraction, peptides are analysed on LC-MS. This combination of techniques is called gel-enhanced LC-MS (GeLCMS), and it represents a standard method in the field of proteomics (22). The advantage of SDS-PAGE is removal of low molecular weight impurities (salts, buffers and detergents) from the sample. Moreover, gel serves as a matrix for digestion by proteolytic enzymes (23). The disadvantage of this method is the possibility of traces of SDS, which in combination with low orthogonality can limit sensitivity of MS analysis (22).

1.1.2.2. 2D-IEF-SDS-PAGE

The two-dimensional gel electrophoresis allows higher resolution for the separation of proteins, as compared to 1D-SDS-PAGE. This is due to the introduction of isoelectric focusing followed by separation based on molecular weight on SDS-PAGE. IEF is the first dimension where proteins are separated according to their isoelectric point (pI). A protein's pI is defined as a pH at which the total charge of the protein is zero (24). Proteins with a more positive net charge will drift to the cathode, while those with a more negative charge will move to the anode, until they reach their pI (Figure 2.) (25). The most significant conditions for a successful IEF-based separation are an unchanged charge of proteins, and that proteins are solubilized during this process (26).

In order to achieve this, combination of neutral chaotropes (urea and thiourea) and electrically neutral detergent (CHAPS) is mostly used (27). The combination of urea/thiourea with reducing agent tributylphosphine (TBP), alkylation agent iodoacetamide (IAA) and protease inhibitors will prevent protein modification, aggregation or precipitation during IEF. Since CHAPS belongs to neutral detergents, it is stable in a wide pH range. Because of this, 4% CHAPS will not interfere with IEF of the samples and it is included in common proteomics practice (28). DTT as a reducing agent is not recommendable for first dimension (IEF), since free-thiol-containing reagents are weak acids- it may migrate during focusing. As a result, reoxidation of sulfhydryl group will occur and reduce protein solubility (29). Instead of DTT, phosphines (TBP) are common reducing agents during IEF, not only because they are neutral, but also because they will not react with acrylamide (30). At last but not the least, blocking of protease activity can change major differences in 2D profile. In absence of protease inhibitors, presence of proteins higher than 50 kD will be reduced (28). Finnie and Svensson showed that presence of protease inhibitors during IEF is crucial, especially during rehydration of IGE strips (31, 32).

A very important condition is a relatively high voltage electric field which will force proteins to travel to their exact *pI* position (33). After IEF, equilibration process with SDS will make proteins mobile in second dimension. Visualization of sample's proteome profile will be possible after staining the gel with the dyes, such as Coomassie, Blue Silver and Silver staining.

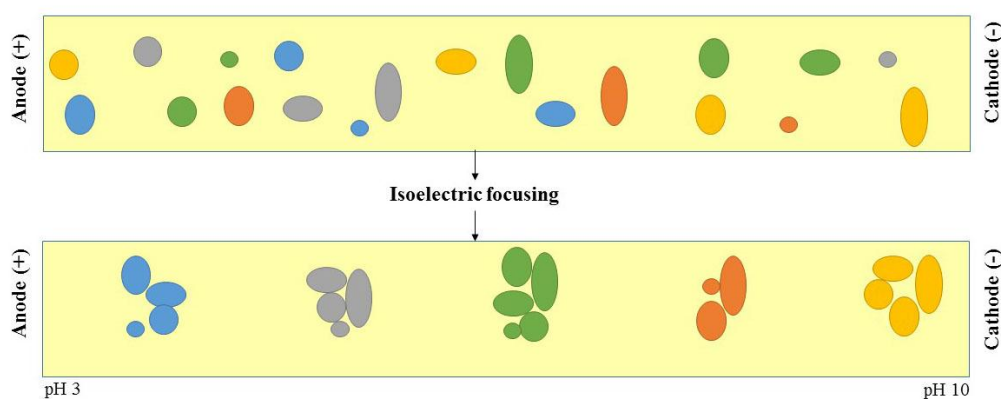


Figure 2. Schematic representation of isoelectric focusing of proteins according to pH

Afterwards, spots of interest can be processed for MS analysis. 2D gel-based proteomics is extensively used for bacterial proteomics (34), micro enzymology (35), immunoproteomics (36, 37), post-translational modifications (38-41).

The most important thing about the sample itself is the dynamic range of protein abundance, since biological samples are complex. By removing the most abundant protein/proteins from the sample (albumin, immunoglobulins from blood), complexity is reduced and sensitivity of analysis is increased (42). Besides this, low abundant proteins could be masked by high abundant proteins on 2-DE profile (11, 43). Removal of protein of interest is possible with affinity chromatography, mimetic ligands and antibodies (44-47). Despite the relatively high costs/time ratio, 2-DE provides insight to proteins as they are in organism, including potential isoforms, PTMs and charges. It is estimated that with proper choice of gel percentage, size, and pH gradient, up to 5000 proteins can be resolved (48). Also, when using more sensitive visualization staining (silver staining), only 1 ng of proteins is enough for determination of proteome profile (49). In case of comparative analysis, two-dimensional difference gel electrophoresis (2-DIGE) was developed. Two different samples (control and treated, for example) will be labeled with different cyanine dyes prior to first dimension, but they will be run and analyzed on the same second dimension, which will reduce gel-to-gel variability (50, 51).

1.1.2.3. Membrane enrichment

The plasma membrane is a structure which separates the cell from the cellular environment. It is a dynamic structure, controlling chemical trafficking in and out of the cell (52). The basic structure of the cell membrane is made of lipids, and membrane proteins are carrying out specific functions, depending on the type of the cell (cell adhesion, ion conductivity, signalling etc.). The amount of membrane proteins depends on the cell type (53). The two basic groups of membrane proteins are integral (constantly attached to the membrane) and peripheral membrane proteins (temporarily attached to the membrane) (Figure 3.) (54). It has been valued that 20-30% of the genome is encoding membrane proteins and that they represent the target for over 50%

of all modern medical drugs. Membrane proteins represent a very important part of medical and biology research, because the changes in their structure and function are responsible for the development of several disorders (55-57).

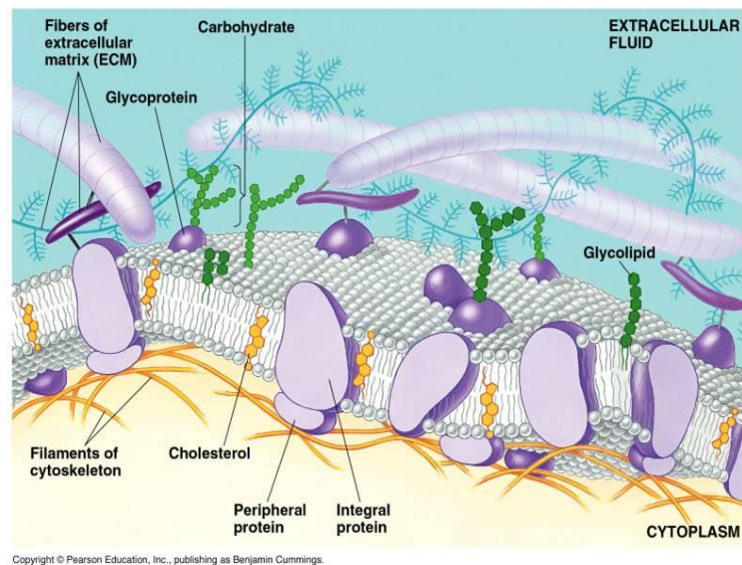


Figure 3. Fluid mosaic model of cell membrane (Picture adopted from Campbell, Neil A.; Reece, Jane B. Biology, 7th Edition)

Analysis of whole cell protein extract remains challenging because of the complexity and of wide dynamic range in abundance of proteins. Subcellular separation of proteins is a common strategy used in order to investigate protein fraction of interest, like membrane proteins which are present in relative low levels in cell (58). According to literature, density gradient centrifugation (e.g., using sucrose, sorbitol, Ficoll, or Percoll), is one of the most common techniques for the separation of plasma membranes (59, 60), mitochondria (61, 62), Golgi and synaptic vesicles (63, 64). There are two types of density gradient centrifugation: rate-zonal and equilibrium. In case of rate-zonal, sucrose is most dense on the bottom of the tube, and then density is gradually decreased to the top of the tube. In equilibrium, two or more layers of different sucrose density are loaded on tube, taking into account that the maximum density of the sucrose is higher than that of the particles of interest (samples) (65).

Besides ultracentrifugation, combination of chloroform and methanol is used in order to enrich membrane fraction. This method is based on hydrophobicity of

transmembrane and membrane associated proteins(66). In addition, Sivars and Tjereld (67) and Everberger et al.(68) describe role of detergents such as n-Dodecyl- β -D-Maltoside (DDM), TritonX-114 and polyethylene glycol (69) Advantages of aqueous two-phase systems are numerous: fast separation, stability of biomolecules, large-scale usage and supplementation of the system with protease/phosphatase (70).

1.1.3. MS based proteomics

1.1.3.1. Proteolytic digestion

Proteolysis is a biochemical process where proteins are disrupted into small peptides and amino acids. This process can be catalysed by enzymes called proteases and they are a very important component of physiological processes including digestion (trypsin, chymotrypsin), blood coagulation (factors VIIa, IXa), immune response (complement factors) and reproduction (acrosin) (71). There are seven known families of proteases, based on catalytic residues: aspartic-, cysteine-, serine-, metallo-, threonine-, glutamic- and asparagine- proteases and each of those families is divided into sub groups (72). In proteomics research the most often used enzymes are serine-proteases (trypsin, chymotrypsin, endoproteinase GluC), and aspartic-protease (pepsin) (Table 1). The enzyme's active site, called the catalytic triad, represents amino acid residues with acid-base-nucleophile motif (73). In case of serine-proteases amino acid residues are histidine, aspartate, and serine. Modifying the electrostatic environment of the serine by increasing nucleophilicity is possible by forming a charge between catalytic triad components. In addition, some enzyme's (trypsin) backbone amide hydrogen atoms of Gly-193 and Ser-195 form an "oxyanion hole" which additionally stabilizes negative charge on the carbonyl oxygen atom of the cleaved amides (Figure 4.) (74).

Table 1. The most common enzymes used in proteomic analysis (75)

HIGHLY SPECIFIC ENDOPROTEASE		
Enzyme	pH of activity	Specificity
Trypsin	7-8,5	Arg, Lys
Arg-C	7,2-8	Arg
Glu-C	7,5-8,5	Arg, Glu
Lys-C	8-9,5	Lys
Asp-N	6-8,5	Asp, Cysteic acid (Glu)
Prolyl endopeptidase	7-7,5	Pro
LESS SPECIFIC ENDOPROTEASE		
Enzyme	pH of activity	Preference
Chymotrypsin	7-9	Phe, Trp, Tyr
Pepsin	2-4	Phe, Met, Leu
Proteinase K	6,5-9,5	Aliphatic residue
Thermolysin	7-9	Leu, Ile, Phe, Trp, Met, Val

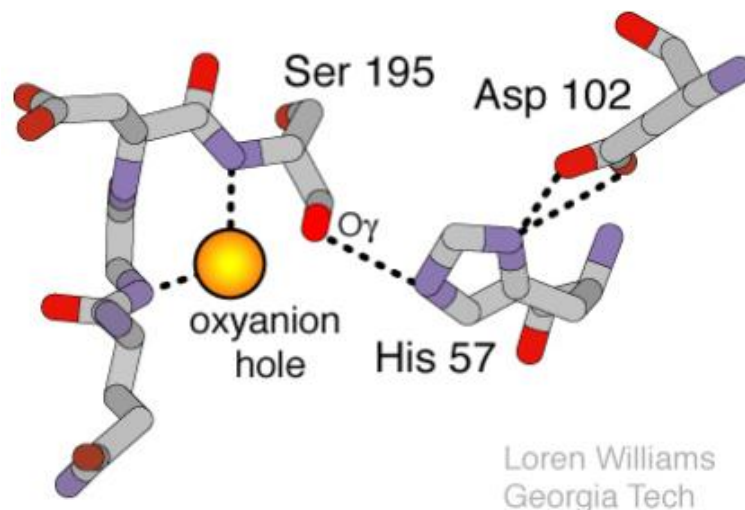


Figure 4. Trypsin active site (The picture is adopted from www.proteopedia.org)

To accomplish the enzyme's maximum access to the protein cleavage sites two important steps are included in sample preparation - reduction and alkylation.

Dithiothreitol (DTT) is the most often used reducing agent. After exposure of samples to DTT solution, cysteine disulphide bonds are reduced into cysteine free sulfhydryl groups. Reduction step is followed by alkylation with iodoacetamide (IAA), which reacts with free sulfhydryl groups of cysteine residues to form S-carboxyamidomethyl-cysteine. In this way sulfhydryl groups cannot be reoxidized (76).

1.1.3.1.1. In-gel digestion

In-gel digestion of samples for MS analysis (Figure 5) is a method which was introduced in 1992 by Rosenfeld (77). This method has some important advantages in comparison to gel-free approaches, such as: fractionation at protein level decreases the complexity of the sample, and in some cases allows identification of post/translational modifications, especially with by mass analysis after 2D-IEF-SDS-PAGE (78).

Optimization of digestion conditions for in-gel digestion is crucial in order to enable the reaction between protein and enzyme. Digestion of proteins in polyacrylamide matrix is restricted by enzyme diffusion and concentration, temperature and pH (79). Comasine, as a gel dye, must be completely removed in order to enable enzymatic cleavage and reduce background during MS analysis of peptides. (80). Extraction of proteins from gel matrix is a multistep process, where in the first step most of peptides are eluted in solution, and an additional step can increase yield of the whole process by only 5-10% (81). Efficiency of extraction depends on many factors, such as thickness of gel, size of gel pores (gel percentage), physico-chemical properties of peptides (e.g. hydrophobicity, *pI*, hydrodynamic radius). It is easier to extract peptides (especially large ones 7-10 kD) from lower percentage of gel than from higher percentage, due to acrylamide pore size (81). Additionally, if it is necessary, gel plug can be submerged in deionized water and freeze. This process will crash pores and ensure recovery of large peptides. Shevchenko and colleagues published a protocol which suggests combination of formic acid/acetonitrile as extraction solution (82). Formic acid will ensure extraction of hydrophilic peptides, while organic solvent will ensure extraction of hydrophobic peptides from gel plug.

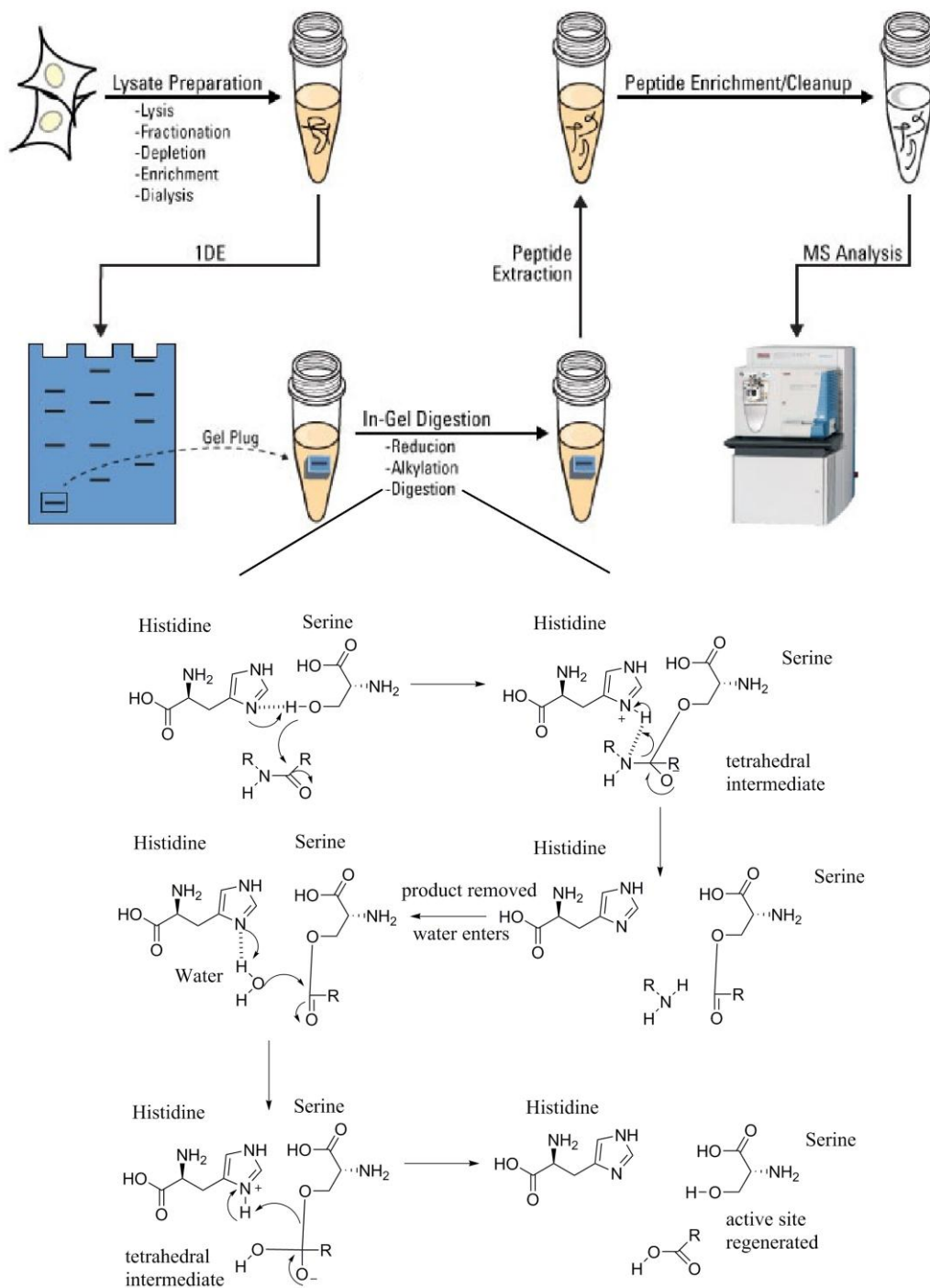


Figure 5. Schematic presentation of in-gel digestion procedure (The picture was adopted and modify from www.sites.psu.edu)

1.1.3.1.2. In-solution digestion

In contrast to the in-gel digestion, in-solution digestion is more efficient, in a sense of producing higher peptide yields and sequence coverage. The digestion protocol is designed in such a way to eliminate salts and detergents which are not compatible with MS analysis, and to keep proteins soluble. Urea is a commonly used denaturant for in-solution digestion (75). Besides this, during the development of the protocol, a great attention has been paid to a non-detergent washing of sample and an elution in low pH range. Usually, in-solution digestion is followed by separation of peptides on HPLC. The most common approach is Multidimensional Protein Identification Technology - Mud PIT strategy (gel-free technique), which enables high resolution separation of peptides and individual identification of complex samples (83).

1.1.3.1.3. FASP

Using gel-free and in-solution digestion enables removal of impurities (detergents, salts, SDS, etc.) and minimizes sample handling. Nevertheless, digestion obstruction and incomplete solubilization of proteins can be potential problems, especially when working with membrane proteins (83). Before usage of SDS, solubilization of membrane proteins was done with formic acid and/or organic solvent (84, 85). Filter-aided sample preparation (FASP) is a universal method of sample preparation for MS analysis. The advantage of this method is that membrane will keep proteins, but not small molecules and peptides. Filter is a "proteomic reactor", which gives us different possibilities for MS sample preparation: detergent removal, buffer exchange, chemical modification (reduction and alkylation) and protein digestion with different enzymes (Figure 6). Numerous citations show that FASP is an appropriate method for proteomic analyses of bacteria, mammalian cells, yeast and paraffin-embedded tissue samples (86-88). Also, it has been successful for membrane proteins, phosphoproteins and N-glycoproteins (89, 90).

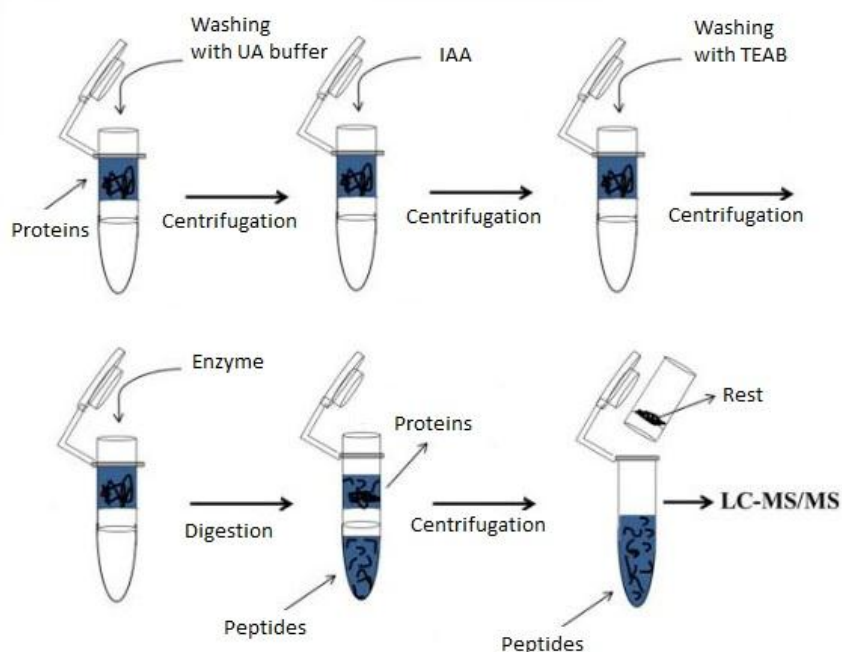


Figure 6. Schematic presentation of Filter-aided sample preparation (91)

1.1.4. HPLC and MS

1.1.4.1. HPLC and pre-fractionation

Chromatography is an analytical technique for separating a mixture of chemical substances into individual components based on different properties of each component. In principle, the separation of components depends on the affinity of the constituents to the stationary phase and their solubility with mobile phase.

High-performance liquid chromatography or high-pressure liquid chromatography (HPLC) utilizes a solid stationary phase with a liquid mobile phase. The operational pressure during the separation is between 50 and 350 bar, which is significantly higher than in regular chromatography. The sample volume is smaller (few microliters), the diameter of the analytical column is 2.1-4.6 mm and it is packed with sorbent particles the size of 2-50 μm . All of those physical characteristics give HPLC a high resolution in separation of mixture of compounds.

In proteomics, the most commonly used HPLC system for the separation of peptides is the reversed-phase liquid chromatography (RPLC). This involves the separation of peptides on the basis of their hydrophobicity. The RPLC is performed on a hydrophobic stationary phase (C18) with an acidic gradient polar mobile phase. The stationary phase consists of a polymeric C18 format (octadodecyl hydrocarbon) integrated via a tri-functional silylation of 2 or 3 silica atoms on the silica gel backbone. This increases the silylation results with a greater column stability, particularly in acidic mobile phase conditions. The hydrophilic peptides pass through the column and are eluted first; this results in the successive elution of hydrophobic peptides by changing the composition of the mobile phase (increasing the percentage of the organic solvent) (Figure 7) (92).

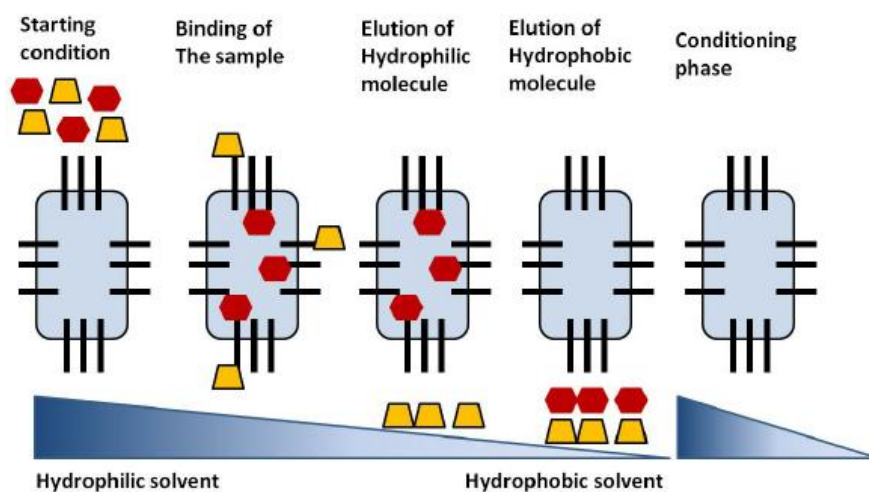


Figure 7. Schematic presentation of reversed-phase chromatography (93)

Given the wide-range of proteins in of biological samples, 2-dimensional peptide-level separation can be performed to reduce sample complexity. This is accomplished by employing two or more methods with different separation selectivities prior to the actual analyses (94). Two-dimensional liquid chromatography, 2D-LC, consists of a liquid chromatography for the pre-fractionation peptides followed by the individual LC MS/MS analyses of the separated fractions. The current separation strategies/modes include: strong cation exchange (SCX), isoelectric focusing (IEF), capillary electrophoresis (CE), capillary isoelectric focusing (CIEF), and mixed mode

pH reverse phase (RP-RP). A strong cation exchange (SCX) was used as a first dimension (95). Traditionally, SCX was the classical peptide pre-fractionation approach. Methods other than SCX were developed as there is a need for sample desalting prior to LCMS analyses leading to sample loss of up to 50%. Further, resolving of peptides in ionic buffers is a concern due to ion suppression of species during MS analyses.

The advent of 2D RPLC/RPLC that utilize buffers that are low in salt or salt-free solutions eventually replaced SCX. MudPIT (2D RPLC/RPLC) is a chromatography-based proteomic technique where a complex peptide sample is separated first into fractions and then every fraction is separately analysed on the RPLC-MS/MS system. Gilar et.al (96) demonstrated that different pH of the mobile phase has a crucial impact on the quality of RPLC on Mud PIT analysis. Peptides contain carboxylic (RCOOH) or basic (RNH₂) functional groups. When the pH of buffer is equal to the isoelectric point of a peptide, the peptide will be in both states: ionized and neutral (Figure 8).

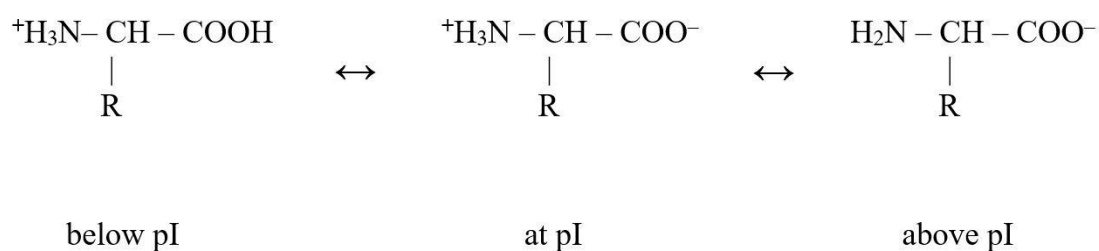


Figure 8. Schematic presentation of ionized and neutral peptide state

The zwitterionic state has poor retention in the reversed phase columns. Peptide retention can be improved by modulating the pH of the aqueous mobile phase. At low pH, positively charged species dominate and at high pH, negatively charged species dominate. In each of these two conditions (below or above its isoelectric pI), the analyte acquires an “ion-suppressed state“. Elution of the retained peptides is then accomplished by increasing the organic solvents gradient (ACN, MeOH, iPrOH). By

pre-fractionating the peptides at pH 10 followed by the LC MS/MS at pH 2.6, the highest orthogonality was accomplished with C18 (96).

1.1.4.2. Mass spectrometry

Mass spectrometry is an analytical, instrumental technique that can identify and/or quantify ionized chemical species based on mass to charge ratio. The technique was developed by A. J. Dempster (1918) and F.W. Aston (1919). The basic components of a mass spectrometer are an ion source, a mass analyser and a detector (97).

1.1.4.2.1. Ion source

Nowadays, we can distinguish different kinds of ionization. The most used ones in chemical/biochemical research are:

- a) Fast atom bombardment (FAB)
- b) Chemical ionization (CI)
- c) Atmospheric-pressure chemical ionization (APCI)
- d) Electrospray ionization (ESI)
- e) Matrix-assisted laser desorption/ionization (69)

Charged species are generated by an ion source, travels to a mass analyser of which signals are recorded in a detector. Data processing of the signals provides an output of the mass to charge ratio (m/z , Thomsons) with the relative abundance of each ion in sample (98). In this study, an ESI source was used to ionize the positively charged peptides in liquid to gas phase (ESI/MS). The ESI/MS technique invented by John Fenn paved the way for his Nobel Prize award for chemistry in 2002 in recognition for its development and application in biochemical, biomedical and pharmacological research.

The analysis of complex peptide mixtures is enabled by coupling the MS instrument as a mass detector, and the high separation resolution of the HPLC.

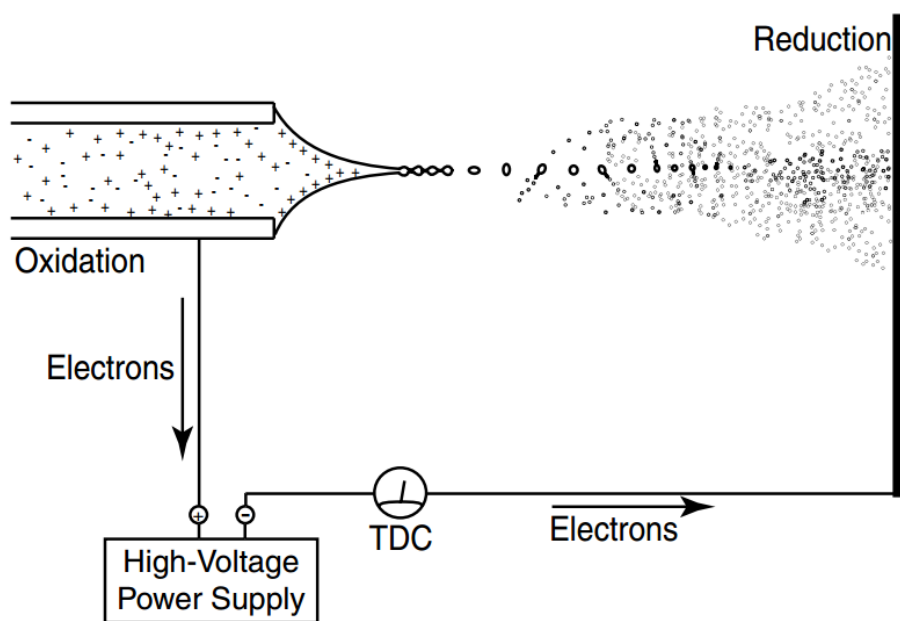


Figure 9. Illustration of main processes of ESI. TDC-total droplet current (I).
 (Copyright: Paul Kebarle and Udo H. Verkerk, On the Mechanism of Electro spray
 Ionization Mass Spectrometry (ESIMS))

The curtail events of ESI that occur at atmospheric pressure are presented in Figure 9. Electrical double layer will be formed as a consequence of infiltration of the established electric field into the liquid, due to the dipole moment of the solvent, which will enable formation of the charged jet by a large amount of positive ions (positive ion mode). Gas-phase ions will be formed in the process called Coulomb effect. During the process of evaporation, charged droplets (formed from jet) will become smaller and positive charges will be closer to each other. As a result, Coulombic repulsion of positive charge will destabilize the droplet, and it will emit a jet of smaller positive charged droplets. This process will be repeated, together with a solvent evaporation, until the gas-phase ions are formed (99). Ions are guided by an electrical potential and pressure differences, while solvent vapour will be removed by nitrogen. "Cleaning-up" of ions will be performed by electrical field and high vacuum system, in order to accelerate ions to mass analysis chamber.

1.1.4.2.2. Mass analyzer

The first MS and mass analysers were developed during the 1980s, but it was in the past 20 years that several types of mass analysers have been established (Table 2). A mass analyser is the part of the MS which is isolated from the outer surroundings, and it is a combination of electric/magnetic fields. Its function is to capture charged particles and separate them based on m/z ratios. The choice of mass analyser type to be used depends on the application and the desired performance (100).

Table 2. Types of mass analysers

Type of analyser	Symbol	Principle of separation
Electric sector	E or ESA	Kinetic energy
Magnetic sector	B	Momentum
Quadrupole	Q	m/z (trajectory stability)
Ion trap	IT	m/z (resonance frequency)
Time-of-flight	TOF	Velocity (flight time)
Fourier transform ion cyclotron resonance	FTICR	m/z (resonance frequency)
Fourier transform Orbitrap	FR-OT	m/z (resonance frequency)

For the performance of the mass analyser, five main points have to be taken into consideration: mass range limit, analysis speed, transmission, mass accuracy and resolution. Mass range limit is expressed in Thomsons (Th) which is based on *mass-to-charge* ratio. The analysis speed (scan speed) describes the speed rate of mass analyser in specific mass range expressed in mass units per second or mass units per millisecond ($u\ s^{-1}$ or $u\ ms^{-1}$). The transmission represents the ratio of ions that reach the detector over number of ions that entered the mass analyser. The mass accuracy represents the ratio or difference between $m_{theoretical}$ and $m_{measured}$ expressed in parts per million (ppm) or milimass units (mmu). The resolution or resolving power is the ability of MS to distinguish two ions with close m/z values (100).

In the course of this study, two types of mass analysers were implemented: 3D ion trap and Orbitrap. Using an oscillatory electric field, ion trap stores ions prior to selected analysis.

The first use of modifying ion trap was performed during 1920 and described by Kingdon (101). The 3D ion trap (quadrupole ions traps - QITs) consists of a circular electrode with two ellipsoid caps on top and bottom. The invention of the 3D ion trap awarded Wolfgang Pauli a Nobel Prize in physics in 1989. The Orbitrap mass analyser consists of an outer cylinder-like electrode and a coaxial inner spindle-like electrode that traps ions in an orbital motion around the spindle (102). It was designed and patented by Makarov and was commercialized by Thermo Scientific in 2005. The conversion of the signal into a mass spectra is enabled by Fourier transformation.

Depending on the kind of ion trap and/or combination of other mass analysers (e.g., 2D linear ion traps), one can distinguish LTQ Orbitrap (linear ion trap), Q Exactive (Quadrupole mass filter), as well as MALDI LTQ Orbitrap (103).

1.1.4.2.3. Ion activation

Tandem mass spectrometry (MS/MS or MS²) implements additional mass analyses after a mass selection/isolation and ion activation/fragmentation. Ion activation can be done based on different mechanisms: collision-induced dissociation (69), electron capture dissociation (ECD), electron transfer dissociation (ETD), negative electron transfer dissociation (NETD), electron-detachment dissociation, charge transfer dissociation (CTD) and a special group of activation called photodissociation (104-106). In proteomics research, the commonly used ion activation techniques are:

- a) CID – activation under impact of ion in gas phase and neutral atom or molecule. Higher-energy collisional dissociation (HCD) is a type of CID typically coupled with Orbitrap mass analysers.
- b) ETD – activation helped by radical anions (fluoranthene). This kind of activation is used for studying of peptide modification, such as phosphorylation.

Depending on the type of ion activation used, fragmentation occurs at distinct peptide bond positions to generate a highly predictable ions species.

In Figure 10 the nomenclature of the generated fragment ions is shown, as previously described by Roepstorff and Fohlman (107). If a charge of a fragment is maintained on the N terminal, ions will be a, b or c, but if it is located on the C terminal than it will be x, y or z (107, 108). CID activation (i.e., HCD) typically cleaves peptides to generate b and y ions species whereas ETD activation generates c and z fragment ions.

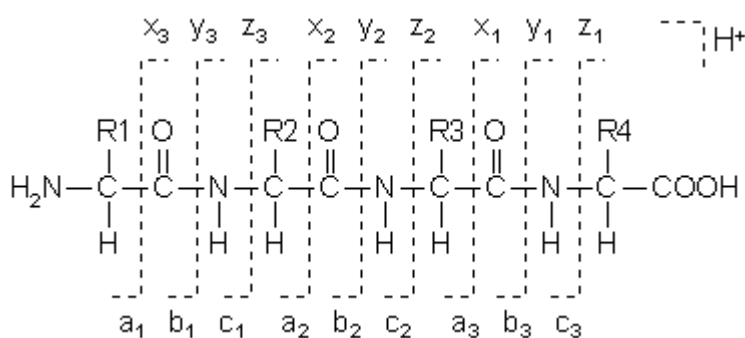


Figure 10. Nomenclature of ions species according to P. Roepstorff and J. Fohlman

1.1.4.2.4. Detector

At the rear end of a mass spectrometer is a detector that measures ion signal intensities proportional to their abundance. Detectors can be divided into two main groups: point ion and array detectors. Point ion detectors measure signals of a single mass in the time point, whereas array detectors detect multiple ion masses simultaneously (109). In both cases, the signals are amplified and processed under different computational platforms for large scale analyses.

1.2. Transcriptomics

1.2.1. Definition, problems and perspectives

Transcriptomics has been a fast evolving field of science during the last years, due to technological development. First concept of transcriptome was

implemented in 1958 with the establishment of central dogma of biology by Francis Crick. He suggested that coding information from DNA is transcribed to RNA. Eventually, it was shown proposed that RNA is used for synthesis of proteins of specific sequence (110). Later on, with the discovery of ribosomal and transfer RNA, and coding (exons) and non-coding sequences (introns), the significance of RNA was elevated to a different level. Today it is known that RNA can be divided into two groups: protein-coding (mRNA) and protein-non-coding RNAs (ncRNAs) (111). Protein-non-coding RNAs are house-keeping RNAs (translation-tRNA; ribosomal-rRNA; small nuclear-snRNA; small nucleolar-sonRNA; guide-gRNA) (112) and regulatory RNAs (antisense-aRNA; CRISPR-crRNA; long noncoding-lncRNA; micro-miRNA; piwi-interacting-piRNA; small interfering-siRNA; short hairpin-shRNA) (113-117). Together, all this molecules shape the future of the cell and organism.

Due to the complexity of biological organisms, one of the first questions was - can we see a correlation between mRNA expression and proteins? Some experiments have shown that the central dogma of molecular biology gives a positive correlation (118-120), while in some cases, there is no significant correlation between mRNA and protein levels (121). Nie and coworkers speculate that the absence of correlation can be caused by biological and/or experimental reasons, such as regulation of translation, half-life differences of mRNA/proteins and methodological limitations (e.g. sample preparation, normalization of data) (122). Eukaryotic polysomes can bind active mRNA and begin a process of protein synthesis, while the mRNA molecule, which is inactive, will be strongly attached to monosomes. There is a constant flow of those two types of mRNA – active and inactive. Besides this, lack of correlation can be a consequence of rapid degradation of mRNA as an answer to different stimuli (123). A combination of proteomics and transcriptomics gives a significant insight in all applied fields of research, such as cancer biology (124), neurosciences (125), ecology, zoology (126) etc. Nevertheless, besides level of mRNA, other processes also have a regulatory function. The most important ones are in proteins, like post-translation modifications (PTMs), which are able to regulate the level of mRNA, protein synthesis and protein activation (127). A large-scale proteomics studies (Mud PIT, tandem MS/MS) in combination with transcriptomics may untangle the regulatory mechanisms and complex signalling networks in living organisms (128-130).

1.2.2. Transcriptomics analysis of model organisms and humans

Collected basic knowledge about biological processes has been expanded thanks to the extensive studies on animal and plant model organisms. The most used ones are *Escherichia coli*, *Saccharomyces cerevisiae*, *Caenorhabditis elegans*, *Drosophila melanogaster*, *Mus musculus*, *Rattus norvegicus*, *Arabidopsis thaliana*, *Danio rerio* and *Xenopus laevis* (131). Common in all these cases is that they are well studied and that their genomes have been sequenced, together with genetic material from mitochondria/chloroplast. Transcriptomics analysis of model organisms is a powerful tool used in order to investigate mRNA changes under a special treatment or condition, and how it is different from the already known mRNA status (132, 133). On the other hand, transcriptomics in human pathologically changed tissues have a tremendous impact on collected knowledge about expression profile and/or response to therapy (134, 135). Alternative to model-organisms is a broad spectrum of *in vitro* studies in different types of cell cultures. Besides this, it is possible to have an animal model of a disease. One of the most applicable and widely used is the transcriptomics profile of cancer affected patient (136), where it is relatively easy to collect tissue. Those results may have impact to improve cancer therapy (137).

1.2.3. Transcriptomics analysis of non-model organisms

In contrast to model organisms, non-model organisms are not well studied. With the development of next generation sequencing (138), numerous possibilities are unlocked for the benefit of evolutionary biology, phylogenetics and population genetics (139). Depending on the type of desired analysis, there are several options of sequencing that can be chosen (Figure 11). For example, whole genome sequencing is applicable to simple biological entities such as bacteria, in order to study evolutionary responses (140), while targeted sequencing, like sequencing of ultra-conserved elements, can be used for comparative genomics studies between large species radiations (141, 142). Transcriptomics analysis of non-model organisms is quite challenging, especially if there is no known genomics background of species. Todd et. al. have shown that

biological variance is more expected in field studies of certain species than in *in vitro* experiments (143).

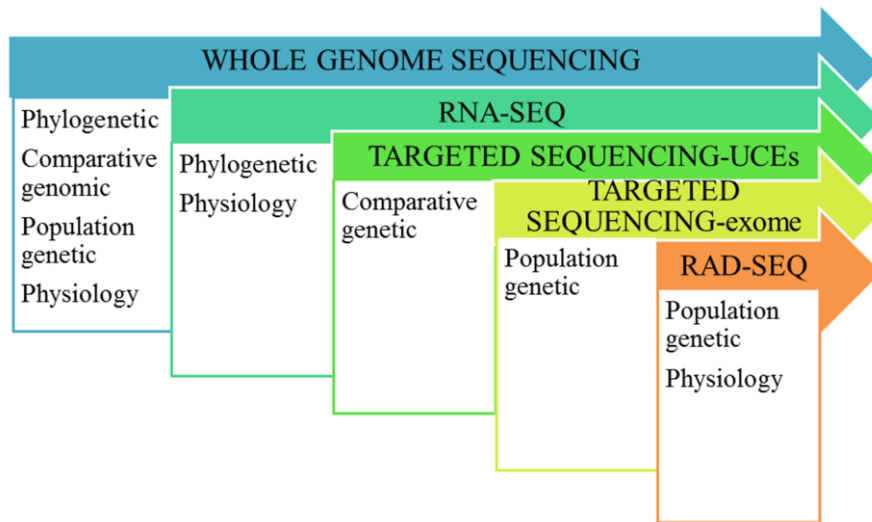


Figure 11. Application of different sequencing techniques depending on requirements of research field (144)

1.3. *Arion vulgaris*

1.3.1. Morphology and phylogeny

Arion vulgaris (Figure 12) was described for the first time by the French naturalist and zoologist Christian Horace Benedict Alfred Moquin-Tandon in 1855. *Arion vulgaris* belongs to phylum Mollusca, class Gastropoda, family Arionidae. Distribution of the family Arionidae is Holarctic and Oriental. The main characteristics of the genus *Arion* (over 40 species) are the absence of internal shell and keel. The animals are 80-150 mm long, brownish or reddish-brown in colour. The caudal mucous pit is located on the tail. The mantle is positioned on the anterior part and covers only one part of the body (145). Like all Pulmonata, *Arion* is a hermaphrodite and the morphology of its reproductive system is a key characteristic to distinguish different species of the genus.



Figure 12. *Arion vulgaris* (*Arion rufus* var. *vulgaris* Moquin-Tandon, 1855, *Arion lusitanicus* auct. non Mabilie)

1.3.2. Ecology and epidemiology

Habitat of the species *Arion vulgaris* are natural (rivers and lakes, forests or dry valleys, mountains – up to 1700 m), as well as cultivated ecosystems (horticultures) (145). Only a small number of animals can be considered as natural enemies (hedgehogs, domestic ducks, some amphibians, reptiles and beetles) of *Arion vulgaris*. Nevertheless, according to DAISIE (Delivery Alien Invasive Species Inventories for Europe), *Arion vulgaris* was classified among 100 worst pests in Europe in 2006. As pests, slugs are characterized by high reproduction rate, high survival rate, sticky mucus and a large body size for a snail. *Arion vulgaris* can live up to two years. According to the research analysis, it has a huge effect on the economy, ecology and health (146). The economic impact is still not precisely calculated, but it clearly represents a huge problem in private gardens and agriculture. Research published in American Malacological Bulletin, from 2009, reports that costs from alien species invasions in United States, United Kingdom, Australia, South Africa, India, and Brazil have been estimated at US\$314 billion per year (147).

Last year's numerous studies have been reporting that *Arion vulgaris* may be a vector for pathogenic species, such as *Clostridium botulinum* (148) and *Listeria monocytogenes* (149). Those findings play a crucial role in agriculture, because they have reported contamination of silage food for cattle. Calculated borderline for the infection is approximately 50 animals per square meter (150). Botulinum neurotoxin

(BoNT A-F) blocks the acetylcholine release in the motor-nerve connection and leads to paralysis and death. *Clostridium botulinum* is very heterogeneous and it is classified into four strains where we can recognize different types of BoNT (C, D - cause of animal botulism; A, B, E and F - human botulism; A and B - bovine botulism outbreaks) (151, 152). *Listeria monocytogenes* is a gram positive bacteria that has an effect on the central nerve system (CNS) and can cause encephalitis. Besides this, it can also cause enteritis, mastitis and septicemia (149). In ruminants, listeriosis is a usual silage associated disease. It has an indirect effect on human listeriosis, due to the fact that fresh milk has been used for production of cheese (153).

In this thesis, we present results from parallel transcriptomics and proteomics analysis of *Arion vulgaris* mantel, in order to identify proteins responsible for its successful survival strategies.

1.4. *Drosophila melanogaster*

Drosophila melanogaster, also known as fruit fly or vinegar fly, belongs to the class Insecta, order Diptera and the family Drosophilidae. It has been used as a model organism for many fields of research, due to the following characteristics - undemanding keeping, presence of sexual dimorphism, 4 pairs of chromosomes, easy and quick reproduction (154). Since 1970s and 1980s, *D. melanogaster* has been a key model organism to uncover gene function, understand development control and genetic origin of disease. Around 75% of human disease genes have homologues in *Drosophila*, such as dysmorphology (155) (single bone in zeugopod, apert syndrome, crouzon syndrome and pfieffer syndrome (156), etc.), cardiac disease (congenital heart disease, venous malformations (157)), neurological disorders (158) (Huntington disease, Parkinson disease (159), Alzheimer disease (160)), cancer (endometrial carcinoma, tuberous sclerosis, melanoma (161)). Explanation for similarity of genetic pathways is that during evolution the control of those processes was intact. Results from investigations of *Drosophila* can be directly applied to vertebrate systems (162).

Including the above mentioned, this invertebrate model system was particularly interesting for the investigation of potential drugs and their mechanisms.

While the genome of *Drosophila* has been decoded and well annotated, 63% of proteins (9124 proteins) were covered by a few proteomics studies (163). 2D analysis of fly heads and brains show approximately 90% of sequence coverage and according to this, heads can be considered for studies of central nervous system (Figure 13) (164). Due to the limitations of 2D electrophoresis, only a few membrane proteins were identified, so studies are still relying on computational predictions.

During work on this thesis, we were focused on membrane proteins, since they are usually drug targets and they have not been successfully investigated until this point. Gel-free bottom-up proteomics approach was used for enriched membrane fraction from fly heads.



Figure 13. *Drosophila melanogaster*, anatomy of an adult CNS (CenBr- central brain hemispheres; OL- olfactory lobe; SubGgl- suboesophageal ganglia; cn- cervical connective; ThAGgl- thoracic ganglia; according to Atlas of Drosophila Development by Volker Hartenstein published by Cold Spring Harbor Laboratory Press, 1993) and external appearance

2. AIMS OF THE STUDY

During work in this thesis, we defined the following aims:

- a) Whole body transcriptome of *Arion vulgaris*, in order to create an mRNA-derived protein database
- b) Proteomic profiling of *Arion vulgaris* mantel using nano-LC-ESI-MS/MS analysis.
- b) Identification of membrane associated and transmembrane proteins from *Drosophila melanogaster* heads using gel-free MS/MS approach

3. PUBLISHED PAPERS

Results presented in this thesis are published in two peer-reviewed journals:

1. Bulat T, Smidak R, Sialana FJ, Jung G, Rattei T, Bilban M, et al. *Transcriptomic and Proteomic Analysis of Arion vulgaris--Proteins for Probably Successful Survival Strategies?* PLoS One. 2016;11(3):e0150614
2. Aradska J, Bulat T, Sialana FJ, Birner-Gruenberger R, Erich B, Lubec G. *Gel-free mass spectrometry analysis of Drosophila melanogaster heads.* Proteomics. 2015;15(19):3356-60

4. RESULTS AND DISCUSSION

4.1. *Arion vulgaris* - whole body transcriptome

Since there are no information about genome or transcriptome, we used RNAseq and *de novo* assembly in order to create the *Arion vulgaris* database. Using Illumina HiSeq 2000 technology we managed to create a cDNA library which contains 339 millions of paired end reads. Using Trinity pipeline (165) assembly of *de novo* transcript resulted in 136,406 contigs. 50% of sequences are 971 base pairs (bp) or longer (NP50), while average length was 671 bp. Combining TransDecoder tool with PFAM version 27.0, we identified 53,523 protein-coding sequences (CDS), illustrated in Figure 14.

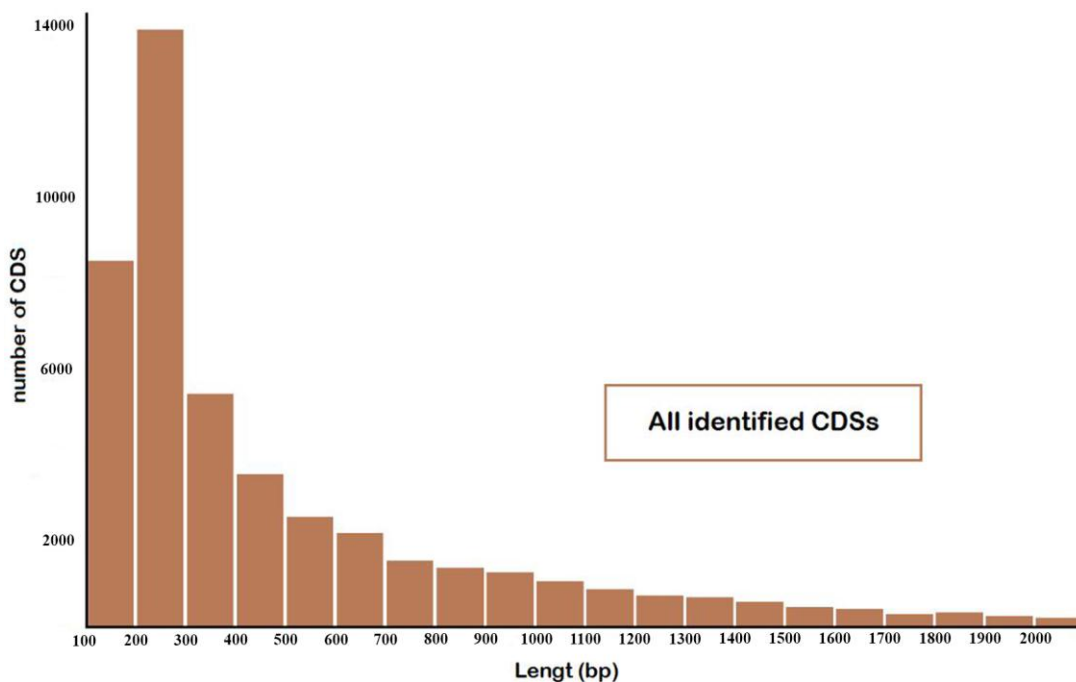


Figure 14. Distribution of 53,523 protein-coding sequences based on length

In order to reach final assembly, all CDS annotated contigs were analysed against the NCBI non-redundant (nr) protein base. In this analysis we used FastAnnotator and cut-off value was set up to $1E-6$ (166). Duplicates and substrings were removed from the translated sequence dataset and resulting protein sequences were post-assembled using an in-house script to reduce redundancy. In Figure 15 arrangement of number of CDSs with hits in nr protein data base (A) and without hits in nr protein database (B) is

presented. The final list of CDSs that we published contains 47,451 non-redundant protein sequences. The full list of non-redundant protein sequences is available as supplementary material (Table S1) published in Bulat T. et al. (167). The sequence data has been submitted to the sequence read archive (69) database of GenBank (<http://www.ncbi.nlm.nih.gov/sra>) with the BioProject accession number PRJEB7891.

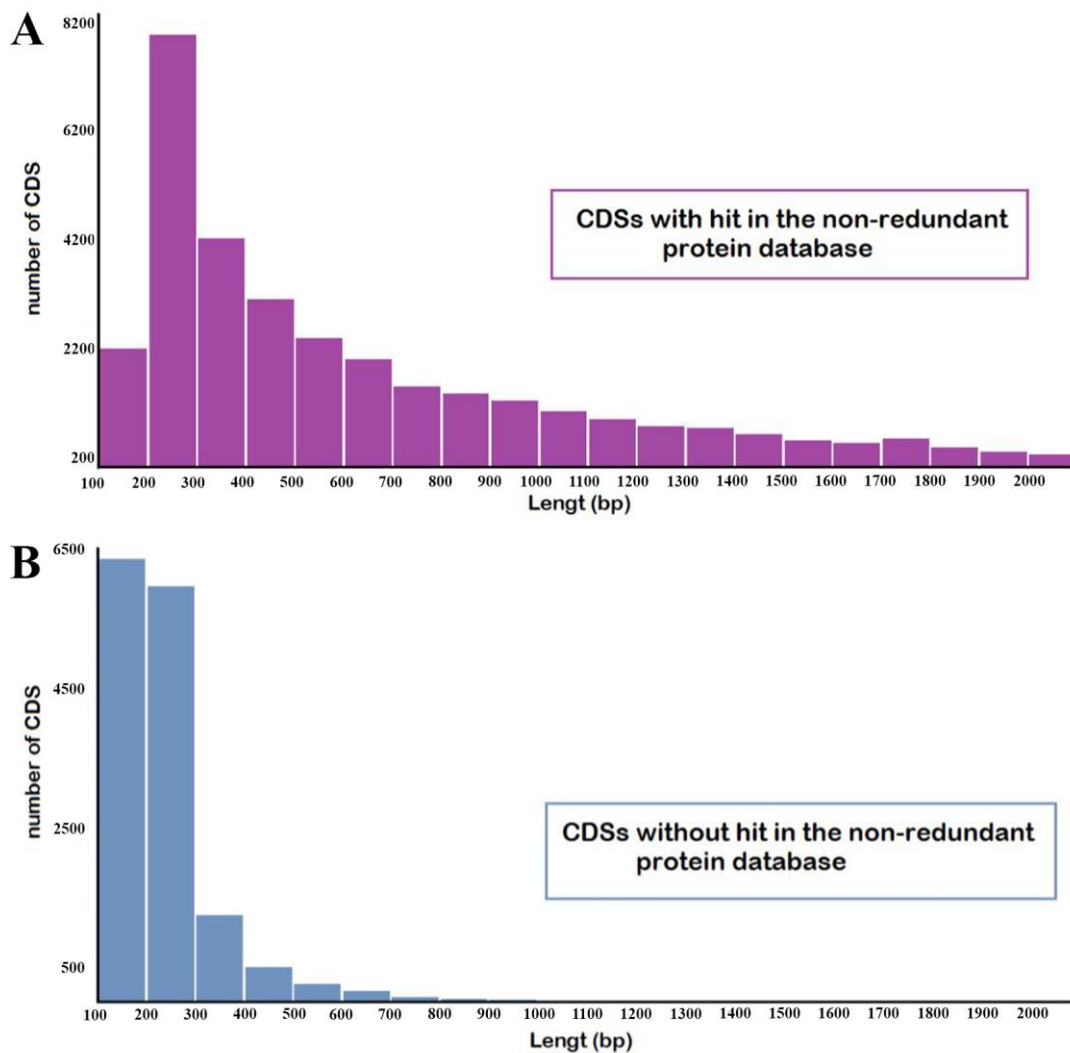


Figure 15. Distribution of number of CDSs with hits in nr database (A) and CDSs without hits in nr database

The analysis of final assembly showed that 1.94 sequences share the same hit against the National Center for Biotechnology Information (NCBI) nr database (Figure 16A), due to sequencing faults and/or possible isoforms of protein. After

comparison with non-redundant data base of the NCBI we obtained the following results: 63.3% of transcriptome, which is 31,463 transcripts, was assigned to at least one protein, while 33.7% (15,988 transcripts) was with no significant hits. The potential reason could be the lack of data for mollusc species.

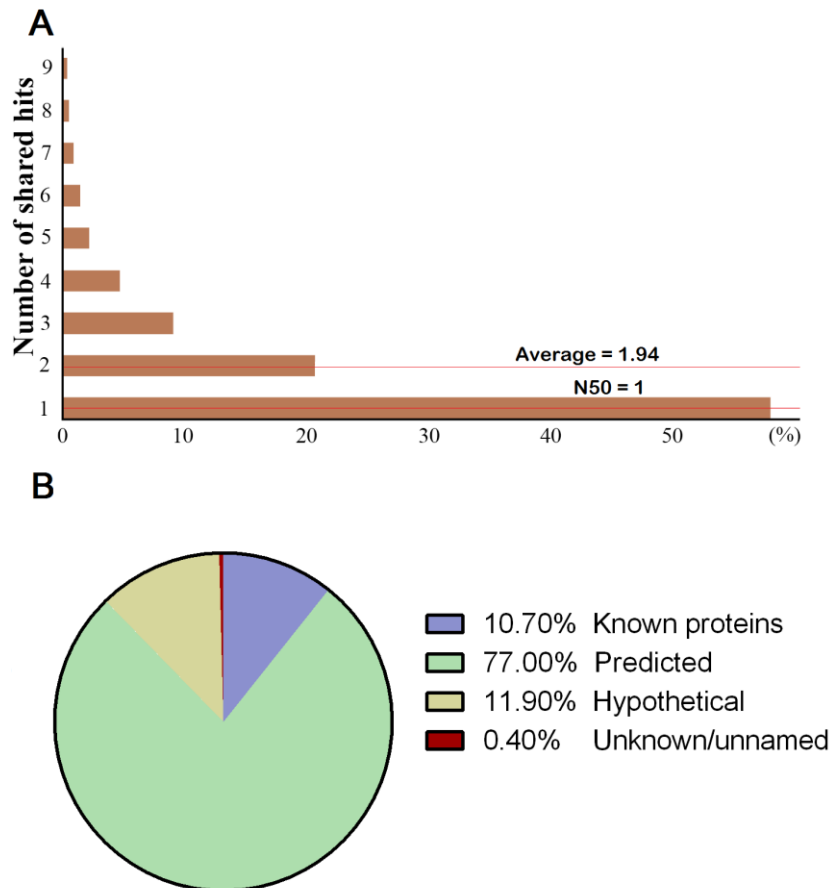


Figure 16. Specification of final assembly of *A. vulgaris* transcripts in comparison with the non-redundant NCBI database (A) and distribution of proteins in comparison with nr NCBI database (B)

Among 31,463 matches, 10.7% (3,380 transcripts) are known proteins, 77% (24,217 transcripts) are predicted, and 11.9% (3,739 transcripts) are hypothetical. Finally, 0.4% (127 transcripts) represents unknown or unnamed proteins (Figure 16B).

From the data that we acquired, one part of sequences, without similarity in the nr database (22,055 transcripts < 300 bp - 46.5% of final assembly), may be

referred as novel genes, ncRNAs or small open reading frames. Kaessmann suggests that novel genes might be a part of a non-coding genome and/or might be fragments of taxonomically ancestral constricted genes (168). Cech and Steitz show that ncRNAs and small open reading frames could be a reason of sequences lacking detectable homology with protein sequences in nr databases (169).

Data mining in genomics is based on functional annotation of individual sequences with potential biological function. According to BLAST2GO tool of FastAnnotator, which is one of the automatic annotation software (170), 39% of transcriptome (18,561 sequences) were functionally annotated with Gene Ontology (GO). The results were grouped into categories “biological process” (33.9%), “molecular function” (28.2%), and “cellular component” (34.8%). The processes with the largest number of sequences in biological process group are: cellular, metabolic, biological regulation, and a response to stimulus. Full list of annotated “biological process” is given in Figure 17.

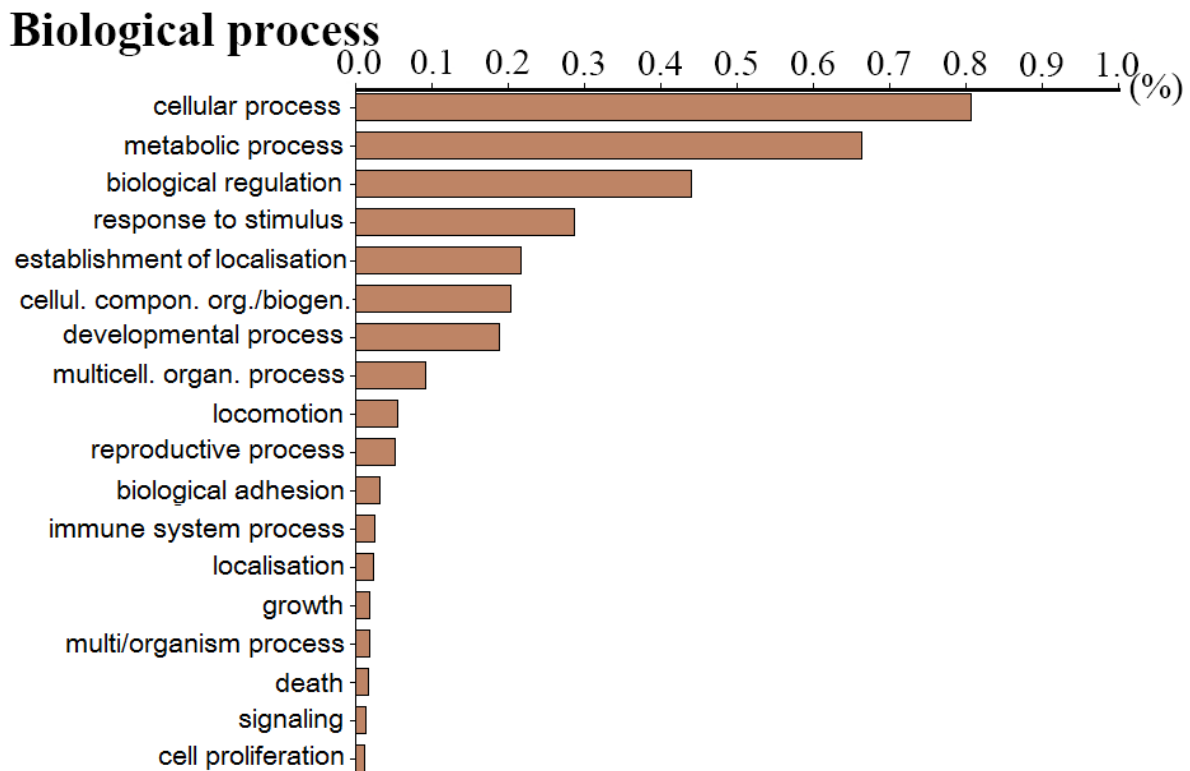


Figure 17. List of annotated biological function according to Gene Ontology (GO) analyses of the *A. vulgaris* transcriptome

The categories which are annotated as “molecular function” and “cellular component” are listed in Figure 18. The highest percentage of GO terms in the „molecular function“ group corresponds to binding, catalytic or transporter activity, while in the „cellular component“ group these are organelle/organelle part, macromolecular complexes or extracellular regions.

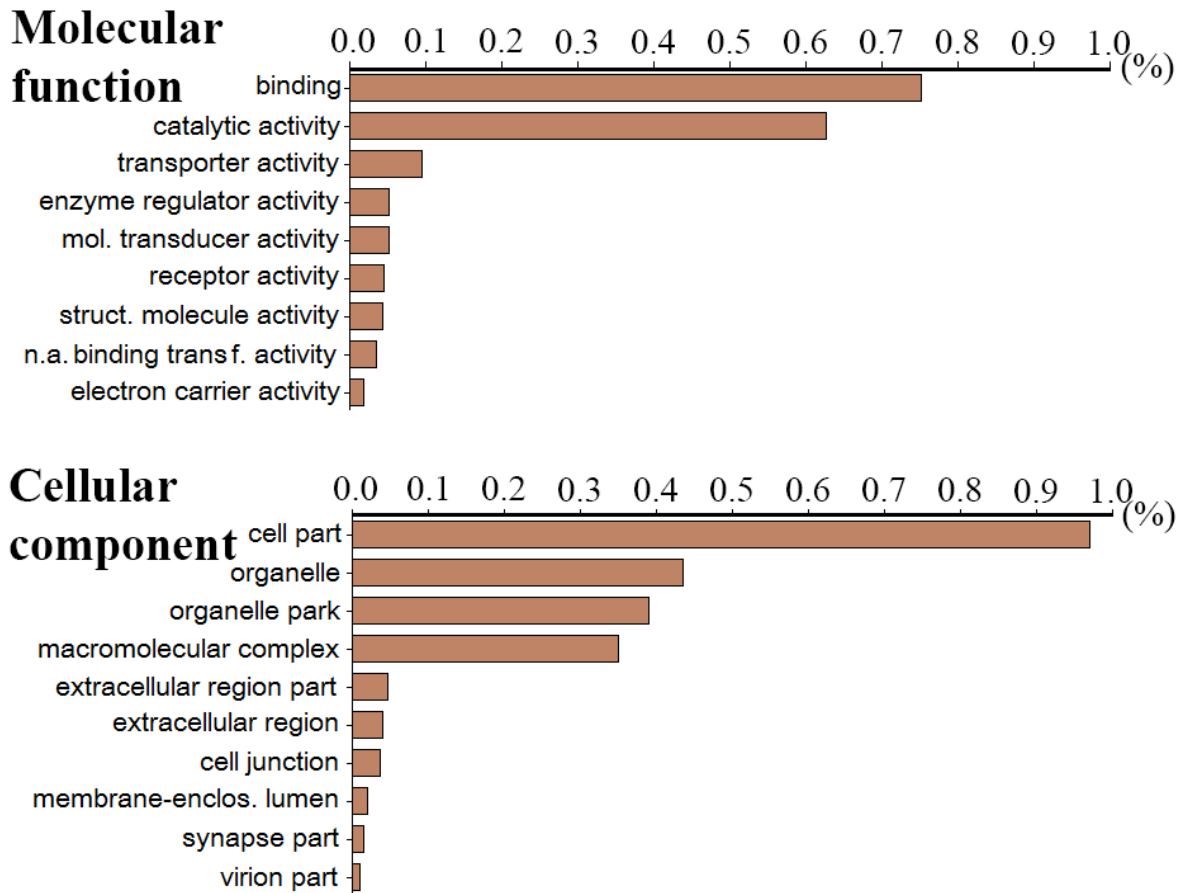


Figure 18. List of annotated molecular function and cellular component according to Gene Ontology (GO) analyses of the *A. vulgaris* transcriptome

A protein domain represents a functional part of a protein, and diverse types of domains can be found in nature. Identification of protein domains can contribute to the understanding of protein function. Many sequence analysis and protein folding algorithms used in computational biology have been built on hidden Markov models (HMMs). This powerful model is now used worldwide (e.g. Human genome project), since laboratories are not able to process the amount of new information so

fast. The Pfam database, created by European Bioinformatics institute contains a large collection of protein families, as well as higher-level groupings of related entries, known as clans. After we ran the transcriptomics data against Pfam database, in 16,994 of entries at least one domain was recognized, and of those entries, 471 sequences was without blastX hit annotation. BlastX is a tool which has a function to compare a nucleotide query sequence translated in all reading frames against a protein sequence database. Three most abundant domains are zinc-finger double domain (zf-H2C2_2), domain of unknown function (DUF4200) and ankyrin repeat (Ank_5). Full list is available as a supplementary material S1 table published in paper by Bulat T. et al. (167), under Pfam numbers 13465, 13863 and 13857. According to output from GO, 1,796 of transcripts were assigned with EC (Enzyme Commission) numbers which specify enzyme-catalysed reactions.

Zinc-finger proteins represent a very structurally diverse group of small proteins with a broad range of functions, such as replication, repair, signalling, cell proliferation and apoptosis. Zinc ion is crucial for the stability of this domain (171). These proteins are usually interaction modules and have a role in binding small molecules, proteins and amino acids. Zinc-finger proteins are divided into few groups according to the type of the domain: Cys2His2, gag-knuckle, treble-clef, zinc ribbon, Zn2/Cys6 and Miscellaneous. Zinc-finger double domain (zf-H2C2_2) belongs to Cys2His2 group and contains an α -helix and an antiparallel β -sheet. Position of zinc ion is organized by two histidine residues and two cysteine residues (Figure 19). These amino acid residues can be present in a protein in various numbers (two, four, six times, etc.).

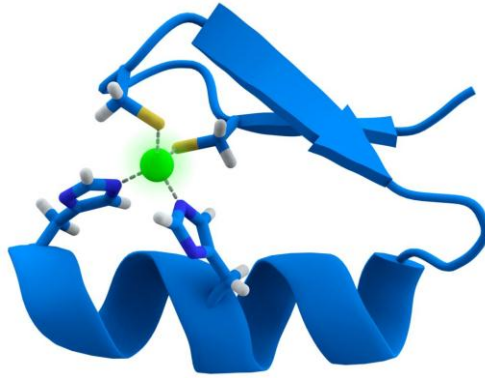


Figure 19. Illustration of zinc-finger domain structure where zinc-ion is stabilize by two histidine residues (dark blue) and two cysteine residues (yellow)

Unlike DUF4200, whose function is unknown, ankyrin repeat domain is found in proteins which have been connected with, among others, transcriptional initiators, ion transporters and signal transducers. Ankyrin domains are among the most common structural motifs within known proteins (172).

Finally, we compared our slug database in blastX in order to check best-match species. Results are presented in Figure 20. 65% of sequences is shared with *Aplysia californica*, 8.7% with *Crassostea gigas* and 2.5% with *Capitella teleta*. More than 80% of the sequences best matched with molluscs species. Some of the transcripts were shared with plants, fungi, bacteria and viruses. These results may also be due to the contamination during sample preparation. Also, it should be taken into consideration that more functional data on these species should be obtained.

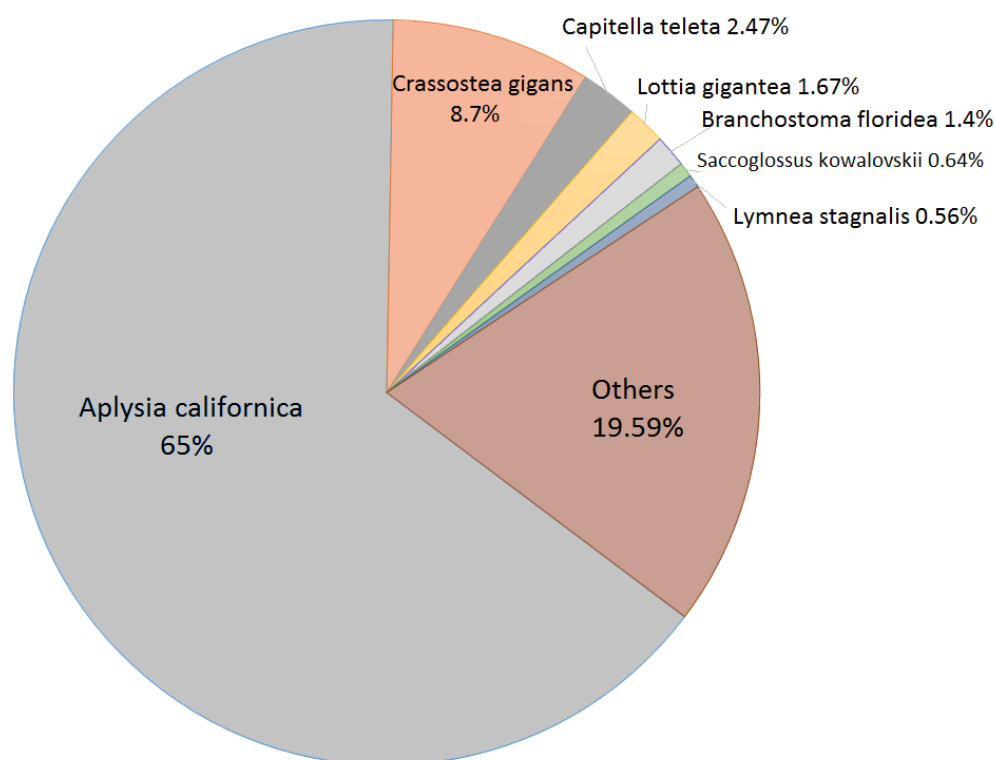


Figure 20. Results of blastX hits against the nr database in comparison with other species

4.2. *Arion vulgaris* mantle proteome

Gel-based proteomics analysis was performed from *A. vulgaris* mantle. Samples were run on 2-DE and 1-DE gel. From 2-DE gel (Figure 21) 356 spots were clearly resolved and trypsin digested. Most of the protein spots in the whole broad range of molecular weight (10-250 kD) are also in the range from pH 5 to pH 9. After running tryptic digested peptides and performing a search against transcriptome-based database, we identified 833 unique proteins, from which 814 had a significant hit in NCBI nr database. Besides this, 19 proteins are with no significant homology in nr NCBI database (Table 3). Some of the novel proteins are detected in unexpected different positions in gel due to possible isoforms and/or posttranslational modifications. The mass spectrometry proteomics data have been deposited to the ProteomeXchange Consortium (173) via the PRIDE partner repository with the dataset identifier PXD002078 and 10.6019/PXD002078. Full list of identified proteins is given in Appendix (Table 1).

Due to the limitation of 2-DE based proteomics, we also ran samples on 1-DE gel, where they were digested and analysed on LC/MS-MS with higher resolution, in order to compensate low separation by 1D-SDS-PAGE. The full list of proteins with hits in nr NCBI database is given in Appendix (Table 2). We identified 2011 proteins from the two biological replicates, from which 48 proteins are without a hit on nr NCBI database (Table 4). Total number of identified proteins from 1-D and 2-D gels, with minimum of one unique peptide and distinct peptides, is 2129. Using a target-decoy search strategy, we estimated a false-positive assignment around 1%. Since the mantel is composed of muscle and skin, a high number of peptides were assigned to paramyosin, myosin, actin and arginine kinase, as well as to others cytoskeletal and muscle-related proteins. Arginine kinase has a highly significant role in invertebrates, since its role is in maintenance of ATP levels, which is achieved by the phosphorylation of phosphagens (174).

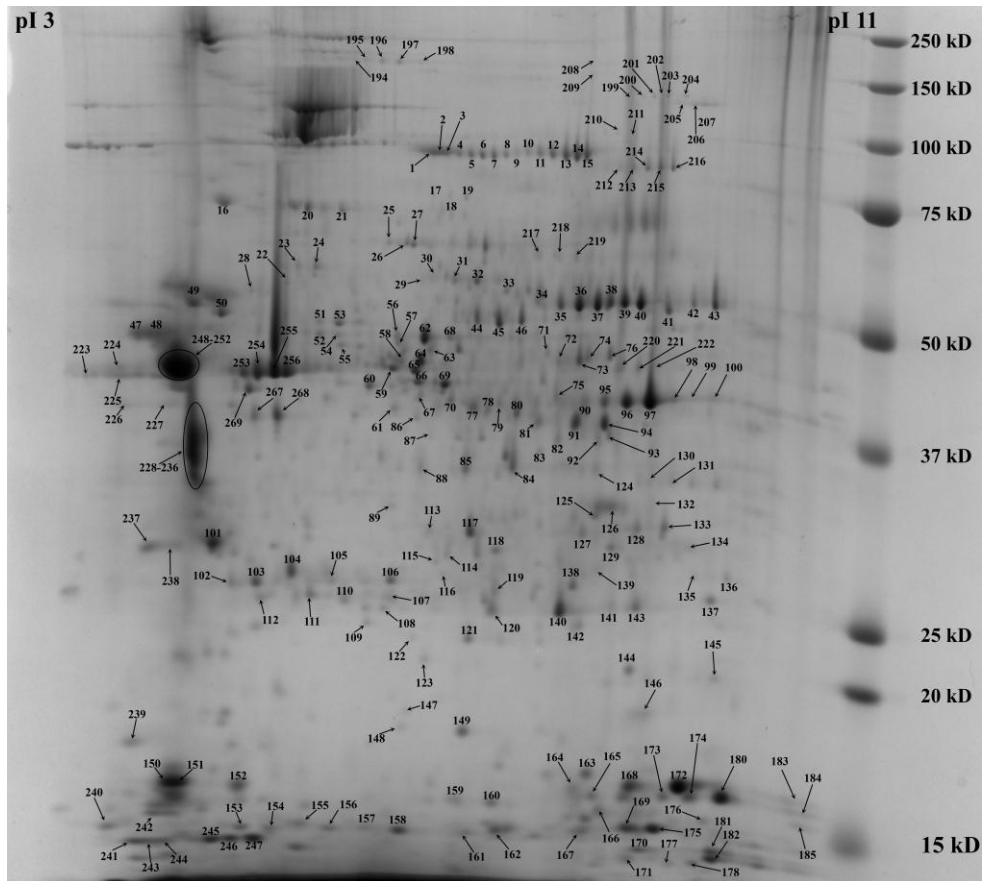


Figure 21. Representative 2-DE gel from *A. vulgaris* mantel proteome study. Numbers represent spots selected for MS analysis

Genus *Arion* belongs to the phylum Mollusca, and it can be considered as the most successful in terms of microbacterial resistance, taking into consideration the fact that molluscs lack adaptive immunity. During their life, they are in contact with many pathogens, including bacteria, fungi, viruses and several lineages of specialized eukaryotic parasites. Body of slugs is protected by mucus-producing epithelium that represents the first physical barrier which defends the organism against pathogens.

Table 3. Detailed information on 19 proteins identified from 2-DE proteomics study without significant hit on NCBI non-redundant database database (acc_embl- EMBL accession number; prot_acc- UniProt accession number; uni_pep- number of unique peptides; prot_cov- protein coverage (%); num_mat- number of assigned peptide matches; num_seq- number of protein sequences; prot_score- MASCOT score)

acc_embl	prot_acc	uni_pep	prot_cov(%)	num_mat	num_seq	prot_score	spot number
HACG01047575	A0A0B7BQP0	26	43	26	122	1681	34,118,180,181,182,183,116,178,251,252,268,49,86,89,114,102,67,298
HACG01032016	A0A0B7ACZ5	9	51	9	83	1050	240,241,243,244,245,334,338,340
HACG01029986	A0A0B7AA27	5	19	5	29	484	162,228,230,231,232,233,235,236,229,234,249,250
HACG01019436	A0A0B6ZEL1	5	46	10	53	1374	56,57,63,62
HACG01023202	A0A0B6ZNW9	4	18	4	32	300	16,49,18,105,109,110,111,194,196,19,7,200,207,211,212,213,98,100,101,103,282,289
HACG01026720	A0A0B7A0U9	4	28	4	56	1389	240,241,242,243,244,245,246
HACG01044712	A0A0B7BE64	4	21	4	7	89	237,238
HACG01005312	A0A0B6Y748	3	20	3	7	95	156,157
HACG01033711	A0A0B7AKN6	3	39	3	10	116	207,213,214,215
HACG01039132	A0A0B7AZH5	3	17	3	20	276	160,163,164,168,183,169,173,174,176,181
HACG01044724	A0A0B7BE79	3	30	3	11	167	231,235,236,232,233,261
HACG01005243	A0A0B6Y7L6	2	7	2	2	53	67
HACG01013119	A0A0B6YUN6	2	8	2	6	130	116,119,139,142
HACG01028842	A0A0B7A4Y5	2	15	2	6	181	244,245,246
HACG01032167	A0A0B7AEI5	2	3	2	2	45	194
HACG01036973	A0A0B7AVD5	2	20	2	3	59	240
HACG01043414	A0A0B7BA86	2	11	2	4	57	156,158,348
HACG01044713	A0A0B7BF10	2	10	2	2	30	102
HACG01019435	A0A0B6ZEY8	1	23	4	5	72	62

Table 4. Detailed information of proteins identified from 1-DE proteomics study without significant hit on NCBI non-redundant database (acc_embl- EMBL accession number; prot_acc- UniProt accession number; uni_pep- number of unique peptides; prot_cov- protein coverage (%); num_mat- number of assigned peptide matches; num_seq- number of protein sequences; prot_sc- MASCOT score)

acc_embl	prot_acc	uni_pep	prot_cov	num_mat	num_seq	prot_sc
HACG01047575	A0A0B7BQP0	21	73	21	218	4285
HACG01026043	A0A0B6ZWD9	7	17	7	30	766
HACG01026345	A0A0B6ZZP9	7	30	7	30	365
HACG01032167	A0A0B7AEI5	7	20	7	7	96
HACG01044712	A0A0B7BE64	7	40	7	35	727
HACG01044713	A0A0B7BF10	7	30	7	25	443
HACG01026720	A0A0B7A0U9	6	32	6	46	540
HACG01013119	A0A0B6YUN6	5	26	5	15	124
HACG01033711	A0A0B7AKN6	5	62	5	14	282
HACG01036259	A0A0B7APZ4	5	22	5	6	146
HACG01005318	A0A0B6Y7S7	4	16	4	4	137
HACG01023202	A0A0B6ZNW9	4	21	4	32	373
HACG01026996	A0A0B7A183	4	21	4	8	92
HACG01028842	A0A0B7A4Y5	4	29	4	19	447
HACG01033499	A0A0B7AKC1	4	8	4	10	164
HACG01036974	A0A0B7AS42	4	34	4	7	82
HACG01039132	A0A0B7AZH5	4	36	4	17	752
HACG01021092	A0A0B6ZHY7	3	17	3	3	44
HACG01023330	A0A0B6ZR28	3	25	3	4	77
HACG01028841	A0A0B7A6R5	3	32	3	7	116
HACG01030092	A0A0B7A8J9	3	30	3	13	381
HACG01030913	A0A0B7AAF1	3	33	3	6	125
HACG01044700	A0A0B7BHE4	3	19	3	6	117
HACG01044706	A0A0B7BGZ4	3	22	3	8	220
HACG01044707	A0A0B7BFB4	3	29	3	10	274
HACG01019435	A0A0B6ZEY8	3	44	14	37	616
HACG01019436	A0A0B6ZEL1	3	55	15	44	760
HACG01002111	A0A0B6XY56	2	41	2	6	109
HACG01005312	A0A0B6Y748	2	14	2	2	36
HACG01026042	A0A0B6ZX56	2	5	2	4	97
HACG01027525	A0A0B7A2T2	2	50	2	3	48
HACG01030091	A0A0B7AAE4	2	31	2	15	358
HACG01032016	A0A0B7ACZ5	2	15	2	7	100
HACG01033415	A0A0B7AK29	2	30	2	4	39
HACG01035358	A0A0B7AN13	2	12	2	2	75

acc_embl	prot_acc	uni_pep	prot_cov	num_ma t	num_seq	prot_sc
HACG01036973	A0A0B7AVD5	2	25	2	4	116
HACG01044893	A0A0B7BFI7	2	2	2	3	32
HACG01047662	A0A0B7BNQ7	2	8	2	2	44
HACG01052636	A0A0B7C2V6	2	33	2	2	37
HACG01044718	A0A0B7BF15	2	37	3	12	207
HACG01044724	A0A0B7BE79	2	50	3	18	271
HACG01044715	A0A0B7BHF6	2	32	5	17	364
HACG01042313	A0A0B7B827	1	8	2	2	33
HACG01044714	A0A0B7BE68	1	12	2	3	88

The modification of immune systems during evolution involves the expansion of particular gene families in given phyla. Interestingly, during the analysis of data that we obtained from a comprehensive transcriptomics and proteomics study, we showed that genes/proteins which are related to immunity are expressed. We identified wide-range of proteins which indicates an unexpected complexity of the immune system with a high grade of pathogen specificity and immune-priming. Some of them may play a role in pattern recognition, as effector proteins or cytokine-like proteins, such as lectins. In addition, we have also detected complement-like proteins, peptidoglycan-recognition proteins (PGRPs), lipopolysaccharide and β 1, 3-glucan-binding proteins, fibrinogen-related proteins (FREPs), pore-forming membrane attack/perforin (MACPF) domain proteins and toxin-like proteins.

Lectins are very important in „self-non-self” recognition. Lectins that we detected (C-type lectin family, galectin, malectin, H-type lectins, L-type lectins, Ricin-B lectins and calnexin) contain conserved carbohydrate-recognition domains (CRDs). In total, lectin-associated functions may be attributed to 149 transcripts, 37 of which were unambiguously identified at the protein level. In Table 5 all lectin-like proteins which we detected on transcriptomics and proteomics level are presented, while all detected lectins at transcriptome level are given in Appendix (Table 3.).

C-type lectins are able to bind carbohydrate in a Ca^{2+} -dependent manner, thanks to CRDs which contain two disulphide bridges located in the base of two loops (175). 47 transcripts encoding proteins with C-type lectin CRDs were found and 14 were also identified at the protein level. 17 of 21 complete transcripts have a predicted signal

peptide demonstrating that they are secreted by cells to utilize their functions (Table 5). In Figure 22 we showed an amazing sequence variability within incilarin-like proteins identified in our study, while in Figure 23 we illustrated a domain structure of C-lectin-like molecules found in the transcriptome of *A. vulgaris*. Incilarins belong to C-type lectins. They were isolated from the surface of the body (mucus) of the land slug, *Incilaria fruhstorfer*. This protein was found in the water-soluble fraction and it is known that it possess hemagglutination activity (176).

Table 5. The list of lectin like proteins detected at the transcriptome (T) and protein level (P) with status- complete or incomplete as the entire CDS from the initial ATG to the STOP codon was represented.

prot_acc UniProt	Protein hit description in nr NCBI database	Status	Evidence	Type
A0A0B7AZQ2	PREDICTED: calnexin-like isoform X1 [Aplysia californica]	complete	T,P	calnexin
A0A0B7AWH4	PREDICTED: calnexin-like isoform X2 [Aplysia californica]	complete	T,P	calnexin
A0A0B7A018	incilarin A [Haliotis discus discus]	complete	T,P	C-type lectin
A0A0B6ZB52	Incilarin A [Meghimatium fruhstorferi]	complete	T,P	C-type lectin
A0A0B6ZSS2	Incilarin A [Meghimatium fruhstorferi]	complete	T,P	C-type lectin
A0A0B6ZT36	Incilarin A [Meghimatium fruhstorferi]	complete	T,P	C-type lectin
A0A0B7AGC8	Incilarin A [Meghimatium fruhstorferi]	complete	T,P	C-type lectin
A0A0B6Y7S8	Incilarin A [Meghimatium fruhstorferi]	complete	T,P	C-type lectin
A0A0B6ZSZ0	Aggrecan core protein [Crassostrea gigas]	complete	T,P	C-type lectin
A0A0B6ZQL8	PREDICTED: cell wall protein DAN4-like [Aplysia californica]	complete	T,P	C-type lectin
A0A0B7B8E9	Incilarin A [Meghimatium fruhstorferi]	complete	T,P	C-type lectin
A0A0B7BB09	putative perlucin 4 [Haliotis discus discus]	incomplete	T,P	C-type lectin
A0A0B6ZZL1	Incilarin C [Meghimatium fruhstorferi]	incomplete	T,P	C-type lectin
A0A0B7BCB3	Low affinity immunoglobulin epsilon Fc receptor [Crassostrea gigas]	incomplete	T,P	C-type lectin
A0A0B6Z234	PREDICTED: uncharacterized protein LOC101860995 isoform X1 [Aplysia californica]	incomplete	T,P	C-type lectin
A0A0B7BV24	PREDICTED: uncharacterized protein LOC101848467 [Aplysia californica]	incomplete	T,P	C-type lectin
A0A0B7AWV4	tandem repeat galectin [Biomphalaria glabrata]	complete	T,P	galectin
A0A0B7B7G5	CHAI like protein precursor [Cepaea hortensis]	complete	T,P	H-type lectin
A0A0B6Z3C5	agglutinin [Helix pomatia]	complete	T,P	H-type lectin
A0A0B6Y9P9	agglutinin [Helix pomatia]	complete	T,P	H-type lectin
A0A0B7AA27		complete	T,P	H-type lectin

A0A0B7BA86		complete	T,P	H-type lectin
A0A0B7AEX4	Chain A, Structure Of Helix Pomatia Agglutinin With No Ligands	incomplete	T,P	H-type lectin
A0A0B7ACW2	Chain A, Structure Of Helix Pomatia Agglutinin With No Ligands	incomplete	T,P	H-type lectin
A0A0B7ANH6	PREDICTED: vesicular integral-membrane protein VIP36-like [Aplysia californica]	complete	T,P	L-type lectin
A0A0B7ADM8	PREDICTED: malectin-B-like [Aplysia californica]	incomplete	T,P	malectin
A0A0B6XX19	PREDICTED: malectin-B-like [Aplysia californica]	incomplete	T,P	malectin
A0A0B6ZWR8	PREDICTED: uncharacterized protein LOC101856230 [Aplysia californica]	complete	T,P	C1qDC
A0A0B7A1M5	sialic acid binding lectin [Cepaea hortensis]	complete	T,P	C1qDC
A0A0B7A6S4	sialic acid binding lectin [Cepaea hortensis]	complete	T,P	C1qDC
A0A0B6ZJ78	sialic acid binding lectin [Helix pomatia]	complete	T,P	C1qDC
A0A0B7ABV6	putative C1q domain containing protein MgC1q41 [Mytilus galloprovincialis]	incomplete	T, P	C1qDC
A0A0B6ZN56	sialic acid binding lectin [Haliotis discus discus]	incomplete	T,P	C1qDC
A0A0B7ALE7	sialic acid binding lectin [Haliotis discus discus]	incomplete	T,P	C1qDC
A0A0B7A907	sialic acid binding lectin [Cepaea hortensis]	incomplete	T,P	C1qDC
A0A0B6ZHG1	sialic acid binding lectin [Helix pomatia]	incomplete	T,P	C1qDC
A0A0B7AJC2	PREDICTED: uncharacterized protein LOC101850618 [Aplysia californica]	complete	T,P	C1qDC

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HACG01005349 -MREIVLLLALSAHCVLSHHDVCIYYGWPVYDGYCYGFLEEVNWLAAASCSQFGERLAEVNNEERDKWLEQAEHRKLGPTWIGGSRARLHPGLFWVPSHR--AVEAYTNIQGVPI--NAGHCLEIYGDKKG---WNRDCSLPRTFCIKVVDV
HACG01018944 -MILAVLVLSLAVTCVTS--LSCQKGLQYQENCYGFVPEPWNLLAGASCRQQTARLLEIEDAGENQFIVSQQLKRFKFDSTWVGGSARLHIGTVEWVETLA--RVGKYTNWQGEFN--NYNGKEHCLELSKVVNRYRWFNFCLENPPFVCKRKAQVI
HACG01023930 -KMLVLLAMALAPPFVSS--HKCYSGWEQKNSCYGLGEEHVDQDAAQFCELNLYLVEINDEENNWLVTYKKGKASGVVWIGGSEHHRGVVEWIPSR--PFGKFSWYGNPE---LVAEDCLEISAGLYKWNENCLAAARFVCEASLV--
HACG01024954 -MIRVLLLALSQCVELS--DDECYNGWAANEGYCYGFVEDVSWLLASANDLFGARLAEIDSQRETWLSQQLVVKLPVAVWGLSSRLHPGVWQWTPSNR--DVSRYVNWIPGEPN--NYENNEQCVELKSTG---WNDLNCLTAKKFCCKLIT1
HACG01027153 -KMLRELLFTLSAQYAAS-EIVCPLGWLIFQNHCFYAFIDRLNYYAAAAACDYLDLYLVKIEAENTWLVSVLRSYNFQETWIGGSQKHLALQFVWVPSQE---PFTLGNWGGGQP--DNTRGRESYLEIRGEFNFSWHDHLEIEISFVCEKPEPHKI
HACG01032978 -MIGVLLALTYQCWVG--AATPSGKVLNHSYGFVIGDQKTWDAQKSCRSYGAELVEINNAEEDQFVATLALKQGNWVWGGASLPGKKEWVWVSHHTVDCQRYVWNAEFPN--NAGSEWCLELRDIG---WHDHCNTRNSFVCEK-----
HACG01041705 -MIRFVLLAVTATCAYG---AGLESWTEFNQYCFSPQKEPTVSAVAAYVCHSISGLRVEIDSAAEDNDWLGELNASHSGSIFAGLNRRS-SPEWVWDPKSR--TADKYSVWNVGEPN-TELSFECLAHYGVARK--WTAVSCSAPPPTCEKKS1KI
HACG01041706 -MIRFVLLAVTITCALG---GCPAGWREPKGYCYSTNREPLWASAYAFONSIGARLAEIDGAERDQWLAQLDQNFQSTFVGLTRRLRLVWVWVPSLA--NTAKYSVWNVGEPNPNYGLNEECLEKFSARKK--WNDYCGESAPPTCEKRINI1
HACG01041708 -MIRFVLLAVTITCAFG---DQLEGWRELNEHCYSFHKEAVTWVAFSYCESIRARLVEIHSEEVKWLWVQLDELGFEKTEAGMSRRRHAPNWLWETSQ---ICPTYTNWIDGEPN-GDASEECLEINVDASKK--WNDVSOVENPTFCEKSAKGI
Incilarin A -MIRVLLLVLAACVFC---ACPNGWKEPKGYCYFYPERVNWLVASASCNLYGGRLEPDISEQDQWLAELTALKFGETWAGGSARLHVWKEWVPSLA--DFSRHSHWVWAGEPN--NVANNVEYCLEINQSPKGNKATTEERQFICERRVDA-

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Figure 22. Alignment of amino acid sequences of incilarin-like proteins from *A. vulgaris*

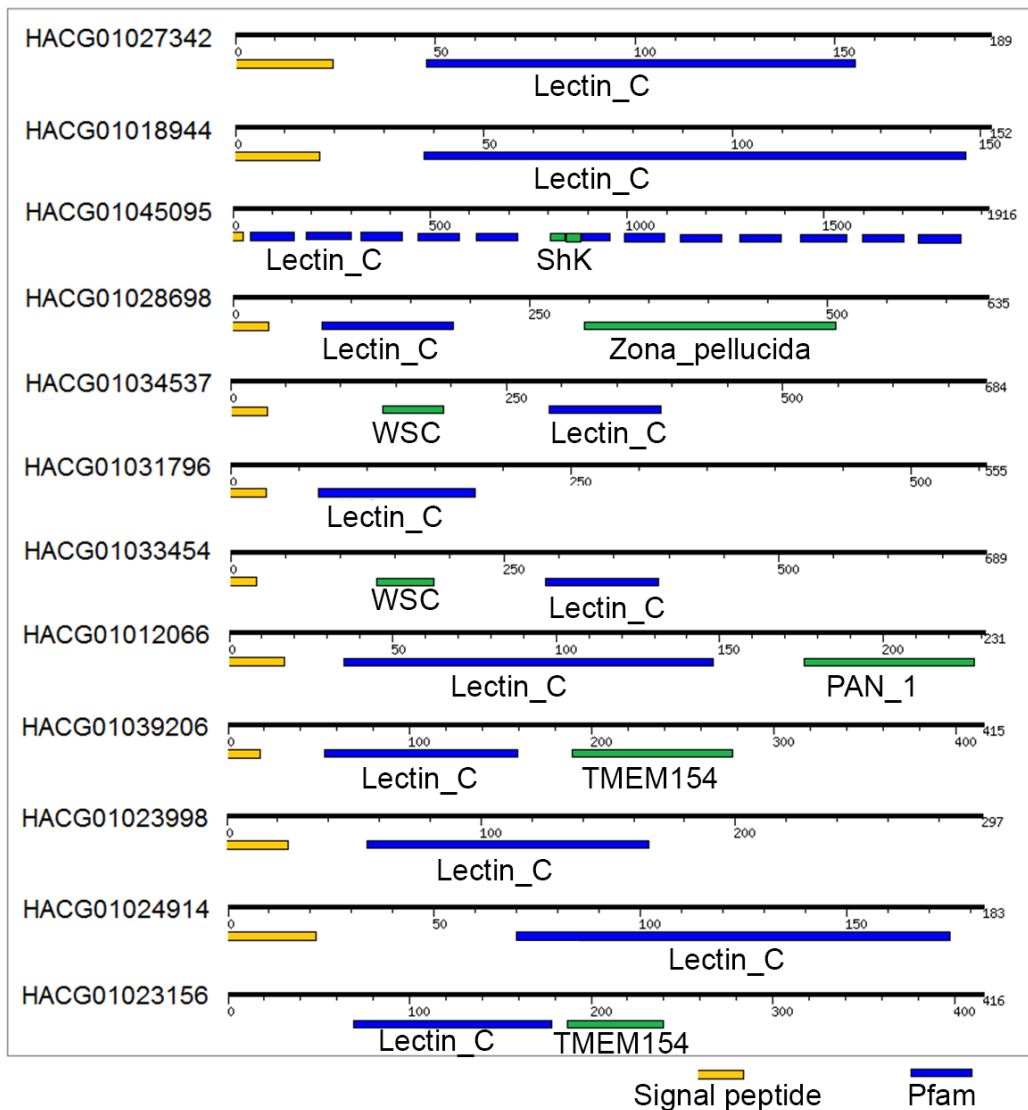


Figure 23. Domain architecture of Lectin_C-like proteins from *A. vulgaris*

Next to Lectin_C, some of the identified peptides contain domains recognized as ShK, WSC, Zona pellucida (ZP), PAN_1 and TMEM154. ShK is a 35 amino acid domain, cross-linked by three disulphide bridges between Cys3-Cys35, Cys12-Cys28, and Cys17-Cys32, which was first identified in a sea anemone *Stichodactyla helianthus* (177). Several independent studies show that the ShK domain blocks the voltage-gated potassium channels - Kv1.1, Kv1.3, Kv1.6, Kv3.2 and KCa3.1 (178-180). WSC (Wall Stress-responsive Component) domain is a cysteine-rich domain, structured by approximately 90 amino acids, and it represents a carbohydrate binding domain. It has been identified in fungi (*Saccharomyces cerevisiae*, *Aspergillus fumigatus*, *Aspergillus*

nidulans), mammalia and insects (referring to UniProt database). ZP domain consist of ~260 amino acids and it has an essential role in development, hearing, immunity, and cancer. It was distinguished in eukaryotes, in glycoproteins from secreted mucus (mammalian sperm receptors, transforming growth factor (TGF)-beta receptor III, deleted in malignant brain tumour-1 (DMBT-1) (181)). PAN (APPLE) domains also contains a cysteine rich structure with three or four bridges and it is associated with protein-protein interaction, or protein-carbohydrate interactions (182). The function of TMEM154 domain is still unknown. Experiments performed with mutation of TMEM genes in sheep indicate that it can be involved in a lentivirus immune response (183).

Transcriptomics analysis of *A. vulgaris* shows 4 calnexin proteins, from which 2 were confirmed at the protein level with a highest similarity to *Aplysia californica* homologs (blastp: E = 0.0; Identity = 70–74%) (Table 5). It contains a signal sequence and an additional C-terminal transmembrane helix (Figure 24). This protein belongs to an evolutionarily conserved calnexin/calreticulin-superfamily and it is connected to glycoprotein secretory pathways (184). Calnexin is one of the pattern recognition receptors and it has been characterised as a main protein in shrimp antibacterial immunity response (185).

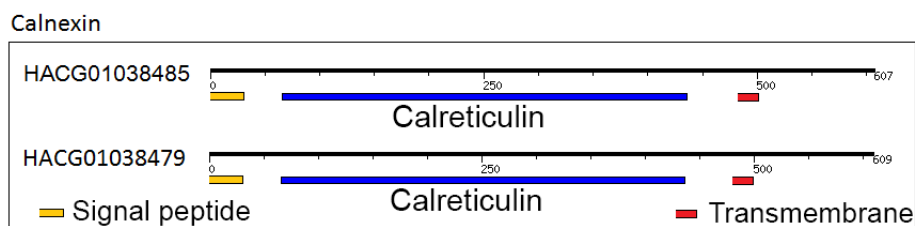


Figure 24. Domain construction of calnexin-like proteins from *A. vulgaris*

The H-type lectins (Figure 25) have a carbohydrate-recognition domain (CRD) structurally similar to *Helix pomatia* agglutinin (HPA) (186). We identified 12 transcripts and from those, 7 were also identified at the protein level (Table 5). These kind of lectins which are detected in *A. vulgaris* share the same size and hexameric arrangement of HPA, thanks to the cysteine residues involved in the intermolecular disulphide bridge construction. HPA function in *H. pomatia* is connected to innate immune system by protecting fertilized eggs from bacteria.

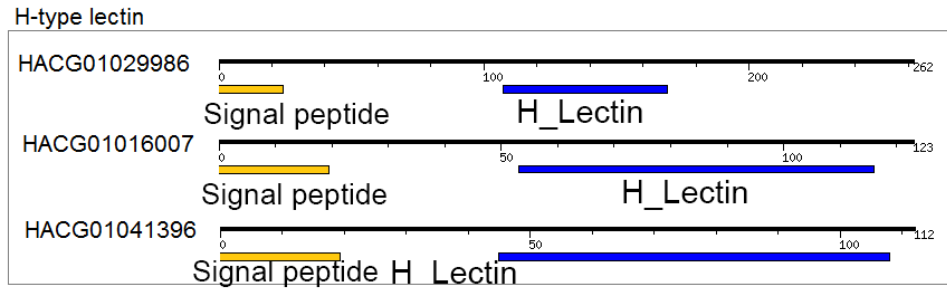


Figure 25. Domain architecture of three H-type lectins of *A. vulgaris*

Additionally, recent studies showed that HPA is able to recognize glycoproteins via O-GlcNAcylation (187). Alignment of amino acid sequences of H-type lectins from *A. vulgaris* with *Helix pomatia* and *Cepaea hortensis* is presented in Appendix (Figure 1).

We reported that ten unigenes coding proteins contain a galectin domain. All of these proteins are the most similar to gastropod galectins (*Aplysia californica*, *Biomphalaria glabrata*, etc. blastp E value ranges from 0 to $2E-32$) (Appendix Table 3). Galectins belong to the family of structurally-related lectins which have a binding specificity towards β -galactoside residues. Proteins which contain a galectin domain are tandem-repeat type galectin (Figure 26). The absence of a signal sequence is consistent with other galectins, which are known to be dynamically secreted from diverse tissue via a “nonclassical” secretory pathway (188).

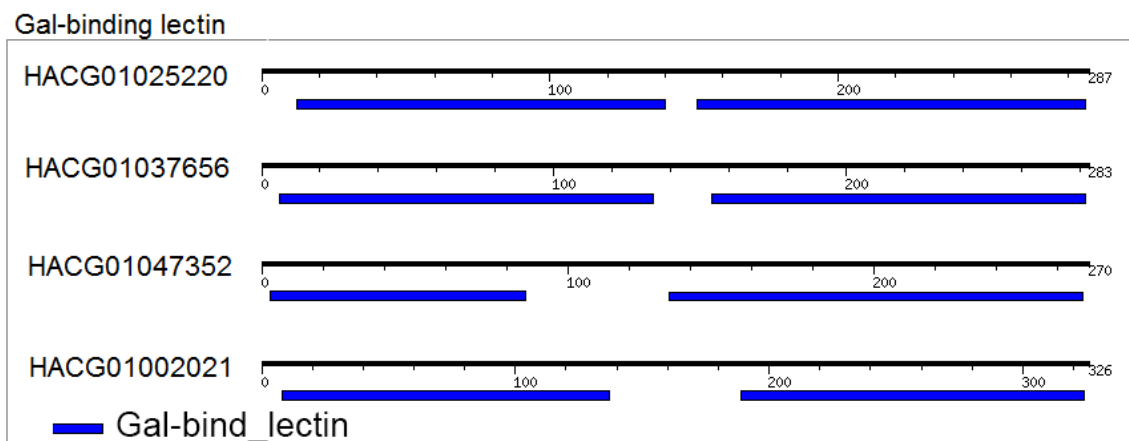


Figure 26. Tandem-repeat structure of galectin like proteins identified in *A. vulgaris*

In total, we detected three L-type lectin sequences. Two of them were at the transcriptome level with the highest similarity to homolog of ERGIC-53 protein from *Littorina littorea* (blastp: E = 0.0; Identity = 57–58%). At the transcriptome and protein level, we detected L-type lectin with VIP36-like homolog from *Aplysia californica*.

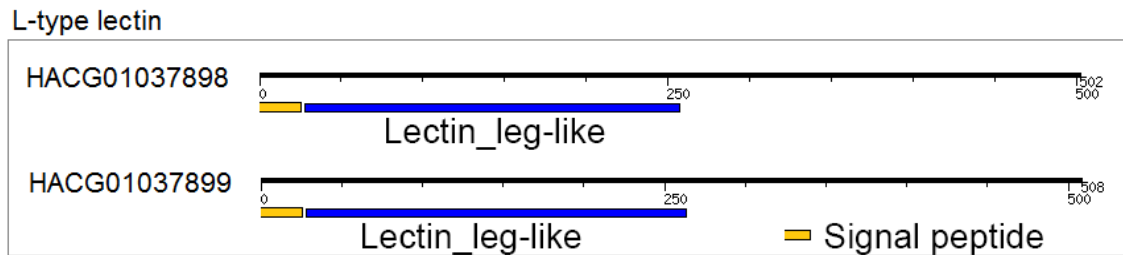


Figure 27. Domain architecture of two L-type lectins of *A. vulgaris*

Both ERGIC-53 and VIP36 are type I transmembrane proteins. ERGIC-53 (ER-Golgi intermediate compartment 53 kDa protein) homologs share the specific architecture with an extracellular signal peptide and a single N-terminal L-type carbohydrate binding site (Figure 27). VIP36 is located in Golgi and post-Golgi portions of the secretory pathway (189).

The R-type lectins belong to a superfamily of proteins which have a CRD structurally similar to the CRD in ricin - toxic, naturally produced lectin in the seeds of *Ricinus communis*. Analysis of the transcriptome predicts 31 unigenes coding proteins of R-type lectins (Table 3. Appendix), which consist of an A chain with enzymatic activity (galactosyltransferase) linked through a disulphide bridge to a B chain with lectin activity (Figure 28). We did not detect them at the protein level probably due to tissue-specificity. Between animal R-type lectins we can distinguish several domains, like mannose receptor (MR) family, EW29, pierisin-1 and pierisin-2. They are identified in the annelid (earthworm) *Lumbricus terrestris* and cabbage butterfly *Pieris rapae*. Pierisin-1 is a 98-kD protein (850 amino acids) that shows extreme toxicity to animal cells in culture (190). Besides this, in *Limulus* horseshoe crab, for example, a coagulation factor G contains two main domains: a central R-type lectin domain and a glucanase-like domain. This protein also has a subunit that is a serine protease, showing that in animals the combination of R-type domain with enzyme is common. The R-type CRD is conserved between animal and bacterial lectins (191).

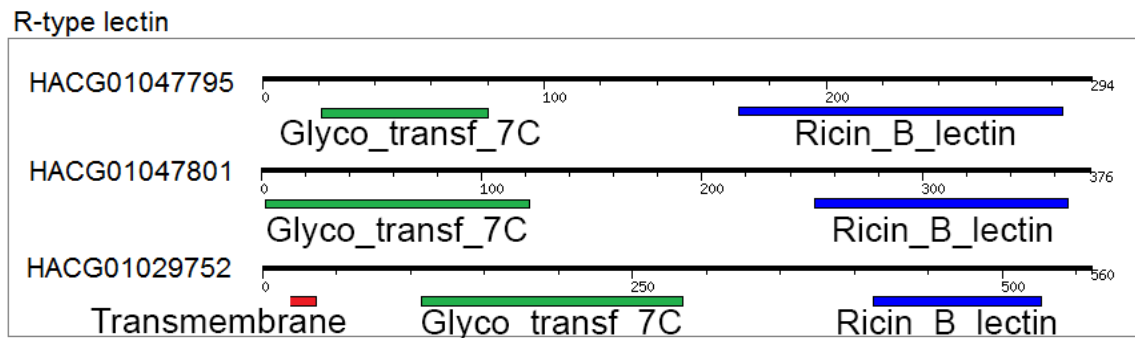


Figure 28. Structure of R-type-like lectin protein from *A. vulgaris*

Malectin is a membrane protein of the ER and it recognises and binds Glc2-N-glycan. At transcriptome and protein level we identified two proteins which have a high similarity with malectin-B-like proteins discovered in *Aplysia californica* (Table 5). The malectin sequence is well conserved in animals (192). The role of malectin in invertebrates is not studied enough, but experimental data from *Xenopus laevis* suggest that malectin could be an important part of the regulatory mechanism in the cleavage process of second glucose from Glc2-N-glycan and in the inhibition of the early entry to the calnexin/calreticulin cycle (CNX/CRT) (193). Taking into consideration that almost all proteins involved in innate and adaptive immune response are glycoproteins (194), and that CNX/CRT cycle ends once glycoproteins reach their native structures, we can postulate that malectin-like proteins detected in *A. vulgaris* can have a significant role in the immune response.

Tachylectins 5A and 5B (TLs-5) are lectins isolated from haemolymph plasma of a horseshoe crab *Tachypleus tridentatus* and they are able to agglutinate human erythrocytes, Gram-positive and Gram-negative bacteria (195). At the transcriptome level we detected three tachylectins like proteins, one TL-5A-like and two TL-5B-like (Table 5). Tachylectin-related proteins have been recognized in numerous organisms (slime molds, sponges, hydroid and bony fish). Their function is correlated with the immune system due to an antibacterial activity: binding to PAMP, phagocytosis of bacteria and binding to lipopolysaccharide (LPS) (196, 197).

Globular C1q domain is the main characteristic of the C1q domain-containing proteins (C1qDC). C1qDC possesses lectin-like properties and has a main role in innate immunity of bivalvia molluscs (198-201). They are involved in several immune responses, such as pathogen recognition (202), microorganism agglutination (203), phagocytosis (204) and mediating cell migration (205). Based on sequence homology and domain similarity, 33 C1qDC sequences from the *A. vulgaris* transcriptome may be classified as members of the C1q family (Table 5 and Appendix Table 3). Ten transcripts have also been identified at the protein level (Table 5). The most of the complete C1qDC sequences revealed a signal peptide or a transmembrane domain on the N-terminus (Figure 29).

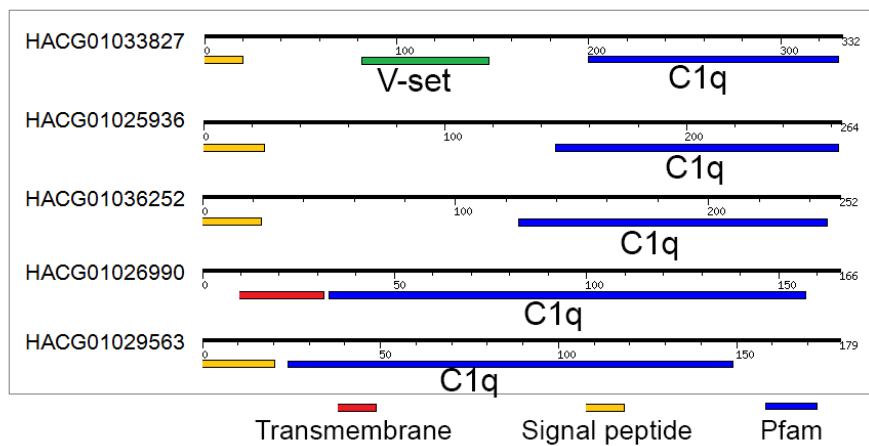


Figure 29. Structure of C1q domain-containing proteins protein from *A. vulgaris*

The C1q domain is involved in the neutralization of viruses, cell adhesion and clearance of apoptotic cells. It is considered as a particularly effective PRD with highly adaptive binding properties, due to the ability of the C1q domain to bind a variety of "self-non-self" ligands, including LPS, virus envelope proteins, phospholipids and some acute-phase proteins (206). Wang et al. suggest that C1qDC proteins could trigger an ancient complement system by the lectin pathway prior to the evolution of immunoglobulins (204).

Besides the C1q domain, C1qDC proteins that we detected, either at protein or transcriptome level, contain homology with V-set and sialic acid-specific binding lectin. V-set are immunoglobulin (Ig)-like domains and can be found in the antibody variable

domain. Sialic acids (Sia) are found mostly at the non-reducing end of oligosaccharide chains on glycoproteins and glycolipids (207), and invertebrate lectins contribute to the innate immune response through binding to Sia on foreign cells (208), like in *Crassostrea virginica* (209), *Tachypleus gigas* (210), *Aphonopelma chalcodes* (211), *Ornithodoros tartakovskyi* and *Ornithodoros tholozani* (212).

During evolution, animals developed different kinds of mechanisms for defence. Some of them have extremely developed escaping skills, while others developed diverse sets of proteins/peptides which they use against predators. Membrane Attack Complex/Perforin (MACPF) consists of proteins with two domains - membrane attack complex (MAC) and perforin (PF), and they belong to pore-forming toxins (PFTs) (213). Recent studies show that MACPF domains are associated with cholesterol-dependent cytotoxicity (CDCs), as well as bacterial PFTs (defence from viruses), which indicates that their mechanisms of action are similar: cholesterol in the target membrane will trigger a pore-forming process, which will lead to cytotoxicity (214). MACPF was discovered as a protecting factor of apple snail eggs against natural enemies (215). In our transcriptomics study of *A. vulgaris*, we found a few candidate effector genes which could be a part of biochemical defence system, including a family of potential pore-forming membrane attack/perforin (MACPF) domain proteins. We assume that together with lectins, they can be a part of an innate immune system. Besides them, in comprehensive transcriptomics data we found mRNA, which according to GO annotation, are coding proteins involved in phagocytosis (71 transcript), coagulation (122 transcript), and Toll-like receptors (50 transcript).

Aside from the importance of the immune system for protection of animals, toxins and toxin-like proteins are also essential. Analysis of *A. vulgaris* shows a novel toxin-like protein with significant similarity to agatoxin. Agatoxin is a toxin from spider glands with a characteristic cysteine motive and it acts as a blocker of glutamate-gated ion channels, voltage-gated sodium channels, or voltage-dependent calcium channels (216). The agatoxin-homolog from *A. vulgaris* shares a specific architecture with a signal peptide, pre-peptide sequence and characteristic cysteine motive in the mature peptide (Figure 30). Proteins with characteristic cysteine pattern are able to build disulphide bonds, and we can distinguish three types of cysteine knots - Growth Factor

Cysteine Knot (GFCK), Inhibitor Cysteine Knot (ICK) and the Cyclic Cysteine Knot (CCK) (Figure 31). Most of ion channel blockers, haemolytic agents, or molecules having antiviral and antibacterial activities, are peptides between 26-48 amino acid residues which contain the cysteine knot (217). The stability of different types of cysteine knots, variety of bioactivities and their exceptional structural scaffold can be used for molecular engineering applications and in drug design (218).

The innate immune system is encoded to distinguish "non-self-materials" mediated by a group of pattern recognition proteins (PRPs) or pattern recognition receptors (PRRs), which can recognize and bind to different pathogen-associated molecular patterns (PAMPs) of invasive microorganisms (219, 220). C-type lectins, PGN-recognition proteins (PGRPs) and galectins belong to PRPs group, indicating that they play a key role in primary defence against invasive microorganisms in *A. vulgaris*. In the last two decades key components of innate immune systems of invertebrates have been discovered established and they include coagulation factors, complement factors, protease inhibitors, antimicrobial peptides, Toll receptors, and other humoral factors. Together they protect the host from being invaded by bacterial, fungal, and viral pathogens (208). A lot of these factors were detected in our study (BioProject accession number PRJEB7891; dataset identifier PXD002078) and together with lectin-like proteins they may protect the host from invading bacterial, fungal, and viral pathogens.

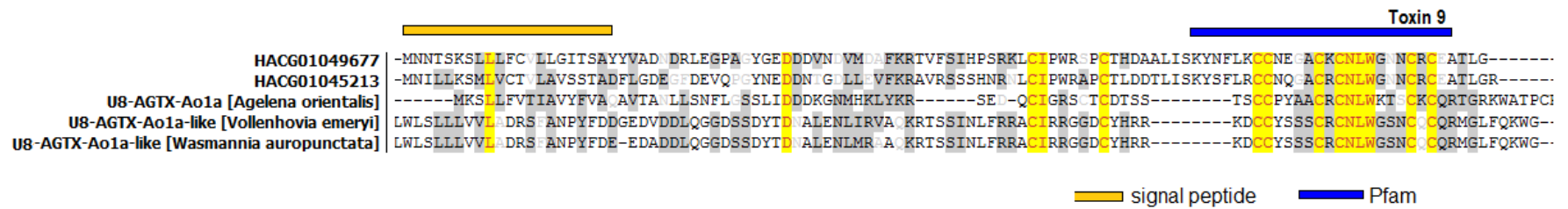


Figure 30. Alignment of amino acid sequences of toxin-like proteins from *A. vulgaris* with U8-agatoxine–Ao1a from *Agelena orientalis*

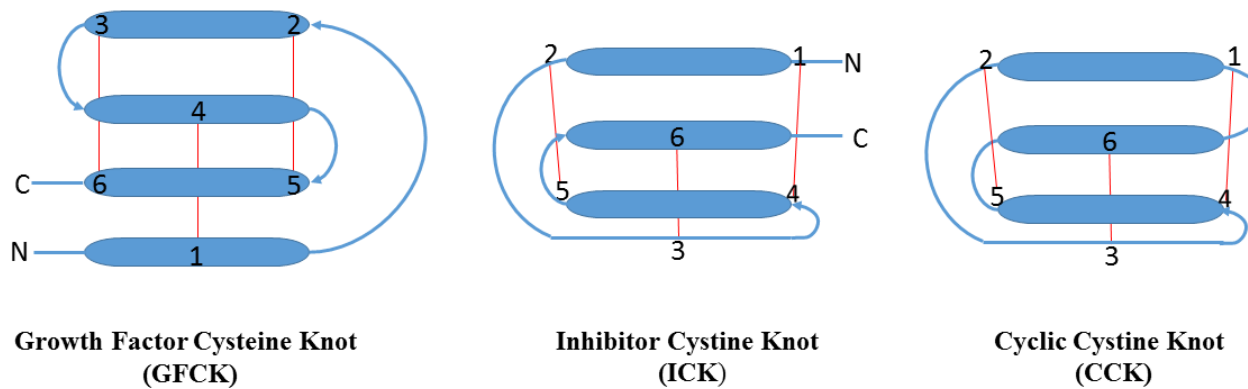


Figure 31. Schematic presentation of the three types of cystine's knots. The ellipsoid shapes represent beta-strands, while the cysteine residues are labelled from 1-6 in order from the N- to C-terminus. Red lines represent the disulfide bonds.

4.3. *Drosophila melanogaster* membrane associated and transmembrane proteins

For the first time a comprehensive analysis of an enriched membrane fraction from fly heads, using a gel-free approach, was reported by Aradska et al. (221). The MS proteomics data have been deposited in the ProteomeXchange Consortium via the PRIDE partner repository with the dataset identifier PXD001712 and 10.6019/PXD001712. A total of 114,865 redundant and 38,179 distinct peptides were identified and assigned to 4812 proteins, including splicing variants. Of those, 3920 proteins (81.4%) were identified with more than two distinct peptides, 624 (13%) with two distinct peptides and 268 (5.6%) with two distinct peptides and multiple hits (Figure 32).

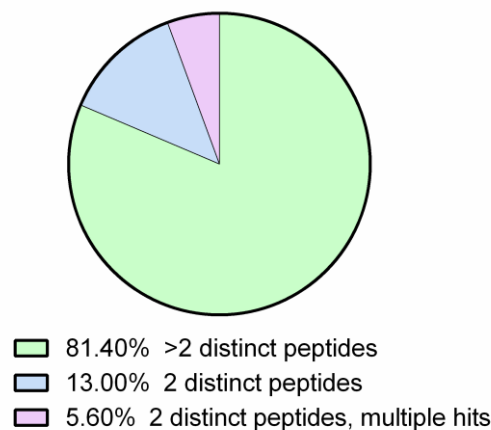


Figure 32. Experimentally identified proteins from membrane fraction of *Drosophila melanogaster*, with respect to the number of distinct peptides

Approximately eight distinct unique peptides were identified per protein. The list of the proteins, arranged according to an exponentially modified protein abundance index (emPAI) (222, 223), is given in Supporting Information Tables S1 (List of experimentally observed proteins) and S2 (List of experimentally observed proteins and peptides) in a paper published in *Proteomics* - Gel-free mass spectrometry analysis of *Drosophila melanogaster* heads (221). All identified proteins correspond to 4559 genes, representing 33,5% of the predicted genes of *Drosophila melanogaster*.

The data from gel-free analysis was classified according to the length of amino acids (Figure 33A) and *pI* value (Figure 33B). Besides this, using the TMHMM algorithm (www.cbs.dtu.dk/services/TMHMM/), transmembrane regions from the identified proteins were predicted (224). 35% of proteins (1679 proteins) were identified as membrane proteins containing at least one transmembrane domain, while 65% (3133 proteins) had no confirmed or predicted transmembrane domain (Figure 34).

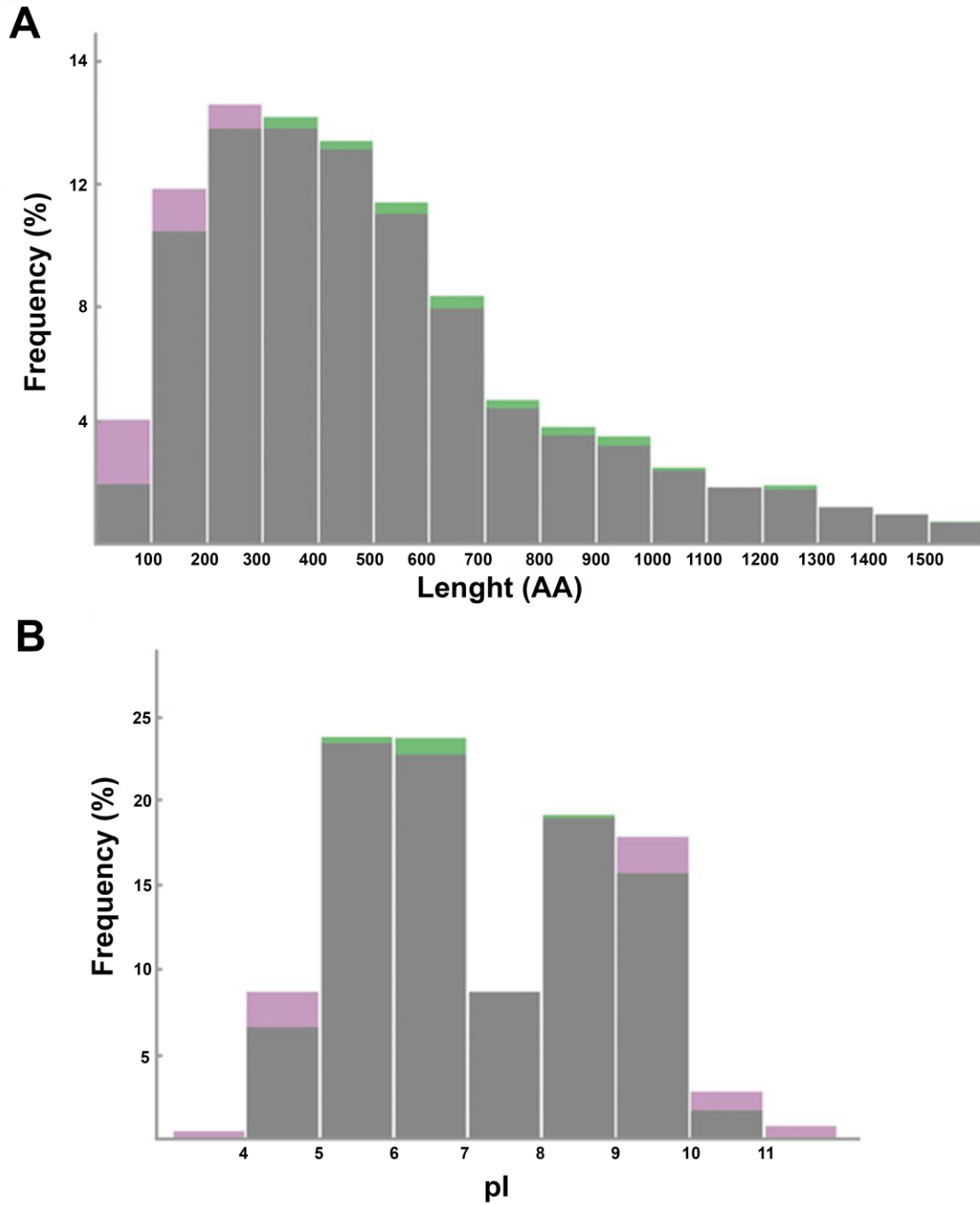


Figure 33. Distributions of length (A) and pI (B) of 4812 experimentally identified proteins in comparison to all proteins (20,328). Under-represented areas are shown in pink, over-represented areas in green

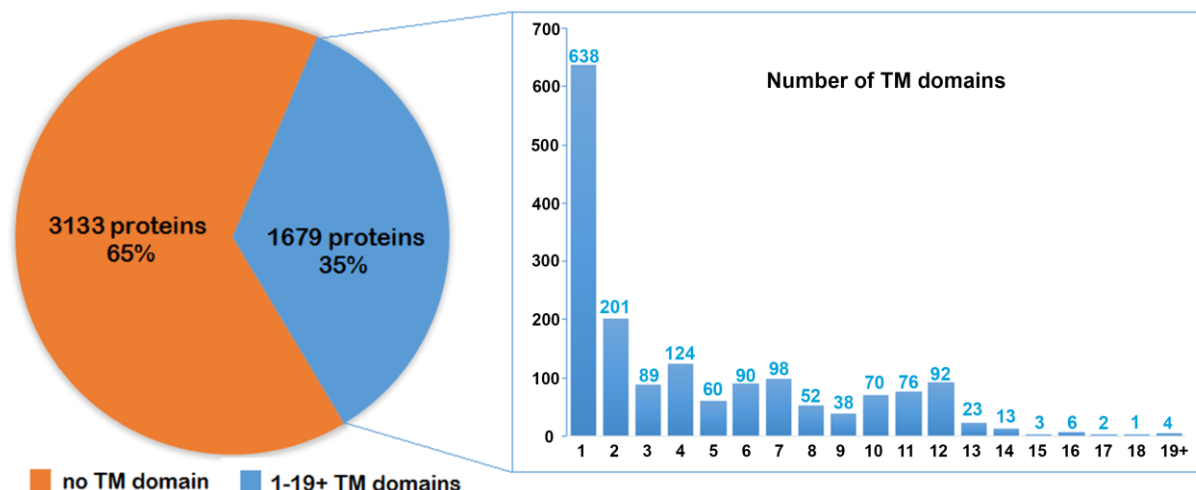


Figure 34. Transmembrane domain distribution among identified proteins

Additionally, the identified proteins were categorized with GO terms, using the AmiGO2 bioinformatics tool (225). AmiGO2 is an open multi-platform software (MATLAB-based) which is used in combination with Gene Ontology (GO), providing information on the ontologies, gene products and annotations, as well as BLAST search, Term Enrichment and GO Slimmer tools (225). For functional analysis, 3565 gene models were considered and an enrichment analysis was performed against *D. melanogaster* background (13 624 gene models). This analysis shows both over-and under-represented GO terms categories. From the data that we obtained, the most under-represented “biological processes” are presented in Figure 35; “molecular functions” and “cellular components” are in Figure 36 and 37, respectively.

Detection of chemical stimuli and sensory perception of chemical stimuli are two of the most under-represented “biological processes”. Proteins involved in odorant binding, olfactory receptor activation and taste receptor activation are the most under-represented, according to AmiGo analysis of “molecular functions” (Figure 33). The most under-represented “cellular components” are: neuron projection membrane, dendrite membrane, leading edge membrane, proteins connected with chemosensory system (Figure 34). They represent the part of the genome which is detected on mRNA, but not on protein level. Chemosensory structures in insects are able to sense an extensive range of volatile, soluble chemicals – signals. Those signals are very important for finding and evaluating the quality of food source, as well as identifying

mates and oviposition sites (226). The olfactory systems of *Drosophila* and mammals are exceptionally similar. They involve the same receptors in olfactory receptor neurons (ORNs), which transmit signals from a single glomerulus (*Drosophila*) or the olfactory bulb (mammals) to the higher brain centres - mushroom body and the lateral horn (*Drosophila*), or to the olfactory cortex (mammals) (227). This similarities make *Drosophila* a great model for medical research, and detection of receptors at the protein level using gel-free methods combined with mass spectrometry opens new perspectives in molecular biology.



Figure 35. Under-represented “biological process” according to AmiGO2 bioinformatics tool

In biomedical research, receptors, transporters, and ion channels are extremely important membrane proteins, since they represent the biggest part of drug targets. The list of identified receptors from our study is presented in Tables 6 and 7. Generally, receptors are divided in two major groups: ionotropic and metabotropic, and receptors belonging to both groups are ligand-gated transmembrane proteins. Neurotransmitters which are released into a synaptic cleft can bind to a receptor and start a signalling cascade. Ionotropic receptors, after binding of a ligand (neurotransmitter), will allow an exchange of ions (K^+ , Na^+ , Cl^- , and Ca^{2+}) between a cell and its extracellular compartment, which triggers a fast response of numerous secondary messengers (228). In contrast, a metabotropic receptor activation initiates a signalling cascade via G-proteins (229).

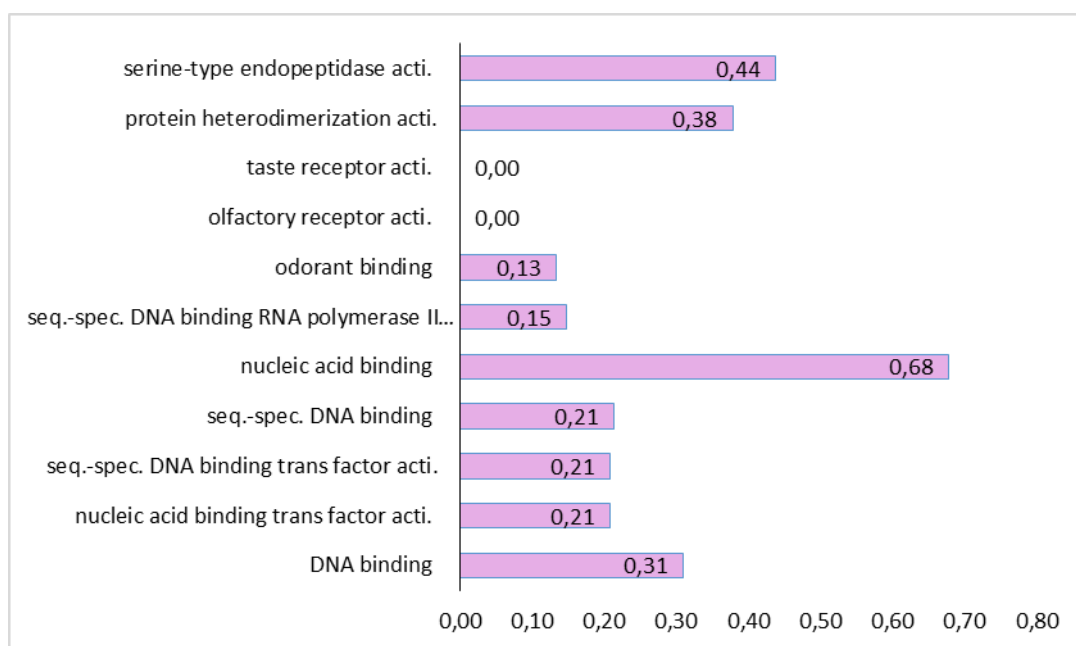


Figure 36. Under-represented “molecular function” according to AmiGO2 bioinformatics tool

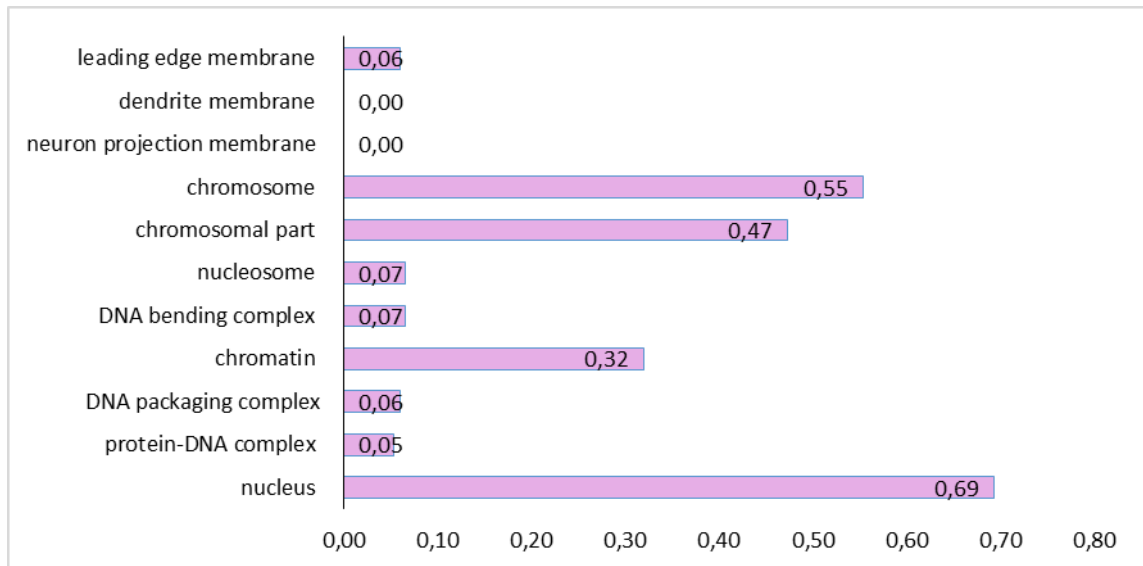


Figure 37. Under-represented “cellular compartment” according to AmiGO2 bioinformatics tool

Ionotropic glutamate receptors are divided into 4 groups: AMPA, 1 kainate, NMDA, and δ receptors, and they play a central role in learning, memory and numerous neurological diseases (230). The most abundant, in regard to emPAI index, are Nmdar1 (0.55) and Nmdar2 (0.41). Nicotinic acetylcholine receptors (nAChRs) are proteins that respond to the neurotransmitter acetylcholine in CNS and peripheral nervous system (PNS) in mammals, whereas in insects it is limited only to the equivalent of CNS. nAChRs are involved in synaptic transmission in the sympathetic/parasympathetic nervous system and as receptors of signals for muscular contraction (231). According to our analysis, $\alpha 4$ (0.29) and $\beta 3$ are the most abundant nicotinic acetylcholine receptor subunits in *Drosophila* brain (0.28).

Table 6. List of ionotropic receptors detected from gel-free MS analysis of *Drosophila melanogaster* heads

Protein Description	Coverage (%)	emP AI	Unique Peptides
IONOTROPIC RECEPTORS			
iGluR			
Glutamate [NMDA] receptor subunit 1 (Nmdar1)	30,7	0,55	25
NMDA receptor 2, isoform G (Nmdar2)	22,6	0,41	19
Glutamate receptor IA (GluRIA)	11,8	0,15	7
Glutamate receptor IB, isoform B (GluRIB)	6,7	0,08	4
nAChR			
Acetylcholine receptor subunit alpha-like 1 (nAChRalpha1)	12,3	0,21	6
Acetylcholine receptor subunit alpha-like 2 (nAChRalpha2)	12,2	0,14	5
Acetylcholine receptor subunit beta-like 2 (nAChRbeta2)	13,3	0,23	5
Nicotinic acetylcholine receptor alpha4, isoform C (nAChRalpha4)	16,9	0,29	6
Nicotinic acetylcholine receptor alpha5, isoform B (nAChRalpha5)	13	0,21	7
Nicotinic acetylcholine receptor alpha6, isoform H (nAChRalpha6)	9,1	0,17	1
Nicotinic acetylcholine receptor Dalpha7 (nAcRalpha18C)	13,3	0,21	4
Nicotinic acetylcholine receptor beta 3 (Dbeta3) subunit (nAChRbeta3)	18,1	0,28	6
Nicotinic acetylcholine receptor (nicra3)	5,6	0,08	2

Table 7. List of metabotropic receptors detected from gel-free MS analysis of *Drosophila melanogaster* heads

Protein Description	Coverage (%)	emP AI	Unique Peptides
METABOTROPIC RECEPTORS			
mGluR			
Metabotropic glutamate receptor, isoform B (mGluR)	14,3	0,21	12
mAChR			
Muscarinic acetylcholine receptor DM1 (mAChR-A)	7	0,11	4
GABA			
Metabotropic GABA-B receptor subtype 2, isoform A (GABA-B-R2)	4,6	0,07	5
Dopamine			
Dopamine receptor 1 (Dop1R1)	13,3	0,24	5
Isoform 606 of Dopamine D2-like receptor (D2R)	8,7	0,14	4
Dopamine/Ecdysteroid receptor, isoform A (DopEcR)	6,2	0,1	2
Octopamine			
Octopamine receptor beta-3R (Octbeta3R)	1,4	0,03	1
Octopamine-Tyramine receptor, isoform B (Oct-TyrR)	15,1	0,21	7
Serotonin			
Serotonin receptor 1B, isoform C (5-HT1B)	14,7	0,14	4
Adenosine			
Adenosine receptor (69)	2,3	0,04	2

Metabotropic receptors also have neurotransmitters as ligands, but they have longer-lasting effects than ionotropic receptors. Receptors involved in memory are divided into several groups: metabotropic glutamate (mGluR), muscarinic acetylcholine (mAChR), gamma-aminobutyric acid (69), serotonin, as well as receptors for norepinephrine, epinephrine, histamine, dopamine, neuropeptides and endocannabinoids (232). From the data that we obtained, the most abundant receptors from the group of metabotropic ones are dopamine receptor 1 (0.24), mGluR (0.21) and octopamine-tyramine receptor (0.21). Dopamine (3,4-dihydroxyphenethylamine) acts as a neurotransmitter in several well-defined distinct pathways, such as motor control, executive functions, arousal, controlling the release of various hormones, and especially in reward-motivated behaviour. Regulation and metabolism of dopamine is also important in Parkinson's

disease, in case of drug and psychostimulants addiction, psychosis and antipsychotic drugs, attention deficit hyperactivity disorder (ADHD), pain in burning mouth syndrome, fibromyalgia, restless legs syndrome, schizophrenia etc. (233-237).

In invertebrates, the monoamines octopamine (OA) and tyramine (TA) take the roles of epinephrine and norepinephrine, which control behaviour, sleep, learning, memory, fight-or-flight response, as well as aggression. Moreover, the mechanism of regulation through OA and TA is very similar to mammalian adrenergic system (238). Adenosine and serotonin receptors are also detected. Adenosine receptors modulate neuronal and synaptic function, whereas serotonin receptors are associated with aggression, anxiety, locomotion, memory and learning (239, 240). Analysis of those receptors at the protein level, using *D. melanogaster* as a model system, may improve the knowledge on the above listed processes.

Ion channels are membrane, pore-forming proteins. Their functions include establishing not only a resting membrane potential, but also modulating action potentials in neurons. Generally, they are controlling the flow of ions in cells, regulating various processes, such as contractions in cardiac, skeletal and smooth muscle system, pancreatic beta-cell insulin release, transport of nutrients and ions in epithelial cells (241, 242). It is known that more than 400 genes encode channel subunits. However, these transcripts can also be alternatively spliced. According to literature, mutation in pathology of ion channels is associated with numerous neurologic disorders (myotonic syndromes, periodic paralyses, partial epilepsies, generalized epilepsies, congenital myasthenic syndromes, hyperekplexia, familial erythremialgia, paraneoplastic channelopathies, etc.) Ion channels can be classified according to:

- 1) Gating
 - Voltage-gated ion channels
 - Ligand-gated ion channel
- 2) Type of ions
 - Chloride channels

- Potassium channels
- Sodium channels
- Calcium channels
- Protons channels
- Non-selective cation channels

3) Localization

- Plasma membrane channels
- Intracellular (endoplasmic reticulum and mitochondrial) channels

Channels which we have detected by gel-free analysis of *D. melanogaster* heads are listed in Table 8. The most abundant one is Calcium channel flower (1,33), followed by diverse types and isoforms of potassium channels (potassium voltage-gated channel protein shaker, inwardly rectifying potassium channel). Calcium channel flower regulates exo- and endocytosis of the synaptic vesicle (SV) in presynaptic terminals, development of photoreceptor terminals and control of gene transcription (243). Two essential functions of SV are transport of proteins involved in an uptake of neurotransmitter and trafficking of other proteins involved in SV endo/exocytosis and recycling. Nevertheless, voltage-gated Ca^{2+} channels are significant transducers of membrane potential changes (244). Potassium channels play an important role in excitable and non-excitable signalling processes, and are linked to diseases like long-QT syndromes, episodic ataxia/myokymia, familial convulsions, hearing/vestibular diseases, Bartter's syndrome, and familial persistent hyperinsulinemic hypoglycemia of infancy (245). Chloride channels are a divergent group of anion-selective channels and their function is related to the regulation of the excitability of neurones, cell volume regulation, the cell cycle and apoptosis (246). Histamine-gated chloride channels (HisCl α 1 and α 2) are well studied since they belong to the Cys-loop receptor superfamily of ligand-gated ion channels, and they are also closely related to the mammalian GABA(A) and glycine receptors (GlyR) (247). The first ion channel which was cloned was one of the sodium channels. Their main characteristic is transmission of

depolarization through the cell networks. Sodium channels play a central role in physiology, because they transmit depolarizing impulses fast throughout cell/cell networks, thus allowing a co-ordination of higher processes like locomotion and cognition (248). At the basic amino acid level, more than 70% of synaptic proteins in *Drosophila* are analogous to mammalian synaptic proteins, which makes *Drosophila* a significant model system for neurobiological studies (249).

A group of proteins that are significant for neurobiological studies is the transporter group. They are integral membrane proteins with a significant role in the transport of ions, proteins and peptides. In contrast to channels, transporters are not open simultaneously to both the extracellular and intracellular environments; either its inner or its outer gate is open. If the movement of a substance across a membrane is against its concentration gradient, they are called active transporters. This process uses chemical energy, such as adenosine triphosphate (ATP), or an electrochemical gradient. On the other hand, transport is also possible without additional energy input. These type of transporters are called facilitated diffusion transporters and they are specific for large polar molecules and charged ions (250). From our analysis, the most abundant ones were amino acid transporter-EAAT1 (1.24) and facilitated trehalose transporter-Tret1-1 (1.02) (Table 9). Glutamate, a dominant neurotransmitter in mammalian CNS, regulates processes in glutamatergic synapses. Besides neurons, glutamates are also present in glial cells (251). EAATs mutations are found in patients with seizures, migraine, cerebellar atrophy, and hemiplegia (252, 253). In addition to that, pathology of EAATs regulation is associated with amyotrophic lateral sclerosis, stroke, epilepsy, schizophrenia, Alzheimer's and Huntington's diseases (254). Trehalose [α -D-glucopyranosyl-(1,1)- α -D-glucopyranoside, Glc(α 1-1 α)Glc] has numerous functions that distinguish it from other disaccharides. These functions are: protection against stress (desiccation, heat, low temperature, and high/low oxygen)(255-257), as well as prevention of osteoporosis (258), oculopharyngeal muscular dystrophy (259), and an induction of the mammalian target of rapamycin-independent autophagy (260). Besides trehalose transporters, glucose transporters, a huge group of membrane proteins which enable the transport of glucose, were also detected. They are present in all phyla. A large number of glucose transporter isophorms, with specific tissue expression, substrate specificity, transport kinetics and altered function in different physiological conditions,

were detected (261). Neural function is also controlled by choline, which is a precursor of the neurotransmitter acetylcholine (ACh) that regulates a huge number of autonomic, cognitive, and motor functions. Choline transporter (CHT) is related with cholinergic dysfunctions such as Alzheimer's disease, Parkinson's disease, schizophrenia, Huntington's disease and dysautonomia (262). Due to the fact that many functional, morphological and molecular characteristics are conserved in *Drosophila*, it represents an optimal model system for studying the above mentioned conditions (263).

Table 8. List of channels detected from gel-free MS analysis of *Drosophila melanogaster* heads

Protein Description	Coverage (%)	emPAI	Unique Peptides
Ca ²⁺ -channel protein alpha[[1]] subunit D, isoform H (Ca-alpha1D)	5,9	0,06	8
Ca ²⁺ -channel protein alpha[[1]] subunit T, isoform C (Ca-alpha1T)	4,4	0,05	7
Ca ²⁺ -channel-protein-beta-subunit, isoform G (Ca-beta)	7,2	0,1	2
Ca ²⁺ -channel-protein-beta-subunit, isoform P (Ca-beta)	6,7	0,11	1
Calcium channel flower	39,2	1,33	5
Chloride channel protein (CG5284-RB)	22,6	0,38	15
Chloride channel protein (ClC-a-RF)	16,5	0,25	13
Chloride channel protein (ClC-b)	22	0,39	14
Cyclic nucleotide-gated ion channel-like, isoform E (Cngl)	4,6	0,03	4
Eag-like K ⁺ channel (Elk)	6,6	0,08	5
Histamine-gated chloride channel subunit (ort)	10,1	0,1	5
Histamine-gated chloride channel subunit 1, isoform D (HisCl1)	5,5	0,07	1
I[[h]] channel, isoform E (Ih)	13,2	0,22	7
Inwardly rectifying potassium channel 3, isoform C (Irk3)	12,7	0,16	1
Inwardly rectifying potassium channel 3, isoform D (Irk3)	20,4	0,32	6
Inwardly rectifying potassium channel, isoform C (Irk1)	20,2	0,43	9
Isoform L of Potassium voltage-gated channel protein Shaker (Sh)	28	0,44	1
Isoform L of Potassium voltage-gated channel protein Shaker (Sh)	24,4	0,42	1
Open rectifier K ⁺ channel 1, isoform D (Ork1)	13	0,21	10
Potassium voltage-gated channel protein Shaker (Sh)	30,4	0,43	1
Small conductance calcium-activated potassium channel, isoform U (SK)	4,9	0,07	4
Sodium channel protein (para)	5,8	0,06	8
Sodium channel protein 60E (NaCP60E)	7,4	0,07	12
Transient receptor potential cation channel protein painless (pain)	21,6	0,45	20
Voltage-dependent calcium channel alpha13 subunit (Ca-alpha1T)	10,4	0,08	1

Table 9. List of transporters detected from gel-free MS analysis of *Drosophila melanogaster* heads

Protein Description	Coverage (%)	emPAI	Unique Peptides
ABC transporter expressed in trachea, isoform C (Atet)	15,4	0,17	7
Adenosine 3~-phospho 5~-phosphosulfate transporter 2 (Papst2)	13,9	0,17	4
Amino acid transmembrane transporter (CG4991)	6,5	0,07	2
Amino acid transporter (Eaat1)	31,9	1,24	16
Amino acid transporter protein (JHI-21)	8,8	0,21	6
CMP-sialic acid/UDP-galactose transporter (Csat)	12,9	0,2	4
ER GDP-fucose transporter, isoform B (Efr)	15,9	0,3	5
Facilitated trehalose transporter (Tret1-1)	28,4	1,02	10
Facilitated trehalose transporter (Tret1-2)	11,7	0,3	2
GDP-fucose transporter 1 (nac)	16	0,26	5
Glucose transporter type 1 (Glut1)	11,8	0,18	4
Glucose transporter type 1 (Glut1)	21,9	0,44	7
High-affinity choline transporter 1 (CG7708)	21,5	0,52	12
Isoform A of Putative inorganic phosphate cotransporter (Picot)	24,2	0,5	12
Major facilitator superfamily transporter 3 (MFS3)	11,5	0,14	4
Na[+]-dependent inorganic phosphate cotransporter (NaPi-T)	11,6	0,27	7
Organic cation transporter protein (Orct)	25	0,58	11
Organic cation transporter-like protein (Orct2)	11,3	0,21	4
Peptide transporter family 1 (yin)	9,4	0,14	4
Putative sodium-dependent multivitamin transporter (CG32669)	11,6	0,2	6
Sodium chloride cotransporter 69, isoform C (Ncc69)	33,6	0,66	2
Transporter (CG10804)	14,7	0,24	8
Transporter (CG13793-RA)	8,3	0,16	4
Transporter (CG13794)	11,8	0,23	5
Transporter (CG13795)	12,3	0,23	5
Transporter (CG15279)	11,6	0,16	6
Transporter (CG1732)	24,7	0,7	19
Transporter (CG3252-RA)	12,2	0,19	7
Transporter (CG5226-RA)	19,6	0,45	13
Transporter (CG5549)(glycin)	21,6	0,29	15
Transporter (DAT)	5,5	0,08	3
Transporter (ine)	15,6	0,24	11
Transporter (List)	11,9	0,15	5
Vesicular glutamate transporter isoform A (Vglut)	21,4	0,39	10

5. CONCLUSIONS

During the work in this thesis, we defined the following aims:

- c) Whole body transcriptome of *Arion vulgaris*, in order to create mRNA-derived protein database
- d) Proteomic profiling of *Arion vulgaris* mantle using nano-LC-ESI-MS/MS analysis.
- b) Identification of membrane associated and transmembrane proteins from *Drosophila melanogaster* heads using gel-free MS/MS approach

5.1. Whole body transcriptome of *Arion vulgaris*

Using Illumina HiSeq 2000 technology we created a cDNA library which contains 339 millions of paired end reads. Using Trinity pipeline we assembled 136,406 contigs of *de novo* transcript, finally resulting in 47,451 protein-coding sequences.

In comparison with non-redundant data base of NCBI we obtained the following results: 63.3% of transcriptome, which is 31,463 transcripts, was assigned to at least one protein, while 33.7% (15,988 transcripts) was with no significant hits.

0.4% (127 transcripts) represents unknown or unnamed proteins.

65% of sequences is shared with *Aplysia californica*, 8.7% with *Crassostea gigas* and 2.5% with *Capitella teleta*. More than 80% of the sequences best matched with mollusc species.

According to Pfam database, in 16,994 entries at least one domain was recognized, and of those entries, 471 sequences were without blastX hit annotation. Three most abundant domains are zinc-finger double domain (zf-H2C2_2), domain of unknown function (DUF4200) and ankyrin repeat (Ank_5).

1,796 of transcripts were assigned with EC (Enzyme Commission) numbers which specify enzyme-catalysed reactions.

5.2. *Arion vulgaris* mantle proteome

We identified 833 unique proteins from 2-DE gel. 814 proteins had a significant hit in NCBI nr database, while 19 proteins are with no significant homology in nr NCBI database.

With combination of 1-DE gel and LC/MS-MS with higher resolution, we identified 2011 proteins, from which 48 proteins are without a hit on nr NCBI database.

Total number of identified proteins from 1-D and 2-D gels, with minimum of one unique peptide and distinct peptides, is 2129.

We identified wide-range of proteins which indicates an unexpected complexity of the immune system, with a high grade of pathogen specificity and immune-priming: cytokine-like proteins, complement-like proteins, peptidoglycan-recognition proteins (PGRPs), lipopolysaccharide and β 1, 3-glucan-binding proteins, fibrinogen-related proteins (FREPs), pore-forming membrane attack/perforin (MACPF) domain proteins and toxin-like proteins.

Lectin-associated functions may be attributed to 149 transcripts, 37 of which were unambiguously identified at the protein level. Lectins that we detected are: C-type lectin family, galectin, malectin, H-type lectins, L-type lectins, Ricin-B lectins and calnexin. They contain conserved carbohydrate-recognition domains (CRDs).

We discovered novel toxin-like proteins with significant similarity to agatoxin in a signal peptide, pre-peptide sequence and characteristic cysteine motive in the mature peptide.

5.3. *Drosophila melanogaster* membrane associated and transmembrane proteins

Using a comprehensive analysis with a gel-free approach of an enriched membrane fraction from fly heads, we identified 114,865 redundant and 38,179 distinct peptides and they were assigned to 4812 proteins, including splicing variants.

3920 proteins (81.4%) were identified with more than two distinct peptides, 624 (13%) with two distinct peptides and 268 (5.6%) with two distinct peptides and multiple hits.

35% of proteins (1679 proteins) were identified as membrane proteins containing at least one transmembrane domain.

According to AmiGo analysis of our data we showed that detection of chemical stimuli and sensory perception of chemical stimuli are two of the most under-represented “biological processes”. Proteins involved in odorant binding, olfactory receptor activation and taste receptor activation are the most under-represented. The most under-represented “cellular components” are: neuron projection membrane, dendrite membrane, leading edge membrane, proteins connected with chemosensory system.

6. EXPERIMENTAL PART

6.1. Chemicals, solutions and buffers

All chemicals and solutions used during experiments were analytical, technical, or LC/MS- grade quality. Details about solutions, chemicals and commercial kits are given in Table 10.

Table 10. List of used chemicals, solutions and commercial kits

Name and abbreviations	Catalog number	Grade quality	Company
4-(2-Hydroxyethyl) piperazine-1-ethanesulfonic acid, N-(2-Hydroxyethyl) piperazine-N'-(2-ethanesulfonic acid) (HEPES)	H4034	Analytical	Sigma Aldrich
Sucrose	S0389		
Phenylmethanesulfonyl fluoride (PMSF)	P7626		
Ammonium bicarbonate	A6141		
Ammonium-sulfate	A4418		
Tris	252859		
Thiourea	T8656		
Iodoacetamide	I1149		
Ethylenediaminetetraacetic acid (EDTA)	E9884		
Ammonium persulfate (APS)	A3678		
Glycerol	G5516		
Tributylphosphate (TBP)	240494		
Isopropanol	I9516		
N,N,N',N'-Tetramethylethylenediamine (TEMED)	T9281		
Hydrochloric acid (HCl)	84415		
Glycine	G8898		
Triethylammonium bicarbonate buffer (TEAB)	17902	LC/MS	
Ammonium formate	17843		

Name and abbreviations	Catalog number	Grade quality	Company
1,4-Dithiothreitol DTT	6908	Analytical	Carl Roth
3-[(3-Cholamidopropyl) dimethylammonio]-1- propansulfonat (CHAPS)	1479		
Urea	2317.1	Technical	
Acrylamide	7906		
Bis-Acrylamide	6755.2		
Acetic acid	6755.2		
Trypsin	V5111	LC/MS	Promega
Chymotrypsin	V1062	LC/MS	Fisher Scientific
Methanol	M/4062/17		
Acetonitrile	A/0638/17	LC/MS	Merck
Formic acid	1.11670.1000		
Triflouracetic acid (TFA)	1.08262.0025		
Isopropanol	1010402500		
Orthophosphoric acid	100573	Technical	
SDS	1610302	Analytical	Bio Rad
Piperazine diacrylamide	161-0202		
Coomassine G-250	1610406	Molecular biology	
Bromophenol blue	1610404		
2-DEclean up kit	1632130		
Protein marker	1610373		
Immobiline DryStrip Gels	17-6003-76	Molecular biology	GE Helthcare
Carrier (3–11pH)	17045601		
Dry cover strip fluid	17-1335-01		
cOmplete protease inhibitor cocktail	11836145001	Molecular biology	Roche
Pierce 660 protein assay	22660	Molecular biology	Thermo Scientific
Colloidal coomassie blue	LC6025	Molecular biology	Novex Life technologies
RNeasy kit	74104	Molecular biology	Qiagen
NEBNext kit	#E7420S/L	Molecular biology	NEB

Following buffers were used during experimental procedures:

1. Urea Buffer

7 M Urea
2 M Thiourea
4 % CHAPS
2 M Tris pH 8,8
0,5 % Carrier (3–11pH)
1 % DTT
1 mM PMSF
1X cOmplete protease inhibitor cocktail

2. Homogenization buffer

10 mM HEPES pH 7,5
300 mM Sucrose
1X complete protease inhibitor cocktail

3. Washing buffer

10 mM HEPES pH 7,5
1X cOmplete protease inhibitor cocktail

4. SDS extraction buffer

1,5 % SDS
100 mM NaCl
20 mM Tris-HCl pH 6,5
1X cOmplete protease inhibitor cocktail

5. Urea A buffer (UA buffer)

8 M Urea
100 mM Tris-HCl (pH 8,5)

6. Rehydration buffer

8 M Urea
10 mM DTT
4 % CHAPS
0,5 % Carrier (3–11pH)
0,008 % Bromphenol Blue

7. Equilibration buffer 1

50 mM Tris-HCl, pH 8,8
6 M Urea
30 % Glycerol
2 % SDS
1 % DTT

8. Equilibration buffer 2

50 mM Tris-HCl, pH 8,8
6 M Urea
30 % Glycerol
2 % SDS
4 % Iodoacetamide
0,002 % Bromophenol blue

9. Leammli buffer

150 mM Tris-HCl pH 6,8
300 mM DTT
6 % SDS
0,3 % Bromophenol blue
30 % Glycerol

10. SDS running buffer

0,025 M Tris
0,192 M Glycine
0,1 % SDS

11. Fixation buffer

50 % Methanol
10 % Acetic acid

12. Blue silver staining solution

12,1 % Orthophosphoric acid
20 % Methanol
0,1 g/mL Ammonium-sulfate
0,0012 g/mL Coomassine G-250

13. Stage Tip Buffer

0,4 % Formic acid
2 % TFA

14. STP Elution Buffer

0,4 % Formic acid

90 % Acetonitrile

15. Solution A for HPLC coupled with amaZon

0,1 % Formic acid

16. Solution B for HPLC coupled with amaZon

0,08 % Formic acid

80 % Acetonitrile

17. Solution A for HPLC coupled with LTQ-OrbitrapVelos

0,4 % Formic acid

18. Solution B for HPLC coupled with LTQ-OrbitrapVelos

0,4 % Formic acid

70 % Methanol

20 % Isopropanol

19. Solution A for HPLC coupled with Thermo Orbitrap Velos Pro

0,01 % Triflouracetic acid

20. Solution B for HPLC coupled with Thermo Orbitrap Velos Pro

0,05 % Triflouracetic acid

80 % Acetonitrile

21. Solution A for HPLC for off-line fractionation

5 % Acetonitrile
20 mM Ammonium formate

22. Solution B for HPLC for off-line fractionation

90 % Acetonitrile
20 mM Ammonium formate

6.2. Apparatus and procedures

6.2.1. Protein isolation and purification

6.2.1.1. *Arion vulgaris*

Arion vulgaris specimens were collected in Woerden, Austria (48°19'40" N, 16°12'34" E) in May 2014. In total, 15 specimens were collected, ranging from 7 to 8 g. Species identification was carried out with the assistance of the Zoological Department of Museum of Natural History in Vienna. According to national law no permission is required to collect and use invertebrates. Individuals of *A. vulgaris* were frozen immediately after harvest in liquid nitrogen and stored at -80°. Mantles were dissected directly before protein extraction procedure. Samples were powdered with mortar and pestle in liquid nitrogen and homogenized in urea buffer. Samples were sonicated (Ultra-TurraxT25, Staufen, Germany) using 5 sec pulse, 5 times in row, following incubation on RT for 1 h, and at 4°C for 2h, with vortexing every 15 min. Samples were centrifuged with Eppendorf benchtop centrifuge (18,000 \times g) at 10°C for 1h. Supernatant was desalted at 1000 \times g with fresh urea buffer in Amicon Ultra-15 Centrifugal filter unit Ultracel-3 membrane (Merck Millipore). Samples were centrifuged until 500 μ l remain.

Protein concentrations were estimated with Pierce 660 nm Protein Assay kit. Samples were cleaned using a 2-DE clean up kit (ReadyPrep™ 2-D Cleanup Kit, BioRad), according to the manufacturer's instructions. Briefly, 400 μ g of sample was

mixed with precipitating agent No#1, and incubated on ice for 15 min before adding precipitating agent No#2. Pellet was obtained by centrifugation of previously mentioned mixture at 12,000 x g for 5 min, following washing with LC/MS water and Washing reagent No#2. Pellet was incubated at -20°C for 30 min, centrifuged at 12,000 x g for 5 min, air dried no more than 5 min and re-suspended by adding an appropriate volume of 2-D rehydration/sample buffer.

6.2.1.2. *Drosophila melanogaster*

Drosophila melanogaster heads were isolated from adult flies using a standard freezing protocol (264). Briefly, flies were collected in vials and flash-frozen by immersion of the vials in liquid nitrogen. Heads were isolated with sieve (upper chamber containing largest mesh for collecting bodies while lower chamber was for collecting heads). Sieve, containing flies, was dipped in the liquid nitrogen and shaken vigorously for 1 min. Heads were then ground into a fine powder with a pre-chilled mortar and pestle. Powder was resuspended in ice-cold homogenization buffer. The homogenate was centrifuged for 10 min at 1000 × g and the supernatant was centrifuged at 50 000 × g for 30 min at 4°C. Subsequently, the pellet was resuspended in 4 mL washing buffer, kept on ice for 30 min and centrifuged at 50 000 × g for 30 min. A total plasma membrane fraction was separated by equilibrium density centrifugation on a discontinuous sucrose gradient. The pellet, resuspended in washing buffer, was layered on top of the sucrose cushion (1 M and 1.25 M sucrose solution) followed by centrifugation at 70 000 × g for 2 h at 4 °C. After centrifugation, fractions from the sucrose interface were collected and diluted ten times with washing buffer and subsequently centrifuged for 30 min at 100 000 × g. The pellet was stored at 80 °C until use. Membrane protein extraction was carried out with urea buffer according to a previous study with minor modifications (265). Protein quantities were estimated by the Pierce 660 protein assay.

6.2.2. Gel electrophoresis and staining

6.2.2.1. One dimensional electrophoresis (1D)

One dimensional electrophoresis was performed on 10% polyacrylamide gel (10% Acrylamide/bis-acrylamide (29:1), 0.625 M Tris-HCl pH 8.8, 0.1% SDS, 0.1% APS, 0.1% TEMED). 4% resolving gel was used in this experiment (4% Acrylamide/bis-acrylamide (29:1), 0.125 M Tris-HCl pH 6.8, 0.1% SDS, 0.1% APS, 0.1% TEMED). 50 µg of extracted proteins were mixed with Laemmli buffer at a 1:3 volume ratio. During electrophoresis voltage was set up as constant: 30 min 50 V, 30 min 100 V and 1h 150 V.

6.2.2.2. Two dimensional electrophoresis (2D-IEF/SDS-PAGE)

2-DE was performed as reported previously with minor modifications (266-268). 750 µg of proteins were loaded on 18 cm pH 3–11 linear IPG strips (GE Healthcare). IEF was performed in a IPGphor electrophoretic system (GE Healthcare, Uppsala, Sweden) using a protocol with gradually increasing voltage from 200 to 8,000 V at 4V/min. Prior to second dimension, IPG strips were incubated in 10 ml of equilibration buffer 1 for 15 min with gentle shaking followed by incubation in 10 ml of equilibration buffer 2. The second-dimensional separation was performed on 10–16% gradient SDS-PAGE gels. During electrophoresis voltage was set up as constant: 14 h 50 V, 2 h 100 V and 2 h 150 V. Molecular masses were determined by comparison with precision Bio Rad protein standard markers spanning from 10 to 250 kDa molecular weight range. After fixation for 4 h in fixation buffer, gels were stained over night with the colloidal coomassie blue stain with mild shaking in cold room at +4 °C.

6.2.3. In gel digestion

All spots were excised from 2-DE gels, and digested with trypsin or chymotrypsin. The gel pieces were washed with 50 % Acetonitrile (69) in 10 mM ammonium bicarbonate for 20 min with vortexing and then 20 min with 10 mM

ammonium bicarbonate. These two steps were repeated until the gel was completely destained. 100 % ACN (69) was added for 10 min and gel pieces were dried completely using Eppendorf concentrator for 30 min at 30 °C. Cysteine residues were reduced with 10 mM DTT at 56 °C for 30 min and alkylated with 55 mM Iodoacetamide for 45 min at 22 °C. After washing with 10 mM ammonium bicarbonate and dehydration with 100% ACN, proteins were digested with 12.5 ng/μl trypsin solution buffered in 10 mM ammonium bicarbonate for 16 h (overnight) at 37 °C. The supernatant was transferred to new Eppendorf LoBind 0.5 mL tubes and peptides were extracted with 1 % formic acid and then with 15 % ACN/0.1 % formic acid (269). 20 μL of extracted peptides were analysed by nano-LC-ESI-(CID/ETD)-MS/MS.

6.2.4. Filter aided sample preparation (FASP)

Membrane protein extraction was carried out according to a previous study with minor modifications (265). 300 μg of the sample (1 × 100 μg each) were digested with trypsin using the filter aided sample preparation (FASP) as previously described with minor modifications (83, 270). Samples were mixed in the filter unit with Urea A buffer (UA) buffer and centrifuged at 14 000 × g for 15 min at 22 °C. Any remaining detergent was exchanged by urea with 200 μl UA. The proteins were alkylated by the addition of 100 μl of 50 mM iodoacetamide in UA and incubated for 30 min at 22 °C. Subsequently, excess reagent was removed by 3 × 100 μL UA solution followed by 3×100 μl of 50 mM TEAB buffer. Proteins were digested overnight at 37 °C with trypsin (an enzyme to protein ratio of 1:100 w/w). Tryptic peptides were recovered from the filter with 40 μL of 50 mM TEAB buffer followed by 50 μl of 0.5 M NaCl. The remaining semi-digested material on the FASP filter was washed with 50 μl 50 mM TEAB buffer and further digested with chymotrypsin (an enzyme to protein ratio of 1:10 w/w). Pooled tryptically-digested and sequential trypsin-chymotrypsin digested samples were acidified, desalted and concentrated with customized reversed phase C18 stage tips (271). Bound peptides were eluted with a buffer containing 90 % acetonitrile and 100 mM TEAB and lyophilized in a vacuum concentrator (Eppendorf, Hamburg, Germany).

6.2.5. Solid phase extraction

The peptides obtained after tryptic digestion were cleaned with ultra-microspin column. Briefly, columns were activated with 2 x 200 μ l methanol and centrifuged in 100 x g for 1 min. After that, they were washed 2 x 400 μ l with stage tip buffer. Acidified samples were loaded and washed with 400 μ l of stage tip buffer. Peptides were eluted with 200 μ l stage tip elution buffer, lyophilized on Eppendorf concentrator and kept at -20 °C, or proceeded immediately for experiments.

6.2.6. Off-line fractionation

Peptides generated in FASP were prefractionated according to protocol that was published by Gilar and coworkers (272). Peptide digests were pooled, purified by solid phase extraction (81) (UltraMicroSpin columns 3–30 μ g capacity, Nest Group Inc., Southboro, MA, USA) and separated at pH 10 by reversed-phase liquid chromatography using an Agilent 1200 series HPLC system. Separation was performed at a flow rate of 100 μ l/min on a Phenomenex column (150 \times 2.0 mm Gemini-NX 3 μ m C18 110 Å, Phenomenex) with a 50 min linear gradient from 5–70% (vol/vol) acetonitrile containing 20 mM ammonium formate. Seventy two time-based fractions were collected, acidified, and pooled into 20 or 50 fractions. Peptides were lyophilized in a vacuum centrifuge and reconstituted in 5% (vol/vol) formic acid for on-line LCMS. Peptide abundance per fraction was estimated based on the UV trace and samples were diluted accordingly to avoid overloading the nano-LCMS system.

6.2.7. LC-MS/MS analysis

6.2.7.1. Nano-LC-MS/MS (amaZon speed ETD)

Nano-LC-MS/MS system was used during execution of these experiments. The HPLC used for peptide separation was an Ultimate 3000 system (Dionex Corporation, Sunnyvale, CA) equipped with a PepMap100 C-18 trap column (300 μ m \times

5 mm) and PepMap100 C18 analytical column (75 μm \times 250 mm). The Amazon speed ETD (Bruker Daltonics, Bremen, Germany) was used to record peptide spectra over the mass range of m/z 400–1,400 and MS/MS spectra in information-dependent data acquisition over the mass range of m/z 100–2,800. Repeatedly, MS spectra were recorded followed by three data-dependent CID MS/MS spectra. An active exclusion of 0.4 min after two spectra was used to detect low abundant peptides. The voltage between ion spray tip and spray shield was set to 1,400 V. Drying nitrogen gas was heated to 150 $^{\circ}\text{C}$ and the flowrate was 3 l/min. The collision energy was set automatically according to the mass and charge state of the peptides chosen for fragmentation. Multiple charged peptides were chosen for MS/MS experiments due to their good fragmentation characteristics. MS/MS spectra were interpreted and peak lists were generated by Data Analysis 4.1 (Bruker Daltonics, Bremen, Germany).

6.2.7.2. Nano-LC-MS/MS (LTQ-Orbitrap Velos)

Samples were analysed on a linear trap quadrupole (LTQ) Orbitrap Velos (Thermo Fisher Scientific, Waltham, MA, USA) coupled to an Agilent 1200 HPLC nano flow system (Agilent Biotechnologies, Palo Alto, CA, USA). Data were acquired using Xcalibur (v2.1.0). The peptide mixture was automatically loaded onto a trap column (Zorbax 300SB-C18 5 μm , 5 \times 0.3 mm, Agilent Biotechnologies), and eluted by back-flushing onto a 16 cm fused silica analytical column packed with C18 reversed phase material (ReproSil-Pur 120 C18-AQ, 3 μm , Dr. Maisch GmbH, Ammerbuch-Entringen, Germany). MS^1 scans were performed from m/z 350-1800 at a resolution of 60,000. Using a data-dependent acquisition mode, the 10 most intense precursor ions with a minimum signal threshold of 5,000 from the MS^1 spectrum were isolated (within a 1.2 Da window) and fragmented to obtain the corresponding MS^2 spectrum. The fragment ions were generated in a higher-energy collisional dissociation (HCD) cell at an NCE of 37% and were detected in an Orbitrap mass analyser at a resolution of 7,500. The dynamic exclusion for the selected ions was 60 s and single-charged precursor ions were excluded.

6.2.7.3. Nano Nano-LC-MS/MS (Orbitrap Velos PRO)

Peptide fractions were separated by nanoLC (Dionex Ultimate 3000) equipped with a μ -precolumn (C18, 5 μ m, pore size 100 Å, 5 \times 0.3 mm, Thermo Fisher Scientific, Vienna, Austria) and an Acclaim PepMap RSLC nanocolumn (C18, 2 μ m, 150 \times 0.075 mm, Thermo Fisher Scientific, Vienna, Austria). The sample was ionized in the nanospray source equipped with stainless steel emitters (Thermo Fisher Scientific, Vienna, Austria) and analysed in a Thermo Orbitrap Velos Pro mass spectrometer in positive ion mode by alternating full-scan MS (m/z 380–2000) in the ICR cell and MS/MS by CID of the 20 most intense peaks in the ion trap with dynamic exclusion enabled.

6.3. Transcriptome sequencing and assembly

6.3.1. Isolation of RNA

Isolation of RNA was performed using RNeasy kit (Qiagen, Hilden, Germany). After removal of the gut, two specimens of *Arion vulgaris* were immersed in liquid nitrogen and pulverized with the help of a mortar and pestle and homogenized in QIAzol Lysis Reagent. Adding gDNA Eliminator Solution and chloroform enabled separation of aqueous (RNA partitions) and organic phases/interphase (DNA and proteins) by centrifugation. The upper, aqueous phase was collected, mixed with ethanol, and RNA was purified using RNeasy spin columns. The aqueous phase was mixed with ethanol to provide appropriate binding conditions, and applied to an RNeasy Mini spin column. Total RNA binds to the spin column membrane, while phenols and other contaminants are efficiently washed away. High-quality RNA was then eluted in RNase-free water.

6.3.2. mRNA isolation, fragmentation and priming

The first step in the workflow involves purifying the poly(A)-containing mRNA molecules using two rounds of poly(T)-oligo-attached magnetic beads. Pooled

total RNA was subjected to RNA-Seq following the mRNA sequencing protocol provided by New England Biolabs (NEBNext kit, NEB, Frankfurt, Germany). Briefly, total RNA was diluted with nuclease-free water in a nuclease-free PCR tube and kept on ice, followed by adding NEBNext Oligo d (T) 25 beads and mixing. Solution was on the magnetic rack at room temperature for 2 min and supernatant was removed from tube. These steps were repeated 2 times. After that beads were resuspended in RNA Binding Buffer and 50 μ l of total RNA. Tube was incubated for 5 min at 65 °C and then cooled down to 4 °C to denature the RNA and facilitate binding of the poly-A mRNA to the beads following 5 min incubation at room temperature. The tube was on the magnetic rack at room temperature for 2 minutes in order to separate the poly-A mRNA binding beads from the solution. The supernatant was removed, while the pellet was washed 2 times with wash buffer. After adding 50 μ l of tris buffer, sample was mixed, incubated at 80 °C for 2 minutes, and then kept at 25 °C to elute the Poly-A mRNA from the beads. Again, 50 μ l of RNA Binding Buffer was added to the sample to allow the mRNA to rebind to the beads. And like in previous step, complete mixture was incubated at room temperature for 5 min, placed on magnetic rack for 2 min, after which supernatant was removed. Samples were washed with wash buffer and then with tris buffer. Removing of beads was performed with 15 μ l of the first strand synthesis reaction buffer and random primer mix (2X) at 94 °C for 15 minutes.

6.3.3. First strand and second cDNA synthesis

The RNA fragments were copied into first-strand complementary DNA (cDNA) using reverse transcriptase and random primers. 10 μ l of mRNA was mixed with Murine RNase Inhibitor, Actinomycin D (0.1 μ g/ μ l), ProtoScript II Reverse Transcriptase (NEBNext kit, NEB) and nuclease free water according to the manufacturer's instructions. Samples were incubated in a preheated thermal cycler: 10 minutes at 25 °C, 15 minutes at 42 °C, 15 minutes at 70 °C and hold at 4 °C. In order to perform second cDNA synthesis, the following solutions are added to previous mix, according to the manufacturer's instructions: nuclease-free water, second strand synthesis reaction buffer (10X) and second strand synthesis enzyme mix and incubated at 25 °C for 1h, with heated lid set at \leq 40 °C.

6.3.4. Purifying the double-stranded cDNA

In order to purify double-stranded cDNA, 1.8X Agencourt AMPure XP beads (NEBNext kit, NEB) were used. 144 µl of the beads was mixed with second strand synthesis reaction (~80 µl), followed by incubation for 5 min at room temperature. While DNA targets were attached to beads, supernatant was removed after 5 min incubation of mixture on magnetic rack. Pellet was washed 2 times with 80 % ethanol and air dried for 10 min on magnetic rack. Eluting of DNA target was performed with nuclease-free water. Samples were mixed with nuclease-free water, vortexed, quickly spun and placed on magnetic rack. Supernatant was used for following steps.

6.3.5. End repair/dA-tail of cDNA library and adaptor ligation

The purified double-stranded cDNA was mixed with dA-Tailed cDNA, NEBNext End Repair Reaction Buffer (10X) and NEBNext End Prep Enzyme Mix (NEBNext kit, NEB) according to the manufacturer's instructions. Samples were incubated in a thermal cycler as follows: 30 minutes at 20 °C, 30 minutes at 65 °C and hold at 4 °C. Blunt/TA ligase master mix, diluted NEBNext adaptor and nuclease-free water were mixed and incubated 15 minutes at 20 °C. Nuclease-free water was used to adjust volume to 100 µl, followed by mixing with the sample from the previous step, and incubation on room temperature for 5 min. Supernatant (containing unwanted fragments) was discarded and pellet was washed two times with 80 % ethanol and air dried. DNA target was eluted with nuclease-free water, mixed with 1 X AMPure XP beads, incubated for 5 min and supernatant was separated from beads on magnetic rack. Pellet was washed 2 times with 80 % ethanol and air dried. DNA target was eluted with nuclease-free water and it was proceeded to the next step.

6.3.6. USER excision, PCR library enrichment and purifying

Sample was mixed with NEBNext USER enzyme, NEBNext high-fidelity PCR master mix (2X), Universal PCR primer and index (273) primer according to the

manufacturer's instructions. PCR cycling conditions were: USER digestion (37 °C, 15 min, and 1 cycle), initial denaturation (98 °C, 30 seconds, and 1 cycle), denaturation (98° C, 10 sec, and 15 cycles), annealing (65 °C, 30 sec, and 15 cycles), extension (72 °C, 30 sec, and 15 cycles), final extension (72 °C, 5 min, 1 cycle). Samples were mixed with AMPure XP beads and incubated 5 min at room temperature. After 5 min at magnetic rack, supernatant was discarded, pellet was washed two times with 80 % ethanol and air dried. DNA target (size distribution of ~200–250 bp) was eluted with nuclease-free water and stored at –20 °C.

6.3.7. Selection of Adaptor-ligated DNA and sequencing

Sample was mixed with AMPure XP beads (NEBNext kit, NEB) according to the manufacturer's instructions and incubated 5 min at room temperature. Magnetic stand was used to separate the beads (containing unwanted large fragments) from the supernatant (containing DNA). Procedure was repeated, but in this step supernatant was discarded and pellet (containing DNA target) was washed two times with 80 % ethanol and air dried. DNA target was eluted with 10 mM Tris-HCl and transferred to a new PCR tube. Finally, the adaptor-ligated DNA was sequenced for 100 cycles on a HiSeq2000 sequencing system (Illumina, San Diego, USA).

Schematic presentation of preparation of biological material for RNA library for next-generation sequencing is given in Figure 38, according to the manufacturer's instructions (NEBNext kit, NEB, Frankfurt, Germany, <https://www.neb.com/products/e7420-nebnext-ultra-directional-rna-library-prep-kit-for-illumina>).

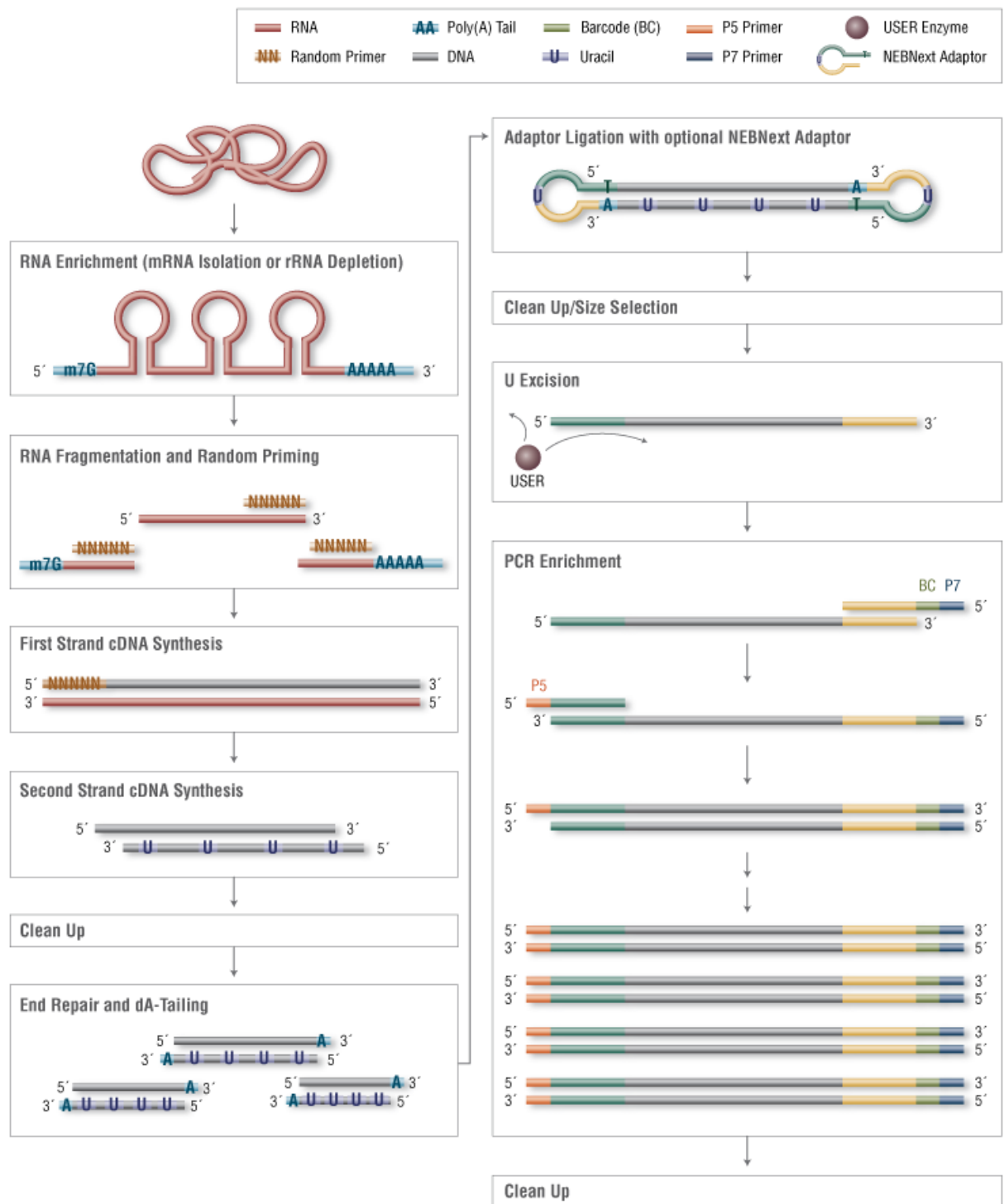


Figure 38. - Workflow for preparation of RNA library

6.4. Bioinformatics and statistics

Approximately 346 million reads were generated from *Arion vulgaris* transcriptomics study. These were filtered and trimmed by prinseq-lite using parameters trim_qual_right 30, min_qual_mean 30, min_len 70, trim_tail_right 6, trim_tail_left6, and rRNA sequences were removed using SortMeRNA 1.99 (274). Digital normalization (maximal coverage 30) and transcript assembly were performed using Trinity pipeline (275) (release 2013-02- 25) resulting in 136,406 assembled contigs with average length of 671.04 bp and N50 of 971 bp. CDS annotation was performed using the TransDecoder tool and PFAM version 27.0. Sequence similarities were obtained by blastx searches (cut-off E-value 1E-6) of the contigs and predicted CDS, respectively, against the NCBI non-redundant protein database (version July 2014). The assembled transcripts were further annotated with GO terms, PFAM protein domains and Enzyme Commission (EC) numbers using web platform FastAnnotator (166) with the default search parameters. 36,948 genes were assigned to the records present in NCBI non-redundant database and 22,868 of them to Gene Orthology terms. Searching against Pfam database identified 21,651 of entries to have at least one domain and 2,336 of genes were assigned with EC numbers. Duplicates and substrings were removed from the translated sequence dataset and resulting protein sequences were post-assembled using an in-house script to reduce redundancy. Final assembly containing 47,451 non-redundant protein sequences was used as an expression dataset for MS protein identification.

7. REFERENCES

1. Wilkins MR, Sanchez JC, Gooley AA, Appel RD, Humphery-Smith I, Hochstrasser DF, et al. Progress with proteome projects: why all proteins expressed by a genome should be identified and how to do it. *Biotechnol Genet Eng Rev.* 1996;13:19-50.
2. Dhingra V, Gupta M, Andacht T, Fu ZF. New frontiers in proteomics research: a perspective. *Int J Pharm.* 2005;299(1-2):1-18.
3. Anderson NL, Anderson NG. Proteome and proteomics: new technologies, new concepts, and new words. *Electrophoresis.* 1998;19(11):1853-61.
4. Neubauer G, Gottschalk A, Fabrizio P, Seraphin B, Luhrmann R, Mann M. Identification of the proteins of the yeast U1 small nuclear ribonucleoprotein complex by mass spectrometry. *Proc Natl Acad Sci U S A.* 1997;94(2):385-90.
5. Shevchenko A, Keller P, Scheiffele P, Mann M, Simons K. Identification of components of trans-Golgi network-derived transport vesicles and detergent-insoluble complexes by nanoelectrospray tandem mass spectrometry. *Electrophoresis.* 1997;18(14):2591-600.
6. Pandey A, Fernandez MM, Steen H, Blagoev B, Nielsen MM, Roche S, et al. Identification of a novel immunoreceptor tyrosine-based activation motif-containing molecule, STAM2, by mass spectrometry and its involvement in growth factor and cytokine receptor signaling pathways. *J Biol Chem.* 2000;275(49):38633-9.
7. Godovac-Zimmermann J, Brown LR. Perspectives for mass spectrometry and functional proteomics. *Mass Spectrom Rev.* 2001;20(1):1-57.
8. Alberts B. The cell as a collection of protein machines: Preparing the next generation of molecular biologists. *Cell.* 1998;92(3):291-4.
9. Celis JE, Gromov P. 2D protein electrophoresis: can it be perfected? *Curr Opin Biotech.* 1999;10(1):16-21.
10. Rabilloud T. Solubilization of proteins for electrophoretic analyses. *Electrophoresis.* 1996;17(5):813-29.
11. Bodzon-Kulakowska A, Bierczynska-Krzysik A, Dylag T, Drabik A, Suder P, Noga M, et al. Methods for samples preparation in proteomic research. *J Chromatogr B Analyt Technol Biomed Life Sci.* 2007;849(1-2):1-31.
12. Martinac A, Filipovic-Grcic J, Perissutti B, Voinovich D, Pavelic Z. Spray-dried chitosan/ethylcellulose microspheres for nasal drug delivery: swelling study and evaluation of in vitro drug release properties. *J Microencapsul.* 2005;22(5):549-61.
13. Pacheco LF, Carmona-Ribeiro AM. Effects of synthetic lipids on solubilization and colloid stability of hydrophobic drugs. *J Colloid Interface Sci.* 2003;258(1):146-54.
14. Niemczewski B. Observations of water cavitation intensity under practical ultrasonic cleaning conditions. *Ultrason Sonochem.* 2007;14(1):13-8.
15. Watkins LK, Bondarenko PV, Barbacci DC, Song S, Cockrill SL, Russell DH, et al. Fast C18 solid-phase desalting/delipidation of the human serum apolipoproteins for matrix-assisted laser desorption ionization and electrospray ionization mass spectrometric analysis. *J Chromatogr A.* 1999;840(2):183-93.
16. Joo WA, Lee DY, Kim CW. Development of an effective sample preparation method for the proteome analysis of body fluids using 2-D gel electrophoresis. *Biosci Biotechnol Biochem.* 2003;67(7):1574-7.
17. Rabilloud T. Solubilization of proteins for electrophoretic analyses. *Electrophoresis.* 1996;17(5):813-29.

18. Wilkins MR. Proteome research : new frontiers in functional genomics. Berlin ; New York: Springer; 1997. xviii, 243 p. p.
19. Schnack HG, van Haren NE, Brouwer RM, van Baal GC, Picchioni M, Weisbrod M, et al. Mapping reliability in multicenter MRI: voxel-based morphometry and cortical thickness. *Hum Brain Mapp.* 2010;31(12):1967-82.
20. Rath A, Glibowicka M, Nadeau VG, Chen G, Deber CM. Detergent binding explains anomalous SDS-PAGE migration of membrane proteins. *P Natl Acad Sci USA.* 2009;106(6):1760-5.
21. Dzieciatkowska M, Hill R, Hansen KC. GeLC-MS/MS analysis of complex protein mixtures. *Methods Mol Biol.* 2014;1156:53-66.
22. Zhang Y, Fonslow BR, Shan B, Baek MC, Yates JR, 3rd. Protein analysis by shotgun/bottom-up proteomics. *Chem Rev.* 2013;113(4):2343-94.
23. Piersma SR, Warmoes MO, de Wit M, de Reus I, Knol JC, Jimenez CR. Whole gel processing procedure for GeLC-MS/MS based proteomics. *Proteome Sci.* 2013;11.
24. Veronesi G, Aude-Garcia C, Kieffer I, Gallon T, Delangle P, Herlin-Boime N, et al. Exposure-dependent Ag⁺ release from silver nanoparticles and its complexation in AgS₂ sites in primary murine macrophages. *Nanoscale.* 2015;7(16):7323-30.
25. Zilberstein GV, Baskin EM, Bukshpan S, Korol LE. Parallel isoelectric focusing II. *Electrophoresis.* 2004;25(21-22):3643-51.
26. Gordon JA, Jencks WP. Relationship of Structure to Effectiveness of Denaturing Agents for Proteins. *Biochemistry-U.S.* 1963;2(1):47-&.
27. Rabilloud T, Adessi C, Giraudel A, Lunardi J. Improvement of the solubilization of proteins in two-dimensional electrophoresis with immobilized pH gradients. *Electrophoresis.* 1997;18(3-4):307-16.
28. Olivieri E, Herbert B, Righetti PG. The effect of protease inhibitors on the two-dimensional electrophoresis pattern of red blood cell membranes. *Electrophoresis.* 2001;22(3):560-5.
29. Herbert B. Advances in protein solubilisation for two-dimensional electrophoresis. *Electrophoresis.* 1999;20(4-5):660-3.
30. Molloy MP, Herbert BR, Walsh BJ, Tyler MI, Traini M, Sanchez JC, et al. Extraction of membrane proteins by differential solubilization for separation using two-dimensional gel electrophoresis. *Electrophoresis.* 1998;19(5):837-44.
31. Finnie C, Svensson B. Proteolysis during the isoelectric focusing step of two-dimensional gel electrophoresis may be a common problem. *Anal Biochem.* 2002;311(2):182-6.
32. Freeman WM, Hemby SE. Proteomics for protein expression profiling in neuroscience. *Neurochemical Research.* 2004;29(6):1065-81.
33. Rabilloud T, Lelong C. Two-dimensional gel electrophoresis in proteomics: A tutorial. *Journal of Proteomics.* 2011;74(10):1829-41.
34. Hecker M, Antelmann H, Buettner K, Bernhardt J. Gel-based proteomics of Gram-positive bacteria: A powerful tool to address physiological questions. *Proteomics.* 2008;8(23-24):4958-75.
35. Afjehi-Sadat L, Lubec G. Identification of enzymes and activity from two-dimensional gel electrophoresis. *Nature Protocols.* 2007;2(10):2318-24.
36. Pitarch A, Nombela C, Gil C. Prediction of the Clinical Outcome in Invasive Candidiasis Patients Based on Molecular Fingerprints of Five Anti-Candida Antibodies in Serum. *Mol Cell Proteomics.* 2011;10(1).
37. Sanchez-Campillo M, Bini L, Comanducci R, Raggiaschi R, Marzocchi B, Pallini V, et al. Identification of immunoreactive proteins of *Chlamydia trachomatis* by Western blot analysis of a two-dimensional electrophoresis map with patient sera. *Electrophoresis.* 1999;20(11):2269-79.
38. Ishigami A, Ohsawa T, Hiratsuka M, Taguchi H, Kobayashi S, Saito Y, et al. Abnormal accumulation of citrullinated proteins catalyzed by peptidylarginine

deiminase in hippocampal extracts from patients with Alzheimer's disease. *J Neurosci Res.* 2005;80(1):120-8.

39. Perluigi M, Poon HF, Hensley K, Pierce WM, Klein JB, Calabrese V, et al. Proteomic analysis of 4-hydroxy-2-nonenal-modified proteins in G93A-SOD1 transgenic mice - A model of familial amyotrophic lateral sclerosis. *Free Radical Bio Med.* 2005;38(7):960-8.

40. Sobel A, Tashjian AH. Distinct Patterns of Cytoplasmic Protein-Phosphorylation Related to Regulation of Synthesis and Release of Prolactin by Gh Cells. *Journal of Biological Chemistry.* 1983;258(17):312-24.

41. Bredemeyer AJ, Lewis RM, Malone JP, Davis AE, Gross J, Townsend RR, et al. A proteomic approach for the discovery of protease substrates. *P Natl Acad Sci USA.* 2004;101(32):11785-90.

42. Patterson SD. Proteomics: evolution of the technology. *Biotechniques.* 2003;35(3):440-4.

43. Anderson NL, Anderson NG. The human plasma proteome: history, character, and diagnostic prospects. *Mol Cell Proteomics.* 2002;1(11):845-67.

44. Sitnikov D, Chan D, Thibaudeau E, Pinard M, Hunter JM. Protein depletion from blood plasma using a volatile buffer. *J Chromatogr B.* 2006;832(1):41-6.

45. Govorukhina NI, Keizer-Gunnink A, van der Zee AGJ, de Jong S, de Bruijn HWA, Bischoff R. Sample preparation of human serum for the analysis of tumor markers - Comparison of different approaches for albumin and gamma-globulin depletion. *Journal of Chromatography A.* 2003;1009(1-2):171-8.

46. Gianazza E, Arnaud P. A general method for fractionation of plasma proteins. Dye-ligand affinity chromatography on immobilized Cibacron blue F3-GA. *Biochem J.* 1982;201(1):129-36.

47. Fountoulakis M, Juranville JF, Jiang L, Avila D, Roder D, Jakob P, et al. Depletion of the high-abundance plasma proteins. *Amino Acids.* 2004;27(3-4):249-59.

48. Gorg A, Obermaier C, Boguth G, Harder A, Scheibe B, Wildgruber R, et al. The current state of two-dimensional electrophoresis with immobilized pH gradients. *Electrophoresis.* 2000;21(6):1037-53.

49. Gorg A, Weiss W, Dunn MJ. Current two-dimensional electrophoresis technology for proteomics. *Proteomics.* 2004;4(12):3665-85.

50. Lee JE, Lee JY, Kim HR, Shin HY, Lin T, Jin DI. Proteomic Analysis of Bovine Pregnancy-specific Serum Proteins by 2D Fluorescence Difference Gel Electrophoresis. *Asian-Australas J Anim Sci.* 2015;28(6):788-95.

51. Stasyk T, Morandell S, Bakry R, Feuerstein I, Huck CW, Stecher G, et al. Quantitative detection of phosphoproteins by combination of two-dimensional difference gel electrophoresis and phosphospecific fluorescent staining. *Electrophoresis.* 2005;26(14):2850-4.

52. Budin I, Devaraj NK. Membrane assembly driven by a biomimetic coupling reaction. *J Am Chem Soc.* 2012;134(2):751-3.

53. Alberts B. *Molecular biology of the cell.* 4th ed. New York: Garland Science; 2002. xxxiv, 1548 p. p.

54. Krogh A, Larsson B, von Heijne G, Sonnhammer EL. Predicting transmembrane protein topology with a hidden Markov model: application to complete genomes. *J Mol Biol.* 2001;305(3):567-80.

55. Almen MS, Nordstrom KJ, Fredriksson R, Schioth HB. Mapping the human membrane proteome: a majority of the human membrane proteins can be classified according to function and evolutionary origin. *BMC Biol.* 2009;7:50.

56. von Heijne G. Membrane-protein topology. *Nat Rev Mol Cell Biol.* 2006;7(12):909-18.

57. Lai X. Reproducible method to enrich membrane proteins with high purity and high yield for an LC-MS/MS approach in quantitative membrane proteomics. *Electrophoresis*. 2013;34(6):809-17.
58. Zhang ZJ, Peck SC. Simplified enrichment of plasma membrane proteins for proteomic analyses in *Arabidopsis thaliana*. *Proteomics*. 2011;11(9):1780-8.
59. Zhang L, Xie J, Wang X, Liu X, Tang X, Cao R, et al. Proteomic analysis of mouse liver plasma membrane: use of differential extraction to enrich hydrophobic membrane proteins. *Proteomics*. 2005;5(17):4510-24.
60. Zhang LJ, Wang XE, Peng X, Wei YJ, Cao R, Liu Z, et al. Proteomic analysis of low-abundant integral plasma membrane proteins based on gels. *Cell Mol Life Sci*. 2006;63(15):1790-804.
61. Hunzinger C, Wozny W, Schwall GP, Poznanovic S, Stegmann W, Zengerling H, et al. Comparative profiling of the mammalian mitochondrial proteome: multiple aconitase-2 isoforms including N-formylkynurenine modifications as part of a protein biomarker signature for reactive oxidative species. *J Proteome Res*. 2006;5(3):625-33.
62. Reifschneider NH, Goto S, Nakamoto H, Takahashi R, Sugawa M, Dencher NA, et al. Defining the mitochondrial proteomes from five rat organs in a physiologically significant context using 2D blue-native/SDS-PAGE. *J Proteome Res*. 2006;5(5):1117-32.
63. Burre J, Beckhaus T, Schagger H, Corvey C, Hofmann S, Karas M, et al. Analysis of the synaptic vesicle proteome using three gel-based protein separation techniques. *Proteomics*. 2006;6(23):6250-62.
64. Coughenour HD, Spaulding RS, Thompson CM. The synaptic vesicle proteome: a comparative study in membrane protein identification. *Proteomics*. 2004;4(10):3141-55.
65. Hinton R, Dobrota M. Density gradient centrifugation. Amsterdam ; New York New York: North-Holland Pub. Co. ;
- sole distributors for the U.S.A. and Canada, Elsevier/North-Holland; 1976. 290 p. p.
66. Ferro M, Seigneurin-Berny D, Rolland N, Chapel A, Salvi D, Garin J, et al. Organic solvent extraction as a versatile procedure to identify hydrophobic chloroplast membrane proteins. *Electrophoresis*. 2000;21(16):3517-26.
67. Sivars U, Tjerneld F. Mechanisms of phase behaviour and protein partitioning in detergent/polymer aqueous two-phase systems for purification of integral membrane proteins. *Bba-Gen Subjects*. 2000;1474(2):133-46.
68. Everberg H, Sivars U, Emanuelsson C, Persson C, Englund AK, Haneskog L, et al. Protein pre-fractionation in detergent-polymer aqueous two-phase systems for facilitated proteomic studies of membrane proteins. *Journal of Chromatography A*. 2004;1029(1-2):113-24.
69. Collaboration CMS, Chatrchyan S, Khachatryan V, Sirunyan AM, Tumasyan A, Adam W, et al. Search for a standard-model-like Higgs boson with a mass in the range 145 to 1000 GeV at the LHC. *Eur Phys J C Part Fields*. 2013;73(6):2469.
70. Walter H, Sasakawa S. Partition of Proteins in Aqueous Dextran D-Polyethylene Glycol Peg Two-Phase Systems. *Federation Proceedings*. 1969;28(2):907-&.
71. Polgar L. The catalytic triad of serine peptidases. *Cellular and Molecular Life Sciences*. 2005;62(19-20):2161-72.
72. Oda K. New families of carboxyl peptidases: serine-carboxyl peptidases and glutamic peptidases. *J Biochem*. 2012;151(1):13-25.
73. Buller AR, Townsend CA. Intrinsic evolutionary constraints on protease structure, enzyme acylation, and the identity of the catalytic triad. *P Natl Acad Sci USA*. 2013;110(8):E653-E61.

74. Dodson G, Wlodawer A. Catalytic triads and their relatives. *Trends Biochem Sci.* 1998;23(9):347-52.
75. Medzihradszky KF. In-solution digestion of proteins for mass spectrometry. *Methods Enzymol.* 2005;405:50-65.
76. Gundry RL, White MY, Murray CI, Kane LA, Fu Q, Stanley BA, et al. Preparation of proteins and peptides for mass spectrometry analysis in a bottom-up proteomics workflow. *Curr Protoc Mol Biol.* 2009;Chapter 10:Unit10 25.
77. Rosenfeld J, Capdevielle J, Guillemot JC, Ferrara P. In-gel digestion of proteins for internal sequence analysis after one- or two-dimensional gel electrophoresis. *Anal Biochem.* 1992;203(1):173-9.
78. Shevchenko A, Loboda A, Ens W, Schraven B, Standing KG, Shevchenko A. Archived polyacrylamide gels as a resource for proteome characterization by mass spectrometry. *Electrophoresis.* 2001;22(6):1194-203.
79. Havlis J, Thomas H, Sebela M, Shevchenko A. Fast-response proteomics by accelerated in-gel digestion of proteins. *Anal Chem.* 2003;75(6):1300-6.
80. Shevchenko A, Wilm M, Vorm O, Mann M. Mass spectrometric sequencing of proteins silver-stained polyacrylamide gels. *Anal Chem.* 1996;68(5):850-8.
81. Speicher KD, Kolbas O, Harper S, Speicher DW. Systematic analysis of peptide recoveries from in-gel digestions for protein identifications in proteome studies. *J Biomol Tech.* 2000;11(2):74-86.
82. Shevchenko A, Tomas H, Havlis J, Olsen JV, Mann M. In-gel digestion for mass spectrometric characterization of proteins and proteomes. *Nat Protoc.* 2006;1(6):2856-60.
83. Wisniewski JR, Zougman A, Nagaraj N, Mann M. Universal sample preparation method for proteome analysis. *Nat Methods.* 2009;6(5):359-62.
84. Washburn MP, Wolters D, Yates JR, 3rd. Large-scale analysis of the yeast proteome by multidimensional protein identification technology. *Nat Biotechnol.* 2001;19(3):242-7.
85. Blonder J, Xiao Z, Veenstra TD. Proteomic profiling of differentiating osteoblasts. *Expert Rev Proteomics.* 2006;3(5):483-96.
86. Erde J, Loo RR, Loo JA. Enhanced FASP (eFASP) to increase proteome coverage and sample recovery for quantitative proteomic experiments. *J Proteome Res.* 2014;13(4):1885-95.
87. Maurer M, Muller AC, Parapatics K, Pickl WF, Wagner C, Rudashevskaya EL, et al. Comprehensive Comparative and Semiquantitative Proteome of a Very Low Number of Native and Matched Epstein-Barr-Virus-Transformed B Lymphocytes Infiltrating Human Melanoma. *Journal of Proteome Research.* 2014;13(6):2830-45.
88. Tanca A, Biosa G, Pagnozzi D, Addis MF, Uzzau S. Comparison of detergent-based sample preparation workflows for LTQ-Orbitrap analysis of the *Escherichia coli* proteome. *Proteomics.* 2013;13(17):2597-607.
89. Wisniewski JR, Zougman A, Mann M. Combination of FASP and StageTip-Based Fractionation Allows In-Depth Analysis of the Hippocampal Membrane Proteome. *Journal of Proteome Research.* 2009;8(12):5674-8.
90. Ostasiewicz P, Zielinska DF, Mann M, Wisniewski JR. Proteome, Phosphoproteome, and N-Glycoproteome Are Quantitatively Preserved in Formalin-Fixed Paraffin-Embedded Tissue and Analyzable by High-Resolution Mass Spectrometry. *Journal of Proteome Research.* 2010;9(7):3688-700.
91. Zhao Q, Fang F, Wu C, Wu Q, Liang Y, Liang Z, et al. imFASP: An integrated approach combining in-situ filter-aided sample pretreatment with microwave-assisted protein digestion for fast and efficient proteome sample preparation. *Anal Chim Acta.* 2016;912:58-64.

92. Molnar I, Horvath C. Reverse-Phase Chromatography of Polar Biological Substances - Separation of Catechol Compounds by High-Performance Liquid-Chromatography. *Clin Chem.* 1976;22(9):1497-502.
93. Cuatrecasas P. Affinity Chromatography, Principles and Applications. *Abstr Pap Am Chem Soc.* 1974:80-.
94. Zhang X, Fang A, Riley CP, Wang M, Regnier FE, Buck C. Multi-dimensional liquid chromatography in proteomics--a review. *Anal Chim Acta.* 2010;664(2):101-13.
95. Slebos RJ, Brock JW, Winters NF, Stuart SR, Martinez MA, Li M, et al. Evaluation of strong cation exchange versus isoelectric focusing of peptides for multidimensional liquid chromatography-tandem mass spectrometry. *J Proteome Res.* 2008;7(12):5286-94.
96. Gilar M, Olivova P, Daly AE, Gebler JC. Two-dimensional separation of peptides using RP-RP-HPLC system with different pH in first and second separation dimensions. *J Sep Sci.* 2005;28(14):1694-703.
97. Sparkman OD. *Mass spectrometry desk reference.* 1st ed. Pittsburgh, Pa.: Global View Pub.; 2000. xvi, 106 p. p.
98. Ho CS, Lam CW, Chan MH, Cheung RC, Law LK, Lit LC, et al. Electrospray ionisation mass spectrometry: principles and clinical applications. *Clin Biochem Rev.* 2003;24(1):3-12.
99. Pozniak BP, Cole RB. Current measurements within the electrospray emitter. *J Am Soc Mass Spectrom.* 2007;18(4):737-48.
100. Rapp U. *Mass Spectrometry and Proteomics: Principles and Applications. Handbook of Toxicogenomics: Strategies and Applications.* 2005:97-114.
101. Kingdon KH. A method for the neutralization of electron space charge by positive ionization at very low gas pressures. *Phys Rev.* 1923;21(4):408-18.
102. Hu Q, Noll RJ, Li H, Makarov A, Hardman M, Graham Cooks R. The Orbitrap: a new mass spectrometer. *J Mass Spectrom.* 2005;40(4):430-43.
103. McAlister GC, Berggren WT, Griep-Raming J, Horning S, Makarov A, Phanstiel D, et al. A proteomics grade electron transfer dissociation-enabled hybrid linear ion trap-orbitrap mass spectrometer. *Journal of Proteome Research.* 2008;7(8):3127-36.
104. Wells JM, McLuckey SA. Collision-induced dissociation (CID) of peptides and proteins. *Methods Enzymol.* 2005;402:148-85.
105. Cooper HJ, Hakansson K, Marshall AG. The role of electron capture dissociation in biomolecular analysis. *Mass Spectrom Rev.* 2005;24(2):201-22.
106. Syka JE, Coon JJ, Schroeder MJ, Shabanowitz J, Hunt DF. Peptide and protein sequence analysis by electron transfer dissociation mass spectrometry. *Proc Natl Acad Sci U S A.* 2004;101(26):9528-33.
107. Roepstorff P, Fohlman J. Proposal for a common nomenclature for sequence ions in mass spectra of peptides. *Biomed Mass Spectrom.* 1984;11(11):601.
108. Johnson RS, Martin SA, Biemann K, Stults JT, Watson JT. Novel fragmentation process of peptides by collision-induced decomposition in a tandem mass spectrometer: differentiation of leucine and isoleucine. *Anal Chem.* 1987;59(21):2621-5.
109. Schilling GD, Andrade FJ, Barnes JH, Sperline RP, Denton MB, Barinaga CJ, et al. Continuous simultaneous detection in mass spectrometry. *Anal Chem.* 2007;79(20):7662-8.
110. Kohler RE. The eighth day of creation: Makers of the revolution in biology. *Isis.* 1997;88(4):730-1.
111. Eddy SR. Non-coding RNA genes and the modern RNA world. *Nat Rev Genet.* 2001;2(12):919-29.
112. Gesteland RF, Cech TR, Atkins JF. *THE RNA WORLD THIRD EDITION The Nature of Modern RNA Suggests a Prebiotic RNA World Preface.* Cold Spring Harb Mon. 2006;43:ix-X.

113. Brantl S. Antisense-RNA regulation and RNA interference. *Biochim Biophys Acta*. 2002;1575(1-3):15-25.
114. Brouns SJ, Jore MM, Lundgren M, Westra ER, Slijkhuis RJ, Snijders AP, et al. Small CRISPR RNAs guide antiviral defense in prokaryotes. *Science*. 2008;321(5891):960-4.
115. Lin SL, Miller JD, Ying SY. Intronic microRNA (miRNA). *J Biomed Biotechnol*. 2006;2006(4):26818.
116. Ghildiyal M, Zamore PD. Small silencing RNAs: an expanding universe. *Nat Rev Genet*. 2009;10(2):94-108.
117. Ahmad K, Henikoff S. Epigenetic consequences of nucleosome dynamics. *Cell*. 2002;111(3):281-4.
118. Anderson L, Seilhamer J. A comparison of selected mRNA and protein abundances in human liver. *Electrophoresis*. 1997;18(3-4):533-7.
119. Chen G, Gharib TG, Huang CC, Taylor JM, Misek DE, Kardina SL, et al. Discordant protein and mRNA expression in lung adenocarcinomas. *Mol Cell Proteomics*. 2002;1(4):304-13.
120. Mootha VK, Bunkenborg J, Olsen JV, Hjerrild M, Wisniewski JR, Stahl E, et al. Integrated analysis of protein composition, tissue diversity, and gene regulation in mouse mitochondria. *Cell*. 2003;115(5):629-40.
121. Lichtinghagen R, Musholt PB, Lein M, Romer A, Rudolph B, Kristiansen G, et al. Different mRNA and protein expression of matrix metalloproteinases 2 and 9 and tissue inhibitor of metalloproteinases 1 in benign and malignant prostate tissue. *Eur Urol*. 2002;42(4):398-406.
122. Nie L, Wu G, Culley DE, Scholten JC, Zhang W. Integrative analysis of transcriptomic and proteomic data: challenges, solutions and applications. *Crit Rev Biotechnol*. 2007;27(2):63-75.
123. Wu X, Brewer G. The regulation of mRNA stability in mammalian cells: 2.0. *Gene*. 2012;500(1):10-21.
124. Yeat NC, Lin C, Sager M, Lin J. Cancer proteomics: developments in technology, clinical use and commercialization. *Expert Rev Proteomic*. 2015;12(4):391-405.
125. Papassotiropoulos A, Fountoulakis M, Dunckley T, Stephan DA, Reiman EM. Genetics, transcriptomics, and proteomics of Alzheimer's disease. *J Clin Psychiatry*. 2006;67(4):652-70.
126. Wang Y, Yan HL, Wang YP, Yang HL, Wei L, Xiao Y, et al. Proteomics and transcriptome analysis coupled with pharmacological test reveals the diversity of anti-thrombosis proteins from the medicinal insect, *Eupolyphaga sinensis*. *Insect Biochem Molec*. 2012;42(8):537-44.
127. Cheng HC, Qi RZ, Paudel H, Zhu HJ. Regulation and function of protein kinases and phosphatases. *Enzyme Res*. 2011;2011:794089.
128. Brotz-Oesterhelt H, Bandow JE, Labischinski H. Bacterial proteomics and its role in antibacterial drug discovery. *Mass Spectrom Rev*. 2005;24(4):549-65.
129. Lee JH, Lee DE, Lee BU, Kim HS. Global analyses of transcriptomes and proteomes of a parent strain and an L-threonine-overproducing mutant strain. *J Bacteriol*. 2003;185(18):5442-51.
130. Shimizu T, Shima K, Yoshino K, Yonezawa K, Shimizu T, Hayashi H. Proteome and transcriptome analysis of the virulence genes regulated by the VirR/VirS system in *Clostridium perfringens*. *J Bacteriol*. 2002;184(10):2587-94.
131. Fields S, Johnston M. Cell biology. Whither model organism research? *Science*. 2005;307(5717):1885-6.
132. Seok J, Warren HS, Cuenca AG, Mindrinos MN, Baker HV, Xu W, et al. Genomic responses in mouse models poorly mimic human inflammatory diseases. *Proc Natl Acad Sci U S A*. 2013;110(9):3507-12.

133. Jubb AW, Young RS, Hume DA, Bickmore WA. Enhancer Turnover Is Associated with a Divergent Transcriptional Response to Glucocorticoid in Mouse and Human Macrophages. *J Immunol.* 2016;196(2):813-22.
134. Yeoh EJ, Ross ME, Shurtleff SA, Williams WK, Patel D, Mahfouz R, et al. Classification, subtype discovery, and prediction of outcome in pediatric acute lymphoblastic leukemia by gene expression profiling. *Cancer Cell.* 2002;1(2):133-43.
135. Chang JC, Wooten EC, Tsimelzon A, Hilsenbeck SG, Gutierrez MC, Elledge R, et al. Gene expression profiling for the prediction of therapeutic response to docetaxel in patients with breast cancer. *Lancet.* 2003;362(9381):362-9.
136. Zhang J, Abrams Z, Parvin JD, Huang K. Integrative analysis of somatic mutations and transcriptomic data to functionally stratify breast cancer patients. *Bmc Genomics.* 2016;17 Suppl 7:513.
137. Li H, Zhu Y, Tang X, Li J, Li Y, Zhong Z, et al. Integrated analysis of transcriptome in cancer patient-derived xenografts. *PLoS One.* 2015;10(5):e0124780.
138. Bings NH. Plasma time-of-flight mass spectrometry as a detector for short transient signals in elemental analysis. *Anal Bioanal Chem.* 2005;382(4):887-90.
139. da Fonseca RR, Albrechtsen A, Themudo GE, Ramos-Madriral J, Sibbesen JA, Maretty L, et al. Next-generation biology: Sequencing and data analysis approaches for non-model organisms. *Mar Genomics.* 2016.
140. Ziemert N, Lechner A, Wietz M, Millan-Aguinaga N, Chavarria KL, Jensen PR. Diversity and evolution of secondary metabolism in the marine actinomycete genus *Salinispora*. *Proc Natl Acad Sci U S A.* 2014;111(12):E1130-9.
141. Faircloth BC, McCormack JE, Crawford NG, Harvey MG, Brumfield RT, Glenn TC. Ultraconserved elements anchor thousands of genetic markers spanning multiple evolutionary timescales. *Syst Biol.* 2012;61(5):717-26.
142. McCormack JE, Faircloth BC, Crawford NG, Gowaty PA, Brumfield RT, Glenn TC. Ultraconserved elements are novel phylogenomic markers that resolve placental mammal phylogeny when combined with species-tree analysis. *Genome Res.* 2012;22(4):746-54.
143. Todd EV, Black MA, Gemmell NJ. The power and promise of RNA-seq in ecology and evolution. *Mol Ecol.* 2016;25(6):1224-41.
144. da Fonseca RR, Albrechtsen A, Themudo GE, Ramos-Madriral J, Sibbesen JA, Maretty L, et al. Next-generation biology: Sequencing and data analysis approaches for non-model organisms. *Mar Genomics.* 2016;30:3-13.
145. Barker GM. The biology of terrestrial molluscs. Wallingford, Oxon, UK ; New York, NY, USA: CABI Pub.; 2001. xiv, 558 p. p.
146. Kałuski T, Kozłowski J, Kozłowska M. Evaluation of damage to seeds and seedlings of different winter wheat cultivars caused by *Deroceras reticulatum* (O.F. Müller, 1774) and *Arion lusitanicus Mabille*, 1868 in laboratory conditions. *Folia Malacologica.* 2005;13:189-95.
147. Cowie RH, Dillon RT, Robinson DG, Smith JW. Alien non-marine snails and slugs of priority quarantine importance in the United States: A preliminary risk assessment. *Am Malacol Bull.* 2009;27(1-2):113-32.
148. Gismervik K, Bruheim T, Rorvik LM, Haukeland S, Skaar I. Invasive slug populations (*Arion vulgaris*) as potential vectors for *Clostridium botulinum*. *Acta Vet Scand.* 2014;56.
149. Gismervik K, Aspholm M, Rorvik LM, Bruheim T, Andersen A, Skaar I. Invading slugs (*Arion vulgaris*) can be vectors for *Listeria monocytogenes*. *Journal of Applied Microbiology.* 2015;118(4):809-16.
150. Briner T, Frank T. The palatability of 78 wildflower strip plants to the slug *Arion lusitanicus*. *Ann Appl Biol.* 1998;133(1):123-33.
151. Lindstrom M, Myllykoski J, Sivela S, Korkeala H. *Clostridium botulinum* in Cattle and Dairy Products. *Crit Rev Food Sci.* 2010;50(4):281-304.

152. Lindstrom M, Korkeala H. Laboratory diagnostics of botulism. *Clin Microbiol Rev.* 2006;19(2):298-+.
153. Driehuis F. Silage and the safety and quality of dairy foods: a review. *Agr Food Sci.* 2013;22(1):16-34.
154. Ashburner M, Roote J. Culture of *Drosophila*: the laboratory setup. *CSH Protoc.* 2007;2007:pdb ip34.
155. Epstein CJ. The new dysmorphology: application of insights from basic developmental biology to the understanding of human birth defects. *Proc Natl Acad Sci U S A.* 1995;92(19):8566-73.
156. Campeau P, Mayson P, Parr H, Farrell J, Scriver C. Development of a web log (blog) for the online metabolic and molecular basis of inherited disorders: Facilitating discussions on inborn errors of metabolism. *J Inherit Metab Dis.* 2006;29:163-.
157. Vikkula M, Boon LM, Carraway KL, 3rd, Calvert JT, Diamonti AJ, Goumnerov B, et al. Vascular dysmorphogenesis caused by an activating mutation in the receptor tyrosine kinase TIE2. *Cell.* 1996;87(7):1181-90.
158. Marsh JL, Pallos J, Thompson LM. Fly models of Huntington's disease. *Hum Mol Genet.* 2003;12 Spec No 2:R187-93.
159. Valente EM, Abou-Sleiman PM, Caputo V, Muqit MM, Harvey K, Gispert S, et al. Hereditary early-onset Parkinson's disease caused by mutations in PINK1. *Science.* 2004;304(5674):1158-60.
160. Kopan R, Goate A. Aph-2/Nicastrin: an essential component of gamma-secretase and regulator of Notch signaling and Presenilin localization. *Neuron.* 2002;33(3):321-4.
161. Sutcliffe JE, Korenjak M, Brehm A. Tumour suppressors - a fly's perspective. *Eur J Cancer.* 2003;39(10):1355-62.
162. Bier E. *Drosophila*, the golden bug, emerges as a tool for human genetics. *Nat Rev Genet.* 2005;6(1):9-23.
163. Brunner E, Ahrens CH, Mohanty S, Baetschmann H, Loevenich S, Potthast F, et al. A high-quality catalog of the *Drosophila melanogaster* proteome. *Nat Biotechnol.* 2007;25(5):576-83.
164. Lee TR, Huang SH, Lee CC, Lee HY, Chan HT, Lin KS, et al. Proteome reference map of *Drosophila melanogaster* head. *Proteomics.* 2012;12(11):1875-8.
165. Grabherr MG, Haas BJ, Yassour M, Levin JZ, Thompson DA, Amit I, et al. Full-length transcriptome assembly from RNA-Seq data without a reference genome. *Nature Biotechnology.* 2011;29(7):644-U130.
166. Chen TW, Gan RCR, Wu TH, Huang PJ, Lee CY, Chen YYM, et al. FastAnnotator- an efficient transcript annotation web tool. *Bmc Genomics.* 2012;13.
167. Bulat T, Smidak R, Sialana FJ, Jung G, Rattei T, Bilban M, et al. Transcriptomic and Proteomic Analysis of *Arion vulgaris*--Proteins for Probably Successful Survival Strategies? *PLoS One.* 2016;11(3):e0150614.
168. Kaessmann H. Origins, evolution, and phenotypic impact of new genes. *Genome Res.* 2010;20(10):1313-26.
169. Cech TR, Steitz JA. The noncoding RNA revolution-trashing old rules to forge new ones. *Cell.* 2014;157(1):77-94.
170. Conesa A, Gotz S, Garcia-Gomez JM, Terol J, Talon M, Robles M. Blast2GO: a universal tool for annotation, visualization and analysis in functional genomics research. *Bioinformatics.* 2005;21(18):3674-6.
171. Krishna SS, Majumdar I, Grishin NV. Structural classification of zinc fingers: survey and summary. *Nucleic Acids Res.* 2003;31(2):532-50.
172. Cunha SR, Mohler PJ. Ankyrin protein networks in membrane formation and stabilization. *J Cell Mol Med.* 2009;13(11-12):4364-76.

173. Vizcaino JA, Deutsch EW, Wang R, Csordas A, Reisinger F, Rios D, et al. ProteomeXchange provides globally coordinated proteomics data submission and dissemination. *Nat Biotechnol.* 2014;32(3):223-6.
174. Voncken F, Gao F, Wadforth C, Harley M, Colasante C. The phosphoarginine energy-buffering system of trypanosoma brucei involves multiple arginine kinase isoforms with different subcellular locations. *PLoS One.* 2013;8(6):e65908.
175. Zelensky AN, Gready JE. The C-type lectin-like domain superfamily. *FEBS J.* 2005;272(24):6179-217.
176. Yuasa HJ, Furuta E, Nakamura A, Takagi T. Cloning and sequencing of three C-type lectins from body surface mucus of the land slug, *Incilaria fruhstorferi*. *Comp Biochem Physiol B Biochem Mol Biol.* 1998;119(3):479-84.
177. Pennington MW, Lanigan MD, Kalman K, Mahnir VM, Rauer H, McVaugh CT, et al. Role of disulfide bonds in the structure and potassium channel blocking activity of ShK toxin. *Biochemistry-U.S.* 1999;38(44):14549-58.
178. Rauer H, Pennington M, Cahalan M, Chandy KG. Structural conservation of the pores of calcium-activated and voltage-gated potassium channels determined by a sea anemone toxin. *J Biol Chem.* 1999;274(31):21885-92.
179. Beeton C, Pennington MW, Wulff H, Singh S, Nugent D, Crossley G, et al. Targeting effector memory T cells with a selective peptide inhibitor of Kv1.3 channels for therapy of autoimmune diseases. *Mol Pharmacol.* 2005;67(4):1369-81.
180. Yan L, Herrington J, Goldberg E, Dulski PM, Bugianesi RM, Slaughter RS, et al. Stichodactyla helianthus peptide, a pharmacological tool for studying Kv3.2 channels. *Mol Pharmacol.* 2005;67(5):1513-21.
181. Jovine L, Qi H, Williams Z, Litscher E, Wassarman PM. The ZP domain is a conserved module for polymerization of extracellular proteins. *Nat Cell Biol.* 2002;4(6):457-61.
182. Tordai H, Banyai L, Patthy L. The PAN module: the N-terminal domains of plasminogen and hepatocyte growth factor are homologous with the apple domains of the prekallikrein family and with a novel domain found in numerous nematode proteins. *FEBS Lett.* 1999;461(1-2):63-7.
183. Heaton MP, Clawson ML, Chitko-Mckown CG, Leymaster KA, Smith TP, Harhay GP, et al. Reduced lentivirus susceptibility in sheep with TMEM154 mutations. *PLoS Genet.* 2012;8(1):e1002467.
184. Lamriben L, Graham JB, Adams BM, Hebert DN. N-Glycan-based ER Molecular Chaperone and Protein Quality Control System: The Calnexin Binding Cycle. *Traffic.* 2016;17(4):308-26.
185. Zhang Q, Wang XQ, Jiang HS, Jia WM, Zhao XF, Wang JX. Calnexin functions in antibacterial immunity of *Marsupenaeus japonicus*. *Dev Comp Immunol.* 2014;46(2):356-63.
186. Sanchez JF, Lescar J, Chazalet V, Audfray A, Gagnon J, Alvarez R, et al. Biochemical and structural analysis of *Helix pomatia* agglutinin. A hexameric lectin with a novel fold. *J Biol Chem.* 2006;281(29):20171-80.
187. Rambaruth ND, Greenwell P, Dwek MV. The lectin *Helix pomatia* agglutinin recognizes O-GlcNAc containing glycoproteins in human breast cancer. *Glycobiology.* 2012;22(6):839-48.
188. Nickel W. The mystery of nonclassical protein secretion. A current view on cargo proteins and potential export routes. *Eur J Biochem.* 2003;270(10):2109-19.
189. Fiedler K, Simons K. A Putative Novel Class of Animal Lectins in the Secretory Pathway Homologous to Leguminous Lectins. *Cell.* 1994;77(5):625-6.
190. Takahashi-Nakaguchi A, Matsumoto Y, Yamamoto M, Iwabuchi K, Totsuka Y, Sugimura T, et al. Demonstration of cytotoxicity against wasps by pierisin-1: a possible defense factor in the cabbage white butterfly. *PLoS One.* 2013;8(4):e60539.

191. Varki A. Essentials of glycobiology. 2nd ed. Cold Spring Harbor, N.Y.: Cold Spring Harbor Laboratory Press; 2009. xxix, 784 p. p.
192. Schallus T, Jaekch C, Feher K, Palma AS, Liu Y, Simpson JC, et al. Malectin: a novel carbohydrate-binding protein of the endoplasmic reticulum and a candidate player in the early steps of protein N-glycosylation. *Mol Biol Cell*. 2008;19(8):3404-14.
193. Deprez P, Gautschi M, Helenius A. More than one glycan is needed for ER glucosidase II to allow entry of glycoproteins into the calnexin/calreticulin cycle. *Mol Cell*. 2005;19(2):183-95.
194. Rudd PM, Elliott T, Cresswell P, Wilson IA, Dwek RA. Glycosylation and the immune system. *Science*. 2001;291(5512):2370-6.
195. Gokudan S, Muta T, Tsuda R, Koori K, Kawahara T, Seki N, et al. Horseshoe crab acetyl group-recognizing lectins involved in innate immunity are structurally related to fibrinogen. *Proc Natl Acad Sci U S A*. 1999;96(18):10086-91.
196. Saito T, Kawabata S, Hirata M, Iwanaga S. A novel type of limulus lectin-L6. Purification, primary structure, and antibacterial activity. *J Biol Chem*. 1995;270(24):14493-9.
197. Schroder HC, Ushijima H, Krasko A, Gamulin V, Thakur NL, Diehl-Seifert B, et al. Emergence and disappearance of an immune molecule, an antimicrobial lectin, in basal metazoa. A tachylectin-related protein in the sponge *Suberites domuncula*. *J Biol Chem*. 2003;278(35):32810-7.
198. Gerlach D, Schlott B, Schmidt KH. Cloning and expression of a sialic acid-binding lectin from the snail *Cepaea hortensis*. *FEMS Immunol Med Microbiol*. 2004;40(3):215-21.
199. Gestal C, Pallavicini A, Venier P, Novoa B, Figueras A. MgC1q, a novel C1q-domain-containing protein involved in the immune response of *Mytilus galloprovincialis*. *Dev Comp Immunol*. 2010;34(9):926-34.
200. He X, Zhang Y, Yu F, Yu Z. A novel sialic acid binding lectin with anti-bacterial activity from the Hong Kong oyster (*Crassostrea hongkongensis*). *Fish Shellfish Immunol*. 2011;31(6):1247-50.
201. Xu T, Xie J, Li J, Luo M, Ye S, Wu X. Identification of expressed genes in cDNA library of hemocytes from the RLO-challenged oyster, *Crassostrea ariakensis* Gould with special functional implication of three complement-related fragments (CaC1q1, CaC1q2 and CaC3). *Fish Shellfish Immunol*. 2012;32(6):1106-16.
202. Yu YH, Huang HQ, Wang Y, Yu YC, Yuan SC, Huang SF, et al. A Novel C1q Family Member of Amphioxus Was Revealed to Have a Partial Function of Vertebrate C1q Molecule. *J Immunol*. 2008;181(10):7024-32.
203. Kong PF, Zhang HA, Wang LL, Zhou Z, Yang JL, Zhang Y, et al. AiC1qDC-1, a novel gC1q-domain-containing protein from bay scallop *Argopecten irradians* with fungi agglutinating activity. *Developmental and Comparative Immunology*. 2010;34(8):837-46.
204. Wang L, Wang L, Zhang H, Zhou Z, Siva VS, Song L. A C1q domain containing protein from scallop *Chlamys farreri* serving as pattern recognition receptor with heat-aggregated IgG binding activity. *PLoS One*. 2012;7(8):e43289.
205. Tahtouh M, Croq F, Vizioli J, Sautiere PE, Van Camp C, Salzet M, et al. Evidence for a novel chemotactic C1q domain-containing factor in the leech nerve cord. *Mol Immunol*. 2009;46(4):523-31.
206. Zhang H, Song L, Li C, Zhao J, Wang H, Qiu L, et al. A novel C1q-domain-containing protein from Zhikong scallop *Chlamys farreri* with lipopolysaccharide binding activity. *Fish Shellfish Immunol*. 2008;25(3):281-9.
207. Lehmann F, Tiralongo E, Tiralongo J. Sialic acid-specific lectins: occurrence, specificity and function. *Cell Mol Life Sci*. 2006;63(12):1331-54.
208. Iwanaga S, Lee BL. Recent advances in the innate immunity of invertebrate animals. *J Biochem Mol Biol*. 2005;38(2):128-50.

209. Tamplin ML, Fisher WS. Occurrence and characteristics of agglutination of *Vibrio cholerae* by serum from the eastern oyster, *Crassostrea virginica*. *Appl Environ Microbiol.* 1989;55(11):2882-7.
210. Tsuboi I, Matsukawa M, Sato N. Isolation and characterization of a sialic acid-specific lectin from hemolymph of the Southeast Asian horseshoe crab *Tachypleus gigas*. *Biosci Biotechnol Biochem.* 1993;57(8):1237-42.
211. Vasta GR, Cohen E. Sialic acid binding lectins in the serum of American spiders of the genus *Aphonopelma*. *Dev Comp Immunol.* 1984;8(3):515-22.
212. Grubhoffer L, Kovar V, Rudenko N. Tick lectins: structural and functional properties. *Parasitology.* 2004;129 Suppl:S113-25.
213. Peitsch MC, Tschopp J. Assembly of macromolecular pores by immune defense systems. *Curr Opin Cell Biol.* 1991;3(4):710-6.
214. Gonzalez MR, Bischofberger M, Pernot L, van der Goot FG, Freche B. Bacterial pore-forming toxins: the (w)hole story? *Cell Mol Life Sci.* 2008;65(3):493-507.
215. Dreon MS, Frassa MV, Ceolin M, Ituarte S, Qiu JW, Sun J, et al. Novel animal defenses against predation: a snail egg neurotoxin combining lectin and pore-forming chains that resembles plant defense and bacteria attack toxins. *PLoS One.* 2013;8(5):e63782.
216. Pringos E, Vignes M, Martinez J, Rolland V. Peptide neurotoxins that affect voltage-gated calcium channels: a close-up on omega-agatoxins. *Toxins (Basel).* 2011;3(1):17-42.
217. Daly NL, Craik DJ. Bioactive cystine knot proteins. *Curr Opin Chem Biol.* 2011;15(3):362-8.
218. Craik DJ, Daly NL, Waine C. The cystine knot motif in toxins and implications for drug design. *Toxicon.* 2001;39(1):43-60.
219. Wang H, Song L, Li C, Zhao J, Zhang H, Ni D, et al. Cloning and characterization of a novel C-type lectin from Zhikong scallop *Chlamys farreri*. *Mol Immunol.* 2007;44(5):722-31.
220. Sato S, St-Pierre C, Bhaumik P, Nieminen J. Galectins in innate immunity: dual functions of host soluble beta-galactoside-binding lectins as damage-associated molecular patterns (DAMPs) and as receptors for pathogen-associated molecular patterns (PAMPs). *Immunol Rev.* 2009;230:172-87.
221. Aradska J, Bulat T, Sialana FJ, Birner-Gruenberger R, Erich B, Lubec G. Gel-free mass spectrometry analysis of *Drosophila melanogaster* heads. *Proteomics.* 2015;15(19):3356-60.
222. Ishihama Y, Oda Y, Tabata T, Sato T, Nagasu T, Rappsilber J, et al. Exponentially modified protein abundance index (emPAI) for estimation of absolute protein amount in proteomics by the number of sequenced peptides per protein. *Mol Cell Proteomics.* 2005;4(9):1265-72.
223. Shinoda K, Tomita M, Ishihama Y. emPAI Calc--for the estimation of protein abundance from large-scale identification data by liquid chromatography-tandem mass spectrometry. *Bioinformatics.* 2010;26(4):576-7.
224. Sonnhammer EL, von Heijne G, Krogh A. A hidden Markov model for predicting transmembrane helices in protein sequences. *Proc Int Conf Intell Syst Mol Biol.* 1998;6:175-82.
225. Carbon S, Ireland A, Mungall CJ, Shu S, Marshall B, Lewis S, et al. AmiGO: online access to ontology and annotation data. *Bioinformatics.* 2009;25(2):288-9.
226. Dahanukar A, Hallem EA, Carlson JR. Insect chemoreception. *Curr Opin Neurobiol.* 2005;15(4):423-30.
227. Stocker RF. The organization of the chemosensory system in *Drosophila melanogaster*: a review. *Cell Tissue Res.* 1994;275(1):3-26.

228. Benton R, Vannice KS, Gomez-Diaz C, Vosshall LB. Variant Ionotropic Glutamate Receptors as Chemosensory Receptors in *Drosophila*. *Cell*. 2009;136(1):149-62.
229. Engelhardt S, Rochais F. G proteins - More than transducers of receptor-generated signals? *Circ Res*. 2007;100(8):1109-11.
230. Traynelis SF, Wollmuth LP, McBain CJ, Menniti FS, Vance KM, Ogden KK, et al. Glutamate Receptor Ion Channels: Structure, Regulation, and Function. *Pharmacol Rev*. 2010;62(3):405-96.
231. Yamamoto I. Nicotine to nicotinoids: 1962 to 1997. *Abstr Pap Am Chem S*. 1997;214:1-Agro.
232. Kandel ER. *Principles of neural science*. 5th ed. New York: McGraw-Hill; 2013. l, 1709 p. p.
233. Jankovic J. Parkinson's disease: clinical features and diagnosis. *J Neurol Neurosurg Psychiatry*. 2008;79(4):368-76.
234. Greenslit N. [Review of: Healy, David. *The Creation of psychopharmacology*. Cambridge, MA: Harvard University Press, 2002]. *Hist Psychiatry*. 2004;15(58 Pt 2):227-32.
235. Kimko HC, Cross JT, Abernethy DR. Pharmacokinetics and clinical effectiveness of methylphenidate. *Clin Pharmacokinet*. 1999;37(6):457-70.
236. Howes OD, Kapur S. The dopamine hypothesis of schizophrenia: version III--the final common pathway. *Schizophr Bull*. 2009;35(3):549-62.
237. Berridge CW, Devilbiss DM. Psychostimulants as Cognitive Enhancers: The Prefrontal Cortex, Catecholamines, and Attention-Deficit/Hyperactivity Disorder. *Biol Psychiat*. 2011;69(12):E101-E11.
238. Sharma AM, Pischon T, Hardt S, Kunz I, Luft FC. beta-Adrenergic receptor blockers and weight gain - A systematic analysis. *Hypertension*. 2001;37(2):250-4.
239. de Boer SF, Koolhaas JM. 5-HT_{1A} and 5-HT_{1B} receptor agonists and aggression: A pharmacological challenge of the serotonin deficiency hypothesis. *Eur J Pharmacol*. 2005;526(1-3):125-39.
240. Cunha RA, Ferre S, Vaugeois JM, Chen JF. Potential therapeutic interest of adenosine A_{2A} receptors in psychiatric disorders. *Curr Pharm Des*. 2008;14(15):1512-24.
241. Korogod SM, Kulagina IB. [Dynamical electrical states of heterogeneous populations of ion channels in the membranes of excitable cells]. *Fiziol Zh*. 2012;58(3):50-9.
242. Camerino DC, Tricarico D, Desaphy JF. Ion channel pharmacology. *Neurotherapeutics*. 2007;4(2):184-98.
243. Yao CK, Lin YQ, Ly CV, Ohyama T, Haueter CM, Moiseenkova-Bell VY, et al. A synaptic vesicle-associated Ca²⁺ channel promotes endocytosis and couples exocytosis to endocytosis. *Cell*. 2009;138(5):947-60.
244. Catterall WA. Voltage-gated calcium channels. *Cold Spring Harb Perspect Biol*. 2011;3(8):a003947.
245. Shieh CC, Coghlan M, Sullivan JP, Gopalakrishnan M. Potassium channels: molecular defects, diseases, and therapeutic opportunities. *Pharmacol Rev*. 2000;52(4):557-94.
246. Salzer I, Gantumur E, Yousuf A, Boehm S. Control of sensory neuron excitability by serotonin involves 5HT_{2C} receptors and Ca²⁺-activated chloride channels. *Neuropharmacology*. 2016;110(Pt A):277-86.
247. Fleck MW, Thomson JL, Hough LB. Histamine-gated ion channels in mammals? *Biochem Pharmacol*. 2012;83(9):1127-35.
248. Marban E, Yamagishi T, Tomaselli GF. Structure and function of voltage-gated sodium channels. *J Physiol-London*. 1998;508(3):647-57.

249. Frank CA, Wang X, Collins CA, Rodal AA, Yuan Q, Verstreken P, et al. New approaches for studying synaptic development, function, and plasticity using *Drosophila* as a model system. *J Neurosci*. 2013;33(45):17560-8.
250. Wright EM, Hirayama BA, Loo DF. Active sugar transport in health and disease. *J Intern Med*. 2007;261(1):32-43.
251. Danbolt NC. Glutamate uptake. *Prog Neurobiol*. 2001;65(1):1-105.
252. Jen JC, Wan J, Palos TP, Howard BD, Baloh RW. Mutation in the glutamate transporter EAAT1 causes episodic ataxia, hemiplegia, and seizures. *Neurology*. 2005;65(4):529-34.
253. de Vries B, Mamsa H, Stam AH, Wan J, Bakker SL, Vanmolkot KR, et al. Episodic ataxia associated with EAAT1 mutation C186S affecting glutamate reuptake. *Arch Neurol*. 2009;66(1):97-101.
254. Beart PM, O'Shea RD. Transporters for L-glutamate: An update on their molecular pharmacology and pathological involvement. *Brit J Pharmacol*. 2007;150(1):5-17.
255. Crowe JH, Crowe LM, Carpenter JF, Wistrom CA. Stabilization of Dry Phospholipid-Bilayers and Proteins by Sugars. *Biochem J*. 1987;242(1):1-10.
256. Crowe JH, Carpenter JF, Crowe LM. The role of vitrification in anhydrobiosis. *Annu Rev Physiol*. 1998;60:73-103.
257. Elbein AD, Pan YT, Pastuszak I, Carroll D. New insights on trehalose: a multifunctional molecule. *Glycobiology*. 2003;13(4):17r-27r.
258. Nishizaki Y, Yoshizane C, Toshimori Y, Arai N, Akamatsu S, Hanaya T, et al. Disaccharide-trehalose inhibits bone resorption in ovariectomized mice. *Nutr Res*. 2000;20(5):653-64.
259. Davies JE, Sarkar S, Rubinsztein DC. Trehalose reduces aggregate formation and delays pathology in a transgenic mouse model of oculopharyngeal muscular dystrophy. *Human Molecular Genetics*. 2006;15(1):23-31.
260. Sarkar S, Davies JE, Huang ZB, Tunnacliffe A, Rubinsztein DC. Trehalose, a novel mTOR-independent autophagy enhancer, accelerates the clearance of mutant huntingtin and alpha-synuclein. *Journal of Biological Chemistry*. 2007;282(8):5641-52.
261. Thorens B. Glucose transporters in the regulation of intestinal, renal, and liver glucose fluxes. *Am J Physiol-Gastr L*. 1996;270(4):G541-G53.
262. Haga T. Molecular properties of the high-affinity choline transporter CHT1. *J Biochem*. 2014;156(4):181-94.
263. Murai KK, Van Meyel DJ. Neuron-glia communication at synapses: Insights from vertebrates and invertebrates. *Neuroscientist*. 2007;13(6):657-66.
264. Jensen K, Sanchez-Garcia J, Williams C, Khare S, Mathur K, Graze RM, et al. Purification of transcripts and metabolites from *Drosophila* heads. *J Vis Exp*. 2013(73):e50245.
265. Luche S, Santoni V, Rabilloud T. Evaluation of nonionic and zwitterionic detergents as membrane protein solubilizers in two-dimensional electrophoresis. *Proteomics*. 2003;3(3):249-53.
266. Bae N, Lodi M, Pollak A, Lubec G. Mass spectrometrical analysis of bilin-binding protein from the wing of *Hebomoia glaucippe* (Linnaeus, 1758) (Lepidoptera: Pieridae). *Electrophoresis*. 2012;33(12):1787-94.
267. Ahmed KE, Chen WQ, John JP, Kang SU, Lubec G. Complete sequencing of the recombinant granulocyte-colony stimulating factor (filgrastim) and detection of biotinylation by mass spectrometry. *Amino Acids*. 2010;38(4):1043-9.
268. Chen WQ, Li L, Lubec G. Proteomic differences between white and brown adipocytes. *Amino Acids*. 2014;46(4):995-1008.
269. Kang SU, Heo S, Lubec G. Mass spectrometric analysis of GABAA receptor subtypes and phosphorylations from mouse hippocampus. *Proteomics*. 2011;11(11):2171-81.

270. Manza LL, Stamer SL, Ham AJ, Codreanu SG, Liebler DC. Sample preparation and digestion for proteomic analyses using spin filters. *Proteomics*. 2005;5(7):1742-5.
271. Rappsilber J, Mann M, Ishihama Y. Protocol for micro-purification, enrichment, pre-fractionation and storage of peptides for proteomics using StageTips. *Nat Protoc*. 2007;2(8):1896-906.
272. Gilar M, Olivova P, Daly AE, Gebler JC. Two-dimensional separation of peptides using RP-RP-HPLC system with different pH in first and second separation dimensions. *J Sep Sci*. 2005;28(14):1694-703.
273. Goldberg ME, ExpertBezancon N, Vuillard L, Rabilloud T. Non-detergent sulphobetaines: A new class of molecules that facilitate in vitro protein renaturation. *Fold Des*. 1996;1(1):21-7.
274. Kopylova E, Noe L, Touzet H. SortMeRNA: fast and accurate filtering of ribosomal RNAs in metatranscriptomic data. *Bioinformatics*. 2012;28(24):3211-7.
275. Grabherr MG, Haas BJ, Yassour M, Levin JZ, Thompson DA, Amit I, et al. Full-length transcriptome assembly from RNA-Seq data without a reference genome. *Nat Biotechnol*. 2011;29(7):644-52.

8. APPENDIX

8.1. Published papers

Bulat T, Smidak R, Sialana FJ, Jung G, Rattei T, Bilban M, et al. *Transcriptomic and Proteomic Analysis of Arion vulgaris--Proteins for Probably Successful Survival Strategies?* PLoS One. 2016;11(3):e0150614

Aradska J, Bulat T, Sialana FJ, Birner-Gruenberger R, Erich B, Lubec G. *Gel-free mass spectrometry analysis of Drosophila melanogaster heads.* Proteomics. 2015;15(19):3356-60

RESEARCH ARTICLE

Transcriptomic and Proteomic Analysis of *Arion vulgaris*—Proteins for Probably Successful Survival Strategies?

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Data Availability Statement: The sequence data has been submitted to the sequence read archive (SRA) database of GenBank (<http://www.ncbi.nlm.nih.gov/sra>) with the BioProject accession number PRJEB7891. The mass spectrometry proteomics data have been deposited to the ProteomeXchange Consortium (Vizcaino et al. 2014) via the PRIDE partner repository with the dataset identifier PXD002078 and 10.6019/PXD002078.

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Abstract

The Spanish slug, *Arion vulgaris*, is considered one of the hundred most invasive species in Central Europe. The immense and very successful adaptation and spreading of *A. vulgaris* suggest that it developed highly effective mechanisms to deal with infections and natural predators. Current transcriptomic and proteomic studies on gastropods have been restricted mainly to marine and freshwater gastropods. No transcriptomic or proteomic study on *A. vulgaris* has been carried out so far, and in the current study, the first transcriptomic database from adult specimen of *A. vulgaris* is reported. To facilitate and enable proteomics in this non-model organism, a mRNA-derived protein database was constructed for protein identification. A gel-based proteomic approach was used to obtain the first generation of a comprehensive slug mantle proteome. A total of 2128 proteins were unambiguously identified; 48 proteins represent novel proteins with no significant homology in NCBI non-redundant database. Combined transcriptomic and proteomic analysis revealed an extensive repertoire of novel proteins with a role in innate immunity including many associated pattern recognition, effector proteins and cytokine-like proteins. The number and diversity in gene families encoding lectins point to a complex defense system, probably as a result of adaptation to a pathogen-rich environment. These results are providing a fundamental and important resource for subsequent studies on molluscs as well as for putative antimicrobial compounds for drug discovery and biomedical applications.

Introduction

Arion vulgaris as the only terrestrial gastropod, is considered among the 100 worst invasive species in Europe. The spread of invasive species, including slugs, is an increasing problem worldwide with an important economical, ecological, health and social impact. The slug *A. vulgaris* is considered a serious pest, both in agriculture and private gardens. Moreover *A. vulgaris*

Competing Interests: The authors have declared that no competing interests exist.

may act as a vector for some pathogenic bacteria, like *Listeria monocytogenes* [1], *Clostridium botulinum* [2],[3] and host species for some parasites, as *Angiostrongylus vasorum* that can cause serious and potentially fatal disease in dogs and other canids [4]. *A. vulgaris* belongs to a species complex *Arion ater* that can only be distinguished by dissecting their reproductive organs.

Currently, transcriptomic studies on molluscs have been restricted to an ecological framework in the marine and fresh water gastropods. However, no transcriptomic or proteomic study has been carried out so far in *A. vulgaris*. Next-generation sequencing has offered a powerful and cost-efficient technique for the generation of transcriptomic datasets in non-model species using diverse platforms such as the Illumina HiSeq, Roche 454, Pacific Biosystems, and Applied Biosystems SOLiD. Several non-model organisms have been characterized by transcriptome sequencing [5–7], which has provided a better understanding of these species.

Since mRNA expression does not necessarily reflect changes at the protein level, complementary proteomic studies can yield more comprehensive biological insight. Mass spectrometry analysis allows large proteome surveys and comparative analysis, where quantities of hundreds or thousands of proteins can be compared between various conditions. However, the lack of a genomic resource for these animals is the major reasons for restricting proteomic applications.

In the current study, the Illumina HiSeq 2000 platform has been used to generate an *A. vulgaris* transcriptome-based protein database. The transcriptome data generated in this study provide a first comprehensive and valuable genomic resource for future research on this slug. With the objective to obtain insight into the slug proteome gel-based proteomic, analysis from the *A. vulgaris* mantle was performed.

Methods

Collection of *Arion vulgaris*

A. vulgaris specimens were collected in May 2014 in a location (48° 19' 40" N, 16° 12' 34" E) in Woerdern, Austria. A total of 15 specimens ranging from 7 to 8 g were collected. Species identification was carried out with the assistance of the Zoological Department of Museum of Natural History in Vienna (H.S.). According to national law no permission is required to collect and use invertebrates.

Transcriptome sequencing and assembly

Isolation of RNA was performed with the RNeasy kit (Qiagen, Hilden, Germany). Following removal of the gut, two specimens of *Arion vulgaris* were immersed in liquid nitrogen and pulverized with the help of a mortar and pestle. Pooled total RNA was subjected to RNA-Seq following the mRNA sequencing protocol provided by New England Biolabs (NEBNext kit, NEB, Frankfurt, Germany). The first step in the workflow involved purifying the poly(A)-containing mRNA molecules using two rounds of poly(T)-oligo-attached magnetic beads. After purification, mRNAs were fragmented into small pieces using divalent cations under elevated temperature. The cleaved RNA fragments were copied into first-strand complementary DNA (cDNA) using reverse transcriptase and random primers. This was followed by second-strand cDNA synthesis using DNA polymerase I and RNase H (NEB, Frankfurt, Germany). These cDNA fragments subsequently went through an end-repair process, the addition of a single A base and ligation of the adaptors. The products were then purified and enriched with 15 cycles of PCR (with a size distribution of ~200–250 bp) to create the final cDNA library. Finally, the adaptor-ligated DNA was sequenced for 100 cycles on a HiSeq2000 sequencing system (Illumina, San Diego, USA) in a paired-end manner according to the manufacturer's

instructions. The generated reads were filtered and trimmed by prinseq-lite using parameters trim_qual_right 30, min_qual_mean 30, min_len 70, trim_tail_right 6, trim_tail_left 6, and rRNA sequences were removed using SortMeRNA 1.99 [8]. Digital normalization (maximal coverage 30) and transcript assembly were performed using Trinity pipeline (release 2013-02-25, [9]).

Transcriptome annotation

Protein-coding sequences were identified in the assembly using the TransDecoder tool and PFAM version 27.0. Sequence similarities were obtained by blastx searches (cut-off E-value 1E-6) of the contigs and predicted CDS, respectively, against the NCBI non-redundant protein database (version July 2014). The assembled transcripts were further annotated with GO terms, PFAM protein domains and Enzyme Commission (EC) numbers using web platform FastAnnotator [10] with the default search parameters. Duplicates and substrings were removed from the translated sequence dataset and resulting protein sequences were post-assembled using an in-house script to reduce redundancy. Final assembly containing 47,451 non-redundant protein sequences were used as an expression dataset for MS protein identification.

Protein extraction

Individuals of *A. vulgaris* were frozen immediately after harvest in liquid nitrogen and stored at -80°. Mantles were dissected directly before protein extraction procedure and homogenized in urea buffer (7 M urea, 2 M thiourea, 4% CHAPS, 2 M Tris (pH 8.8), 0.5% Carrier (3–11 pH), 1% DTT, 1 mM EDTA, 1 mM PMSF, Protease Inhibitor C). Protein samples were cleaned using a 2-DE clean up kit (BioRad, Hercules, CA, USA) and protein concentration was estimated by the Pierce 660 kit (ThermoScientific, Rockford, USA).

Two dimensional gel electrophoresis (2-DE) and nano-LC-ESI-MS/MS analysis

2-DE was performed as reported previously with minor modifications [11–13]. 750 µg of proteins were loaded on 18 cm 3–11 linear IPG strips (GE Healthcare). IEF was performed in a IPGphor electrophoretic system (GE Healthcare, Uppsala, Sweden) using a protocol with gradually increasing voltage from 200 to 8,000 V at 4 V/min. Prior to second dimension electrophoresis, IPG strips were incubated in 10 ml of equilibration buffer (50 mM Tris-HCl, pH 8.8, 6 M urea, 30% glycerol, 2% SDS and 0.002% bromophenol blue) containing 1% DTT for 15 min with gentle shaking followed by incubation in 10 mL of equilibration buffer with 4% iodoacetamide. The second-dimensional separation was performed on 10–16% gradient SDS-PAGE gels. After fixation for 4 h in 50% methanol and 10% acetic acid, gels were stained overnight with the colloidal coomassie blue (Novex life technologies, Invitrogen, CA). Molecular masses were determined by comparison with precision protein standard markers (#1610373, BioRad, Hercules, CA, USA) spanning the 10 to 250 kDa molecular weight range. All spots were excised from 2-DE gels and digested with trypsin and/or chymotrypsin. The gel pieces were cut into small pieces and washed with 50% ACN in 10mM ammonium bicarbonate for 20 min with vortexing and then 20 min with 10mM ammonium bicarbonate. These two steps were repeated until the gel was completely destained. 100% ACN was added for 10min and gel pieces were dried completely using a SpeedVac concentrator for 30 min at 30°C. Cysteine residues were reduced with 10 mM DTT at 56°C for 30 min and alkylated with 55 mM iodoacetamide for 45 min at 22°C. After washing with 10 mM ammonium bicarbonate and dehydration with 100% ACN, proteins were digested with 12.5 ng/µL trypsin (Promega,

Mannheim, Germany) solution buffered in 10 mM ammonium bicarbonate for 16 h (overnight) at 37°C. The supernatant was transferred to new LoBind 0.5 mL tubes and peptides were extracted with 1% formic acid and then with 15% ACN/0.1% formic acid [14]. 20 μ L of extracted peptides were analyzed by nano-LC-ESI-(CID/ETD)-MS/MS essentially as described previously [14]. The HPLC used was an Ultimate 3000 system (Dionex Corporation, Sunnyvale, CA) equipped with a PepMap100 C-18 trap column (300 μ m \times 5 mm) and PepMap100 C-18 analytical column (75 μ m \times 250 mm). The gradient was (A = 0.1% formic acid in water, B = 0.08% formic acid in ACN) 4–30% B from 0 to 105 min, 80% B from 105 to 110 min, 4% B from 110 to 125 min. The Amazon speed ETD (Bruker Daltonics, Bremen, Germany) was used to record peptide spectra over the mass range of m/z 400–1,400, and MS/MS spectra in information-dependent data acquisition over the mass range of m/z 100–2,800. Repeatedly, MS spectra were recorded followed by three data-dependent CID MS/MS spectra. An active exclusion of 0.4 min after two spectra was used to detect low abundant peptides. The voltage between ion spray tip and spray shield was set to 1,400 V. Drying nitrogen gas was heated to 150°C and the flow rate was 3 L/min. The collision energy was set automatically according to the mass and charge state of the peptides chosen for fragmentation. Multiple charged peptides were chosen for MS/MS experiments due to their good fragmentation characteristics. MS/MS spectra were interpreted and peak lists were generated by Data Analysis 4.1 (Bruker Daltonics, Bremen, Germany).

One dimensional (1-D) SDS-PAGE and LC-MS/MS

Although 2-DE followed by LC-MS/MS is a proven method for protein separation and identification, it suffers from poor sensitivity, poor representation of very acidic or basic proteins and low solubility of hydrophobic proteins. Therefore, we also separated proteins by 1-D SDS-PAGE coupled with high resolution LC-MS/MS. 50 μ g of extracted proteins were mixed with Laemmli buffer (150 mM Tris HCl pH 6.8, 300 mM DTT, 6% SDS, 0.3% bromophenol blue, 30% glycerol) at a 1:3 volume ratio. Proteins in each sample were separated by 10% SDS-PAGE and stained by Blue silver [15]. After destaining with Milli-Q water, gels were cut into 12 slices with proteins of different mass. Proteins underwent in-gel trypsin digestion as above and were analyzed using a LTQ-Orbitrap Velos (ThermoFisher Scientific, Waltham, MA, USA) coupled with nano-LC (Dionex Ultimate 3000) as described previously [16].

Database search

Data generated from Amazon speed ETD for each of the 2-DE protein spots were analyzed by searching the mRNA-derived *Arion vulgaris* database (47,451 sequences) including 115 commonly observed contaminants with Mascot Search engine (version 2.4) using the Mascot Daemon interface (Matrix Science, London, UK). Detailed search criteria were used as follows; enzyme: trypsin or chymotrypsin with a maximum of two missing cleavage sites; search mode: MS/MS ion search with decoy database search included; fixed modification: carbamidomethylation (C); variable modification: oxidation (M); search mode: MS/MS ion search with decoy database search included; peptide mass tolerance 0.35 Da; MS/MS mass tolerance \pm 0.35 Da.

For the mass spectrometry data generated from the LTQ Orbitrap Velos, the MS raw files were first processed with Proteome Discoverer 1.4 (ThermoFisher Scientific, IL, USA) to generate separate Mascot generic files. The database search was performed using Mascot version 2.4 (MatrixScience, London, UK) against the combined database as mentioned above. The search criteria were 10 ppm for precursor and 0.5 Da for fragments; search mode: MS/MS ion search with decoy database search included; fixed modification: carbamidomethyl (cysteine); variable modification: oxidation (methionine).

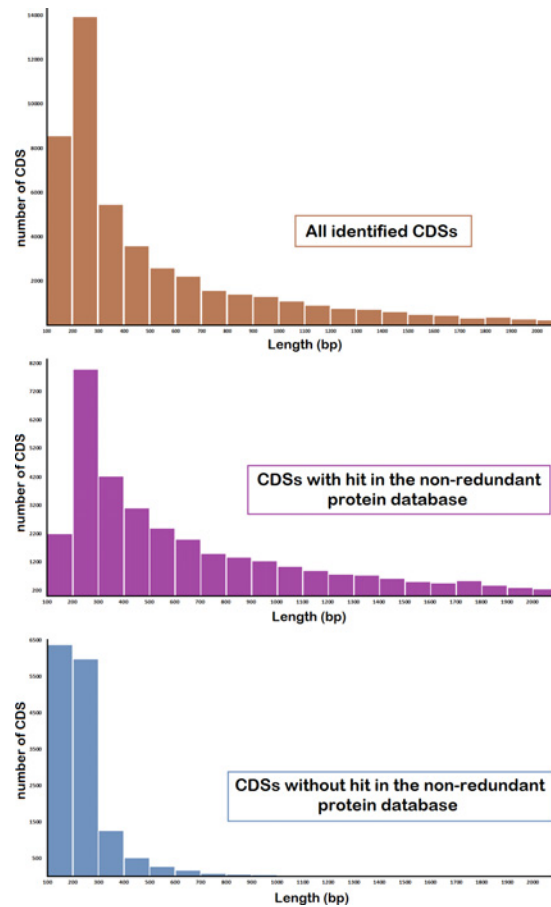


Fig 1. Protein-coding sequences (CDS) distribution showing the majority of sequences in the range from 200 to 300 bp.

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Peptide identifications were filtered to a < 1% false discovery rate (FDR) using the target-decoy strategy [17]. Acceptance parameter for protein identification was a minimum of one unique peptide and two distinct peptides.

Results

Transcriptome of whole body

The slug-specific database was constructed using a RNAseq and *de novo* assembly strategy. A cDNA library from the adult slug was generated and using Illumina HiSeq 2000 technology an approximate number of 339 millions of paired end reads was obtained. *De novo* transcript assembly performed by Trinity pipeline [9] resulted in 136,406 contigs with average lengths of 671.04 bp and N50 of 971 bp, meaning that 50% of the assembled sequences are 971 bp long or longer. 53,523 protein-coding sequences (CDS) were identified using the TransDecoder tool from the Trinity package with PFAM version 27.0. Fig 1 shows the distribution of CDS annotated contig lengths. All CDS annotated contigs of the final assembly were subjected to blastx analysis against the NCBI non-redundant protein database (nr) using the web platform Fastanotator [10] with a cut-off value of 1E-6. Duplicates and substrings were removed from the translated sequence dataset and resulting protein sequences were post-assembled using an in-house script to reduce redundancy. The final assembly contains 47,451 non-redundant protein

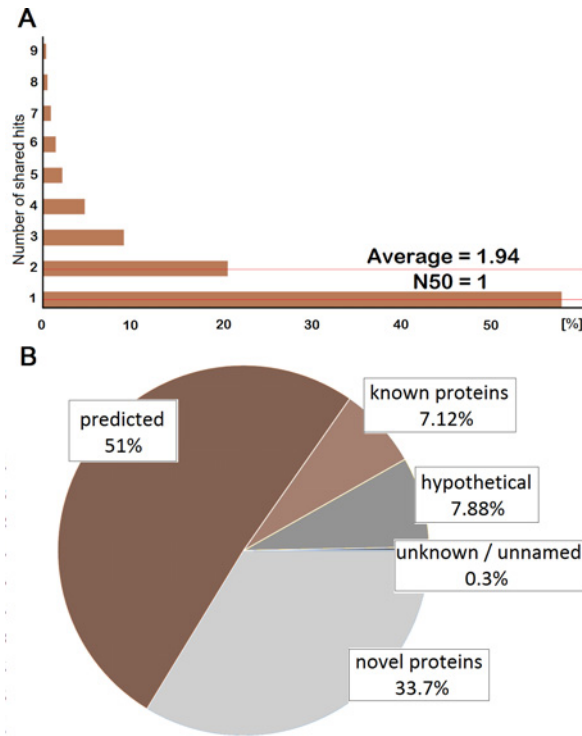


Fig 2. Characteristics of sequences from *A. vulgaris* transcriptome. (a) Distribution of shared top-hits. (b) Classification of top-hits in the current transcriptome.

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sequences. On average 1.94 sequences of the final assembly share the same hit against the NCBI nr database (Fig 2A), due to sequencing errors and/or potential isoforms. As a result, 31,463 sequences (66.3%) were assigned to at least one protein and 15,988 (33.7%) transcripts did not have a significant hit in the NCBI nr database probably because of the lack of molecular data of mollusc species. Among 31,463 matches, only 3,380 (10.7%) were known proteins, 24,217 are predicted (77%), 3,739 are hypothetical (11.9%), and 127 are either uncharacterized, unknown or unnamed proteins (0.4%) (Fig 2B). Sequences without significant similarity to sequences present in the nr database are being referred as putative novel genes. These include taxonomically constricted genes derived from ancestral genes or appeared *de novo* from non-coding sequences [18]. The high number of transcripts lacking detectable homology with protein sequences described in the database might be also due to the high number of short transcripts (147–300 bp) in the final assembly. Of the 47,451 transcripts in the final assembly, 46.5% (22,055 transcripts) are shorter than 300 bp (Fig 1). It has been shown that non-coding RNAs [19] as well as small open reading frames are potential sources of sequences lacking detectable homology with protein sequences in nr databases. A high number of putative novel genes was identified but additional genetic as well as proteomic studies are needed to annotate them correctly and this was one of the aims of the current proteomic study described below.

39% (18,561) of all sequences were functionally annotated with Gene Ontology (GO) terms using BLAST2GO tool of Fastannotator [20]. The results were summarized to the categories “biological process” (33.9%), “molecular function” (28.2%), and “cellular component” (34.8%). Within the biological process classification, metabolic processes (66%), biological regulation (43%) and response to stimulus (29%) were the most representative (Fig 3). In the molecular function category, the highest percentage of GO terms corresponded to “binding”

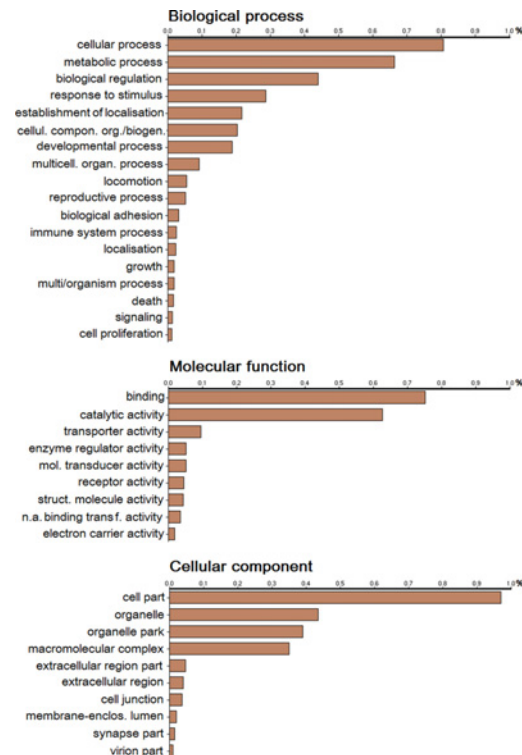


Fig 3. Gene Ontology (GO) analyses of the *A. vulgaris* transcriptome.

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(75%), “catalytic activity” (62%) and “transporter activity” (9.8%) (Fig 3). For the cellular component classification, the cluster sizes of “organelle/organelle part”, “macromolecular complex” and “extracellular region” were relatively large (Fig 3). Domain analysis against the Pfam database identified 16,994 of entries to have at least one domain of which 471 sequences without blastx hit were further annotated. The zinc-finger double domain zf-H2C2_2 (PF13465) was the most abundant domain followed by domain of unknown function DUF4200 (PF13863) and ankyrin repeat Ank_5 (PF13857) (S1 Table). Ankyrin domains are among the most common structural motifs in known proteins. 1,796 of transcripts were assigned with EC (Enzyme Commission) numbers.

We also analyzed taxonomic distribution of the best-match species of the annotable slug sequences. The highest proportion of the best blastx hits was found for *Aplysia californica* (65%), *Crassostea gigas* (8.7%) and *Capitella teleta* (2.5%) (Fig 4). Despite the limited number of molluscan sequences in the public databases, more than 80% of the sequences best matched to molluscan species. Assignment of transcripts also to chordate, plants, fungi, bacteria and viruses was probably due to more functional data on these species as well as contamination from environmental organisms during sample preparation has to be also considered.

Proteome of slug mantle

With the objective to gain insight into the slug proteome gel-based proteomic analysis from the *A. vulgaris* mantle was performed. In total, 356 spots corresponding to 833 unique proteins were clearly resolved in 2-DE reproducible gels (S2 Table). Fig 5 shows the representative 2-DE gel covering pH 3–10. Most of the spots were located between pH 5 to 9 over a broad range of MW (10–250 kDa). Among these proteins, 814 were assigned to protein sequence with significant hit in NCBI nr database and 19 represent a proteins with no significant homology in nr

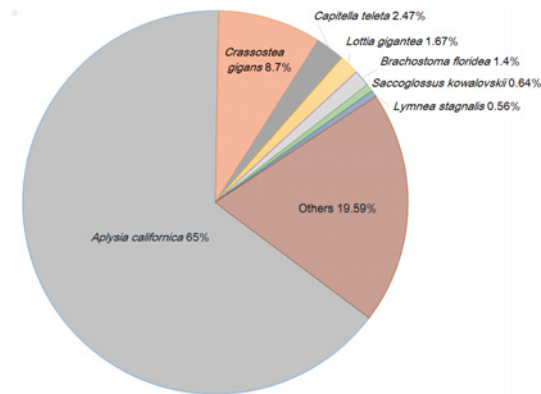


Fig 4. Species distribution of best blastx hits against the nr database.

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database. Several putative novel proteins have been identified in several spots representing isoforms with different posttranslational modifications deduced from different position in the gel. Because of the limitation of 2-DE for very small and very large proteins, alkaline proteins and hydrophobic proteins, we also analyzed the protein samples by 1-D SDS-PAGE coupled with LC-MS/MS. 2011 proteins were identified in two biological replicates from 1-D SDS-PAGE (S3 Table). Combining the results from 1-D and 2-D gel separation methods, a total of 2129 proteins were identified with minimum of one unique peptide and two distinct peptides (S4 Table). The overall false-positive assignment was estimated around 1% by a target-decoy search strategy suggesting high quality of our dataset. 1770 identified proteins (83.1%) had matches to Gene Ontology (GO) term annotations and were categorized to molecular function (1647 proteins), cellular component (1337 proteins), and biological process (1596 proteins). Of these, 23.7% have functions that are associated with cytoskeletal protein binding. The mantle, made up of muscle and skin, hence many of the cytoskeletal and muscle-related proteins were observed. The highest number of identified peptides in the combined proteomic approach were observed for paramyosine, myosine, actin and arginine kinase. Arginine kinase belongs to a class of kinases that play a crucial role in invertebrates in the maintenance of ATP levels by the phosphorylation of phosphagens which then serve as a high energy source for rapid ATP replenishment [21]. The GO terms with the largest number of assigned sequences in the biological process category were small molecule metabolic process, catabolic process, biosynthetic process and response to chemical stimulus. 1882 (88.3%) had at least one PFAM domain match and 398 (18.7%) an EC number. The RNA binding motifs RRM_6 and RRM_1 (PF14259 and PF00076) were the most abundant domains followed by Tropomyosin_1 domain (PF12718) and EF hand domains (PF13833, PF13405). (S4 Table).

Combined transcriptomic and proteomic analysis revealed the presence of a high number of sequences sharing carbohydrate-recognition domains, like lectin domains, proteins with C1q domain and toxin-like proteins were present as well (S5 Table).

Discussion

The combined transcriptomic and proteomic approach allowed the identification of a great number of new transcripts and proteins related to metabolic, functional and cellular components. These transcriptomics and proteomics data are by far the most comprehensive among terrestrial gastropods. Moreover, one may speculate that innate immune response-related and defense elements identified in this study may contribute to uncover the success of *A. vulgaris* and indeed, *A. vulgaris* is spreading Europe-wide and is well protected against tentative

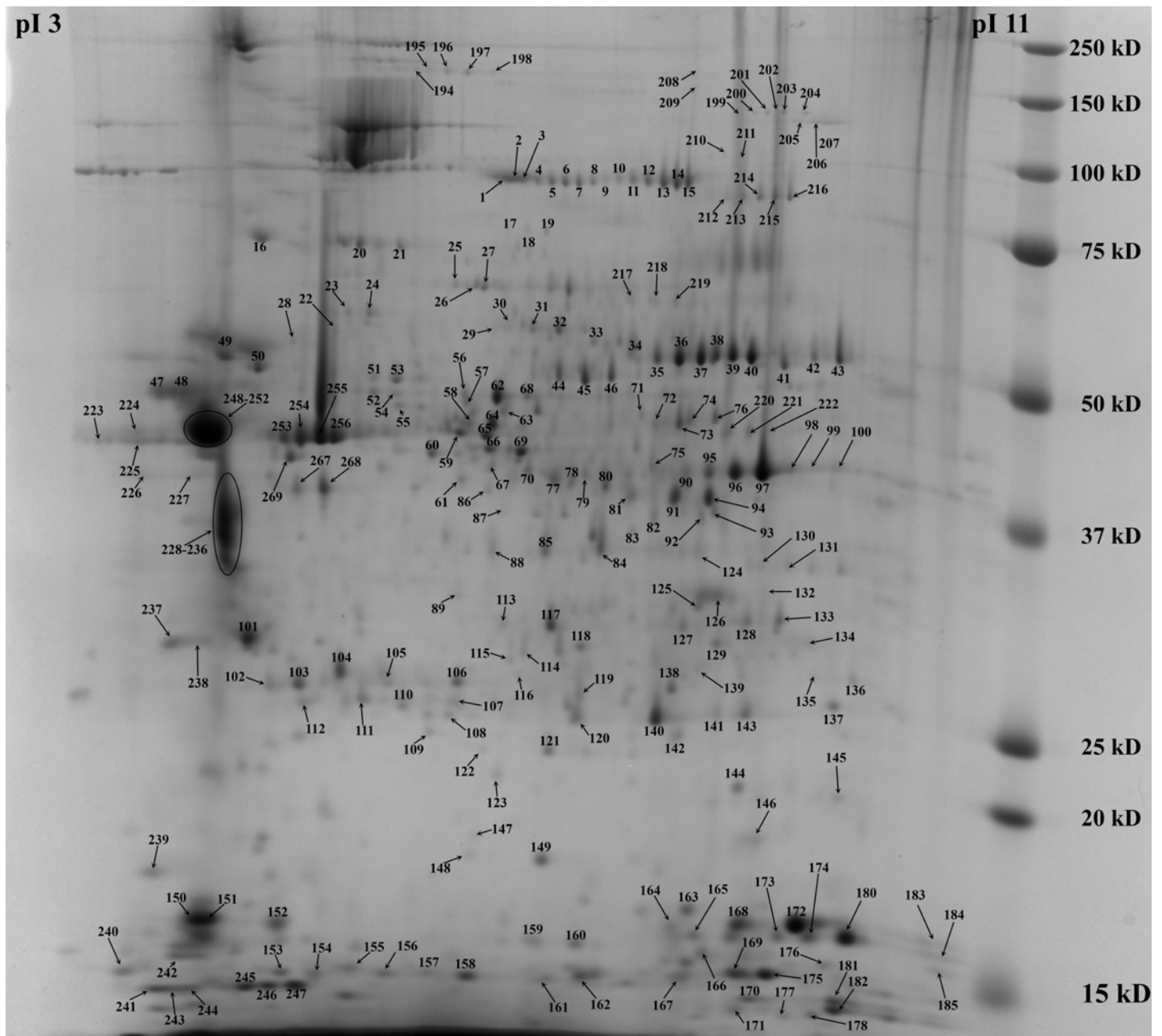


Fig 5. Representative 2-DE gel image from slug mantle showing protein spots selected for mass spectrometry analysis. The identified proteins are listed in [S2 Table](#).

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predators and pathogens [22–24]. 2129 *A. vulgaris* mantle proteins were identified through this study, markedly expanding the list of known proteins in terrestrial gastropods, demonstrating the powerfulness of gel-based approach (2-DE and 1-D SDS-PAGE) combined with mass spectrometry identification (LC-ESI-MS/MS). Of the two protein identification approaches used in this study, the 1-D approach has shown markedly higher sensitivity as demonstrated by the significantly higher number of detected proteins. The 2-DE method is useful in separating protein isoforms with different isoelectric point and protein mobility [25].

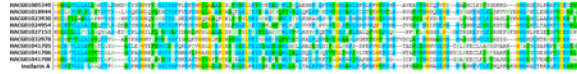


Fig 6. Domain architecture of C-lectin-like molecules found in the transcriptome of *A. vulgaris*.

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Slugs belonging to the one of the most successful phyla, Mollusca, lack clear evidence of adaptive immunity. They contend with many pathogens, including bacteria, fungi, viruses and several lineages of specialized eukaryotic parasites, and how they without adaptive immunity effectively and sufficiently defend themselves, is not well understood. The innate immune system in gastropoda is provided by physical barriers (e.g., shell, skin and epithelium), an evolutionary archaic mechanism of molecular "self-non-self" recognition, as well as by a variety of defense-related factors. The soft, moist slug body protected by a ciliated, mucus-producing epithelium that provides an initial physical barrier to colonization by the pathogens, plays a crucial role in host defense. The diversification of immune systems during evolution involves the expansion of particular gene families in given phyla. Analysis of transcriptomic data from *A. vulgaris* shows a comprehensive repertoire of genes related to innate immunity including many associated pattern recognition, effector proteins and cytokine-like proteins, such as lectins, complement-like proteins, peptidoglycan-recognition proteins (PGRPs), lipopolysaccharide and β 1,3-glucan-binding proteins, fibrinogen-related proteins (FREPs), pore-forming membrane attack/perforin (MACPF) domain proteins and toxin-like proteins that indicates an extraordinary complexity of immune system with a high degree of pathogen specificity and immune-priming.

Lectin-like proteins

Lectins play an important role in „self-non-self“ recognition and clearance of invaders in gastropods. The slug repertoire of lectin-like proteins bearing conserved carbohydrate-recognition domains (CRDs) is highly diversified, including C-type lectin family, galectin, malectin, H-type lectins, L-type lectins, Ricin-B lectins and calnexin. The diversity of CRDs in lectins may reflect the different functions that the proteins perform. In total, lectin-associated functions may be attributed to 149 transcripts, 37 were unambiguously identified at the protein level (S5 Table). These lectin-like proteins show low overlap between species of invertebrates, suggesting relatively rapid evolution of pattern recognition proteins involved in innate immunity. Most of transcripts/proteins described belong to tree lectin families, C-type, R-type and H-type.

C-type lectins

C-type lectins are a superfamily of diverse proteins able to bind specific carbohydrates in a Ca^{2+} -dependent manner. The CRDs contain characteristic double-loop structures stabilized by two highly conserved disulfide bridges located at the bases of the loops [26]. The second loop is structurally and evolutionarily flexible and is involved in Ca^{2+} -dependent carbohydrate binding. In total, 47 transcripts encoding proteins with C-type lectin CRDs have been found and 14 were identified also at the protein level. 17 of 21 complete transcripts have a predicted signal peptide indicating that they are secreted by cells to exert their functions (S5 Table) (Fig 6). Fig 7 shows remarkable sequence variability within incilarin-like proteins identified in our study. Incilarins are C-type lectins originally isolated from the water-soluble fraction of the body surface mucus of the land slug, *Incilaria fruhstorferi*, possessing hemagglutination activity [27].

R-type lectins

The R-type lectins are members of a superfamily of proteins which contain a carbohydrate-recognition domain (CRD) structurally similar to the CRD in ricin. All identified R-type lectins,

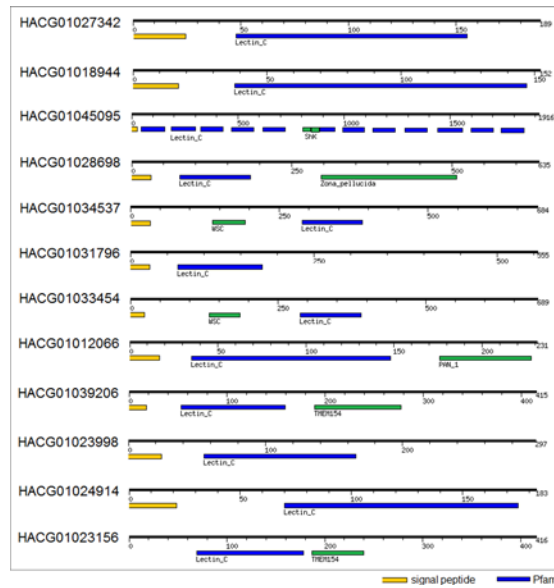


Fig 7. Alignment of amino acid sequences of incilarin-like proteins from *A. vulgaris*.

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are chimeric proteins consisting of an A chain with enzymatic activity (galactosyltransferase) linked through a disulfide bridge with a B chain with lectin activity (Fig 8). An analysis of the transcriptome predicts 32 unigenes coding proteins of R-type lectins. R-type lectine-like proteins were not identified at the protein level probably due to tissue-specific expression.

H-type lectins

The H-type lectins are proteins which contain a carbohydrate-recognition domain (CRD) structurally similar to *Helix pomatia* agglutinin (HPA) [28]. It has been shown that HPA is part of the innate immunity system of *H. pomatia* and a component of perivitelline fluid protecting fertilized eggs from bacteria. We identified 12 transcripts, 7 at the protein level. H-type lectins from *A. vulgaris* have a similar size and are predicted to share the same hexameric arrangement achieved by conserved cysteine residues involved in the intermolecular disulfide bridge formation (Figs 8 and 9).

Galectins

Galectins represent a family of structurally-related diverse lectins with carbohydrate binding specificity primarily to β -galactoside residues. Screening of the *A. vulgaris* transcriptome

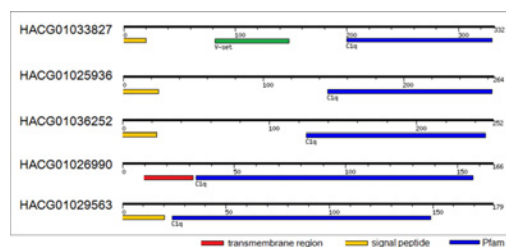


Fig 8. Domain architecture of galectins, H-type lectins, R-type lectins, L-type lectins and calnexin-like proteins found in the transcriptome of *A. vulgaris*.

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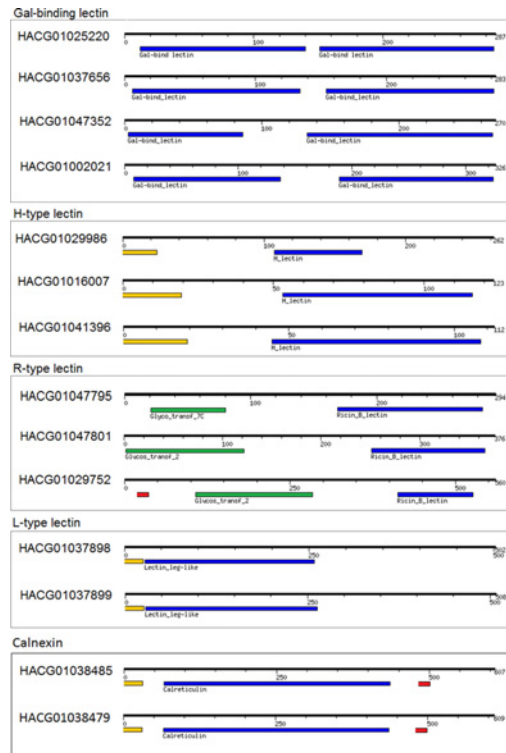


Fig 9. Alignment of amino acid sequences of H-type lectins from *A. vulgaris* with *Helix pomacea* and *Cepaea hortensis*.

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revealed that ten unigenes coding proteins share a galectin domain. Galectin domain-containing proteins expressed in *A. vulgaris* are not polymorphic, only a tandem-repeat type galectin has been identified (Fig 8). The absence of a signal sequence is consistent with other galectins, which are known to be actively secreted from diverse tissue via a “nonclassical” secretory pathway [29]. All of these lectins are most similar to gastropod galectins (*Aplysia californica*, *Biomphalaria glabrata*, etc.;blastp E value ranges from 0 to 2E–32).

L-type lectins

A member of L-type (legume-like) lectin family, ERGIC-53 is known as a pattern recognition receptor involved in the immune system of *E. sinensis* [30]. Transcriptomic analysis revealed the presence of two L-type lectin sequences with highest similarity to homolog of ERGIC-53 protein from *Littorina littorea* (blastp: E = 0.0; Identity = 57–58%). ERGIC-53 homologs share the characteristic architecture with an extracellular signal peptide and a single N-terminal L-type carbohydrate binding site (Fig 8).

Calnexin

Members of the evolutionarily conserved calnexin/calreticulin-superfamily bind to oligosaccharides containing terminal glucose residues. Calnexin acts as one of the pattern recognition receptors and has a crucial role in shrimp antibacterial immunity [31]. The search of *A. vulgaris* transcriptomic data revealed four calreticulin-like unigenes coding calnexin proteins with a highest similarity to *Aplysia californica* homologs (blastp: E = 0.0; Identity = 70–74%).

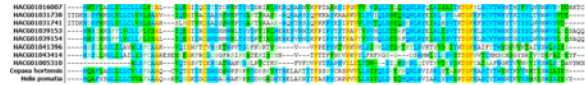


Fig 10. Domain architecture of C1q domain-containing proteins found in the transcriptome of *A. vulgaris*.

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Calnexin homologs contain a signal sequence and an additional C-terminal transmembrane helix (Fig 8).

C1q domain-containing proteins

The C1q domain-containing proteins (C1qDC) possessing lectin-like features are a family of proteins characterized by a globular C1q domain [32] regarded as an important player in innate immunity of bivalvia molluscs [33–37]. C1qDC proteins participate in several immune responses, such as pathogen recognition [38], microorganism agglutination [39] and mediating cell migration [40]. In addition, the C1q domain is involved in other immunological processes, such as phagocytosis [41], neutralization of viruses, cell adhesion and clearance of apoptotic cells. It is proposed that C1qDC proteins can activate an ancient complement system by the lectin pathway prior to the evolution of immunoglobulins [41]. The C1q domain has been considered as an extremely efficient pattern recognition domain with highly adaptive binding properties. The extreme versatility of C1q is due to the capability of the C1q domain to bind a variety of self and non-self ligands, including lipopolysaccharides [42], virus envelope proteins, outer membrane proteins from Gram-negative bacteria, phospholipids and some acute-phase proteins.

Based on sequence homology, molecular architecture and domain similarity, 34 C1q domain-containing sequences from the *A. vulgaris* transcriptome may be classified as members of the C1q family (S4 Table). Ten transcripts have been identified also at the protein level. Most of the complete C1qDC sequences displayed a signal peptide or a transmembrane domain on the N-terminus (Fig 10).

Toxin-like proteins

Escaping predation is essential to survival. To reduce predation, organisms have developed a diverse defence mechanisms. A few candidate effector genes were found in the slug transcriptome, including a family of potential pore-forming membrane attack/perforin (MACPF) domain proteins. Pore-forming membrane attack/perforin (MACPF) domain proteins have been described to be involved in the biochemical defence of apple snail eggs against predators [43].

Transcriptomic analysis of *A. vulgaris* revealed a novel toxin-like protein showing significant similarity to agatoxin, expressed in spider glands with a characteristic cysteine motive in the mature peptide. The agatoxin-homolog from *A. vulgaris* shares characteristic architecture with signal peptide, prepeptide sequence and characteristic cysteine motive in the mature peptide (Fig 11).



Fig 11. Alignment of amino acid sequences of toxin-like proteins from *A. vulgaris* with U8-agatoxine-Ao1a from *Agelena orientalis*.

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Taken together, one may speculate that the diversity and multitude of proteins with defensive potential along with already reported defensins (diterpene) of this slug [44] may contribute to success and survival of this terrestrial mollusc.

Data Accessibility

The sequence data has been submitted to the sequence read archive (SRA) database of GenBank (<http://www.ncbi.nlm.nih.gov/sra>) with the BioProject accession number PRJEB7891.

The mass spectrometry proteomics data have been deposited to the ProteomeXchange Consortium [45] via the PRIDE partner repository with the dataset identifier PXD002078 and 10.6019/PXD002078.

Supporting Information

S1 Table.

(XLSX)

S2 Table. List of 814 proteins identified from the mantel of *A. vulgaris* using 2-D electrophoretic separation.

(XLSX)

S3 Table. List of 2011 proteins identified from the mantel of *A. vulgaris* using 1-D electrophoretic separation.

(XLSX)

S4 Table. List of 2129 proteins identified from the mantel of *A. vulgaris*.

(XLSX)

S5 Table. List of lectin-like sequences and toxin-like sequences identified in *A. vulgaris*.

(XLSX)

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Author Contributions

Conceived and designed the experiments: JA GL. Performed the experiments: TB FJS GJ. Analyzed the data: RS TR JA. Contributed reagents/materials/analysis tools: MB. Wrote the paper: JA GL. Species identification: HS.

References

1. Gismervik K, Aspholm M, Rorvik LM, Bruheim T, Andersen A, Skaar I (2015) Invading slugs (*Arion vulgaris*) can be vectors for *Listeria monocytogenes*. J Appl Microbiol 118: 809–816. doi: [10.1111/jam.12750](https://doi.org/10.1111/jam.12750) PMID: [25580873](https://pubmed.ncbi.nlm.nih.gov/25580873/)
2. Gismervik K, Bruheim T, Rorvik LM, Haukeland S, Skaar I (2014) Invasive slug populations (*Arion vulgaris*) as potential vectors for *Clostridium botulinum*. Acta Vet Scand 56: 65. doi: [10.1186/s13028-014-0065-z](https://doi.org/10.1186/s13028-014-0065-z) PMID: [25277214](https://pubmed.ncbi.nlm.nih.gov/25277214/)
3. Stalder GL, Loncaric I, Walzer C (2014) Diversity of enterobacteria including beta-lactamase producing isolates associated with the Spanish slug (*Arion vulgaris*). Sci Total Environ 479–480: 11–16. doi: [10.1016/j.scitotenv.2014.01.103](https://doi.org/10.1016/j.scitotenv.2014.01.103) PMID: [24534696](https://pubmed.ncbi.nlm.nih.gov/24534696/)
4. Patel Z, Gill AC, Fox MT, Hermosilla C, Backeljau T, Breugelmans K, et al. (2014) Molecular identification of novel intermediate host species of *Angiostrongylus vasorum* in Greater London. Parasitol Res 113: 4363–4369. doi: [10.1007/s00436-014-4111-6](https://doi.org/10.1007/s00436-014-4111-6) PMID: [25195057](https://pubmed.ncbi.nlm.nih.gov/25195057/)

5. Feldmeyer B, Wheat CW, Krezdorn N, Rotter B, Pfenninger M (2011) Short read Illumina data for the de novo assembly of a non-model snail species transcriptome (*Radix balthica*, *Basommatophora*, *Pulmonata*), and a comparison of assembler performance. BMC Genomics 12: 317. doi: [10.1186/1471-2164-12-317](https://doi.org/10.1186/1471-2164-12-317) PMID: [21679424](https://pubmed.ncbi.nlm.nih.gov/21679424/)
6. Amin S, Prentis PJ, Gilding EK, Pavasovic A (2014) Assembly and annotation of a non-model gastropod (*Nerita melanotragus*) transcriptome: a comparison of de novo assemblers. BMC Res Notes 7: 488. doi: [10.1186/1756-0500-7-488](https://doi.org/10.1186/1756-0500-7-488) PMID: [25084827](https://pubmed.ncbi.nlm.nih.gov/25084827/)
7. Sun J, Wang M, Wang H, Zhang H, Zhang X, Thiyagarajan V, et al. (2012) De novo assembly of the transcriptome of an invasive snail and its multiple ecological applications. Mol Ecol Resour 12: 1133–1144. doi: [10.1111/1755-0998.12014](https://doi.org/10.1111/1755-0998.12014) PMID: [22994926](https://pubmed.ncbi.nlm.nih.gov/22994926/)
8. Kopylova E, Noe L, Touzet H (2012) SortMeRNA: fast and accurate filtering of ribosomal RNAs in meta-transcriptomic data. Bioinformatics 28: 3211–3217. doi: [10.1093/bioinformatics/bts611](https://doi.org/10.1093/bioinformatics/bts611) PMID: [23071270](https://pubmed.ncbi.nlm.nih.gov/23071270/)
9. Grabherr MG, Haas BJ, Yassour M, Levin JZ, Thompson DA, Amit I, et al. (2011) Full-length transcriptome assembly from RNA-Seq data without a reference genome. Nat Biotechnol 29: 644–652. doi: [10.1038/nbt.1883](https://doi.org/10.1038/nbt.1883) PMID: [21572440](https://pubmed.ncbi.nlm.nih.gov/21572440/)
10. Chen TW, Gan RC, Wu TH, Huang PJ, Lee CY, Chen YY, et al. (2012) FastAnnotator—an efficient transcript annotation web tool. BMC Genomics 13 Suppl 7: S9. doi: [10.1186/1471-2164-13-S7-S9](https://doi.org/10.1186/1471-2164-13-S7-S9) PMID: [23281853](https://pubmed.ncbi.nlm.nih.gov/23281853/)
11. Bae N, Lodl M, Pollak A, Lubec G (2012) Mass spectrometrical analysis of bilin-binding protein from the wing of *Hebomoia glaucippe* (Linnaeus, 1758) (Lepidoptera: Pieridae). Electrophoresis 33: 1787–1794. doi: [10.1002/elps.201100569](https://doi.org/10.1002/elps.201100569) PMID: [22740467](https://pubmed.ncbi.nlm.nih.gov/22740467/)
12. Ahmed KE, Chen WQ, John JP, Kang SU, Lubec G (2010) Complete sequencing of the recombinant granulocyte-colony stimulating factor (filgrastim) and detection of biotinylation by mass spectrometry. Amino Acids 38: 1043–1049. doi: [10.1007/s00726-009-0312-1](https://doi.org/10.1007/s00726-009-0312-1) PMID: [19526310](https://pubmed.ncbi.nlm.nih.gov/19526310/)
13. Chen WQ, Li L, Lubec G (2014) Proteomic differences between white and brown adipocytes. Amino Acids 46: 995–1008. doi: [10.1007/s00726-013-1657-z](https://doi.org/10.1007/s00726-013-1657-z) PMID: [24390458](https://pubmed.ncbi.nlm.nih.gov/24390458/)
14. Kang SU, Heo S, Lubec G (2011) Mass spectrometric analysis of GABAA receptor subtypes and phosphorylations from mouse hippocampus. Proteomics 11: 2171–2181. doi: [10.1002/pmic.201000374](https://doi.org/10.1002/pmic.201000374) PMID: [21538884](https://pubmed.ncbi.nlm.nih.gov/21538884/)
15. Candiano G, Bruschi M, Musante L, Santucci L, Ghiggeri GM, Carnemolla B, et al. (2004) Blue silver: a very sensitive colloidal Coomassie G-250 staining for proteome analysis. Electrophoresis 25: 1327–1333. PMID: [15174055](https://pubmed.ncbi.nlm.nih.gov/15174055/)
16. Bennett KL, Funk M, Tschernutter M, Breitwieser FP, Planyavsky M, Ubaida Mohien C, et al. (2011) Proteomic analysis of human cataract aqueous humour: Comparison of one-dimensional gel LCMS with two-dimensional LCMS of unlabelled and iTRAQ(R)-labelled specimens. J Proteomics 74: 151–166. doi: [10.1016/j.jprot.2010.10.002](https://doi.org/10.1016/j.jprot.2010.10.002) PMID: [20940065](https://pubmed.ncbi.nlm.nih.gov/20940065/)
17. Elias JE, Gygi SP (2007) Target-decoy search strategy for increased confidence in large-scale protein identifications by mass spectrometry. Nat Methods 4: 207–214. PMID: [17327847](https://pubmed.ncbi.nlm.nih.gov/17327847/)
18. Kaessmann H (2010) Origins, evolution, and phenotypic impact of new genes. Genome Res 20: 1313–1326. doi: [10.1101/gr.101386.109](https://doi.org/10.1101/gr.101386.109) PMID: [20651121](https://pubmed.ncbi.nlm.nih.gov/20651121/)
19. Cech TR, Steitz JA (2014) The noncoding RNA revolution—trashing old rules to forge new ones. Cell 157: 77–94. doi: [10.1016/j.cell.2014.03.008](https://doi.org/10.1016/j.cell.2014.03.008) PMID: [24679528](https://pubmed.ncbi.nlm.nih.gov/24679528/)
20. Conesa A, Gotz S, Garcia-Gomez JM, Terol J, Talon M, Robles M (2005) Blast2GO: a universal tool for annotation, visualization and analysis in functional genomics research. Bioinformatics 21: 3674–3676. PMID: [16081474](https://pubmed.ncbi.nlm.nih.gov/16081474/)
21. Voncken F, Gao F, Wadforth C, Harley M, Colasante C (2013) The phosphoarginine energy-buffering system of trypanosoma brucei involves multiple arginine kinase isoforms with different subcellular locations. PLoS One 8: e65908. doi: [10.1371/journal.pone.0065908](https://doi.org/10.1371/journal.pone.0065908) PMID: [23776565](https://pubmed.ncbi.nlm.nih.gov/23776565/)
22. Pfenninger M, Weigand A, Balint M, Klussmann-Kolb A (2014) Misperceived invasion: the Lusitanian slug (*Arion lusitanicus* auct. non-Mabille or *Arion vulgaris* Moquin-Tandon 1855) is native to Central Europe. Evol Appl 7: 702–713. doi: [10.1111/eva.12177](https://doi.org/10.1111/eva.12177) PMID: [25067951](https://pubmed.ncbi.nlm.nih.gov/25067951/)
23. Hatteland BA, Roth S, Andersen A, Kaasa K, Stoa B, T S (2013) Distribution and spread of the invasive slug *Arion vulgaris* MoquinTandon in Norway. Fauna norvegica 32: 13–26.
24. Kozłowski J (2007) The distribution, biology, population dynamics and harmfulness of *Arion lusitanicus* Mabillee, 1868 (Gastropoda: *Pulmonata*: *Arionidae*) in Poland. Journal of Plant Protection Research 47: 219–230.
25. Sun J, Zhang H, Wang H, Heras H, Dreon MS, Ituarte S, et al. (2012) First proteome of the egg perivitelline fluid of a freshwater gastropod with aerial oviposition. J Proteome Res 11: 4240–4248. doi: [10.1021/pr3003613](https://doi.org/10.1021/pr3003613) PMID: [22738194](https://pubmed.ncbi.nlm.nih.gov/22738194/)

26. Zelensky AN, Gready JE (2005) The C-type lectin-like domain superfamily. *FEBS J* 272: 6179–6217. PMID: [16336259](#)
27. Yuasa HJ, Furuta E, Nakamura A, Takagi T (1998) Cloning and sequencing of three C-type lectins from body surface mucus of the land slug, *Incilaria fruhstorferi*. *Comp Biochem Physiol B Biochem Mol Biol* 119: 479–484. PMID: [9734332](#)
28. Sanchez JF, Lescar J, Chazalet V, Audfray A, Gagnon J, Alvarez R, et al. (2006) Biochemical and structural analysis of *Helix pomatia* agglutinin. A hexameric lectin with a novel fold. *J Biol Chem* 281: 20171–20180. PMID: [16704980](#)
29. Nickel W (2003) The mystery of nonclassical protein secretion. A current view on cargo proteins and potential export routes. *Eur J Biochem* 270: 2109–2119. PMID: [12752430](#)
30. Huang Y, Tan JM, Wang Z, Yin SW, Huang X, Wang W, et al. (2014) Cloning and characterization of two different L-type lectin genes from the Chinese mitten crab *Eriocheir sinensis*. *Dev Comp Immunol* 46: 255–266. doi: [10.1016/j.dci.2014.04.015](#) PMID: [24796868](#)
31. Zhang Q, Wang XQ, Jiang HS, Jia WM, Zhao XF, Wang JX (2014) Calnexin functions in antibacterial immunity of *Marsupenaeus japonicus*. *Dev Comp Immunol* 46: 356–363. doi: [10.1016/j.dci.2014.05.009](#) PMID: [24858031](#)
32. Carland TM, Gerwick L (2010) The C1q domain containing proteins: Where do they come from and what do they do? *Dev Comp Immunol* 34: 785–790. doi: [10.1016/j.dci.2010.02.014](#) PMID: [20214925](#)
33. Gerlach D, Schlott B, Schmidt KH (2004) Cloning and expression of a sialic acid-binding lectin from the snail *Cepaea hortensis*. *FEMS Immunol Med Microbiol* 40: 215–221. PMID: [15039097](#)
34. Allam B, Pales Espinosa E, Tanguy A, Jeffroy F, Le Bris C, Paillard C (2014) Transcriptional changes in Manila clam (*Ruditapes philippinarum*) in response to Brown Ring Disease. *Fish Shellfish Immunol* 41: 2–11. doi: [10.1016/j.fsi.2014.05.022](#) PMID: [24882017](#)
35. Gestal C, Pallavicini A, Venier P, Novoa B, Figueras A (2010) MgC1q, a novel C1q-domain-containing protein involved in the immune response of *Mytilus galloprovincialis*. *Dev Comp Immunol* 34: 926–934. doi: [10.1016/j.dci.2010.02.012](#) PMID: [20219531](#)
36. He X, Zhang Y, Yu F, Yu Z (2011) A novel sialic acid binding lectin with anti-bacterial activity from the Hong Kong oyster (*Crassostrea hongkongensis*). *Fish Shellfish Immunol* 31: 1247–1250. doi: [10.1016/j.fsi.2011.08.021](#) PMID: [21906682](#)
37. Xu T, Xie J, Li J, Luo M, Ye S, Wu X (2012) Identification of expressed genes in cDNA library of hemocytes from the RLO-challenged oyster, *Crassostrea ariakensis* Gould with special functional implication of three complement-related fragments (CaC1q1, CaC1q2 and CaC3). *Fish Shellfish Immunol* 32: 1106–1116. doi: [10.1016/j.fsi.2012.03.012](#) PMID: [22484279](#)
38. Yu Y, Huang H, Wang Y, Yu Y, Yuan S, Huang S, et al. (2008) A novel C1q family member of amphioxus was revealed to have a partial function of vertebrate C1q molecule. *J Immunol* 181: 7024–7032. PMID: [18981122](#)
39. Kong P, Zhang H, Wang L, Zhou Z, Yang J, Zhang Y, et al. (2010) AiC1qDC-1, a novel gC1q-domain-containing protein from bay scallop *Argopecten irradians* with fungi agglutinating activity. *Dev Comp Immunol* 34: 837–846. doi: [10.1016/j.dci.2010.03.006](#) PMID: [20346969](#)
40. Tahtouh M, Croq F, Vizioli J, Sautiere PE, Van Camp C, Salzet M, et al. (2009) Evidence for a novel chemotactic C1q domain-containing factor in the leech nerve cord. *Mol Immunol* 46: 523–531. doi: [10.1016/j.molimm.2008.07.026](#) PMID: [18952286](#)
41. Wang L, Wang L, Zhang H, Zhou Z, Siva VS, Song L (2012) A C1q domain containing protein from scallop *Chlamys farreri* serving as pattern recognition receptor with heat-aggregated IgG binding activity. *PLoS One* 7: e43289. doi: [10.1371/journal.pone.0043289](#) PMID: [22905248](#)
42. Zhang H, Song L, Li C, Zhao J, Wang H, Qiu L, et al. (2008) A novel C1q-domain-containing protein from Zhikong scallop *Chlamys farreri* with lipopolysaccharide binding activity. *Fish Shellfish Immunol* 25: 281–289. doi: [10.1016/j.fsi.2008.06.003](#) PMID: [18603000](#)
43. Dreon MS, Frassa MV, Ceolin M, Ituarte S, Qiu JW, Sun J, et al. (2013) Novel animal defenses against predation: a snail egg neurotoxin combining lectin and pore-forming chains that resembles plant defense and bacteria attack toxins. *PLoS One* 8: e63782. doi: [10.1371/journal.pone.0063782](#) PMID: [23737950](#)
44. Schroeder FC, Gonzalez A, Eisner T, Meinwald J (1999) Miriamin, a defensive diterpene from the eggs of a land slug (*Arion sp.*). *Proc Natl Acad Sci U S A* 96: 13620–13625. PMID: [10570122](#)
45. Vizcaino JA, Deutsch EW, Wang R, Csordas A, Reisinger F, Rios D, et al. (2014) ProteomeXchange provides globally coordinated proteomics data submission and dissemination. *Nat Biotechnol* 32: 223–226. doi: [10.1038/nbt.2839](#) PMID: [24727771](#)

DATASET BRIEF

Gel-free mass spectrometry analysis of *Drosophila melanogaster* heads

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Membrane proteins play key roles in several fundamental biological processes such as cell signalling, energy metabolism and transport. Despite the significance, these still remain an under-represented group in proteomics datasets. Herein, a bottom-up approach to analyse an enriched membrane fraction from *Drosophila melanogaster* heads using multidimensional liquid chromatography (LC) coupled with tandem-mass spectrometry (MS/MS) that relies on complete solubilisation and digestion of proteins, is reported. An enriched membrane fraction was prepared using equilibrium density centrifugation on a discontinuous sucrose gradient, followed by solubilisation using the filter-aided sample preparation (FASP), tryptic and sequential chymotryptic digestion of proteins. Peptides were separated by reversed-phase (RP) LC at high pH in the first dimension and acidic RP-LC in the second dimension coupled directly to an Orbitrap Velos Pro mass spectrometer. A total number of 4812 proteins from 114 865 redundant and 38 179 distinct peptides corresponding to 4559 genes were identified in the enriched membrane fraction from fly heads. These included brain receptors, transporters and channels that are most important elements as drug targets or are linked to disease. Data are available via ProteomeXchange with identifier PXD001712 (<http://proteomecentral.proteomexchange.org/dataset/PXD001712>).

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Animal proteomics / *Drosophila melanogaster* / Mass spectrometry / Membrane proteins



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The fruit fly *Drosophila melanogaster* is used as prime model organism for experimental studies of eukaryotic organisms, possessing genetic, anatomic, behavioural, methodological and even economic advantages [1]. A complete genome of *D. melanogaster* has been determined [2] and protein-coding regions are well-annotated. Surprisingly, the human and the drosophila genome are apparently interrelated. The majority of human genes have counterparts in the fly genome and almost 75% of all known human disorder-related genes have fly homologues [3]. This, together with amenability in the genetic manipulations has made *Drosophila* a powerful, simple

model to understand human biology, and molecular and cellular mechanisms of human diseases. In the last decade, a few large-scale proteomics analyses were performed to validate gene models and uncover protein expression profiles in different parts of the fly body, cell types, developmental states and cellular fractions [4–7]. So far the most extensive study has covered 63% of the fly proteome by detecting 9124 proteins. This high coverage was achieved by combining sample diversity and applying multiple fractionation strategies [4]. Lee et al. [8] have reported a proteomics dataset from fly heads based on two-dimensional electrophoresis separations. They have shown a high overlap of proteins found in heads and brains, nearly 90% and therefore heads can be considered for analyses of proteins from the central nervous system. Due to

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limitations of their two-dimensional gel-based approach only very few membrane proteins were identified in the above-mentioned dataset.

A comprehensive protein coverage of *Drosophila* proteome has not been achieved; membrane proteins still remain under-represented in proteomics studies and there are still gene models relying on computational predictions only. Herein, analysis of an enriched membrane fraction from fly heads through a gel-free approach is reported. The used gel-free bottom-up proteomic approach employs an initial proteolytic digestion of the complex mixture with fractionation at the peptide level using multidimensional liquid chromatography prior to nano-LC separation reducing the complexity of the sample.

D. melanogaster heads were isolated from adult flies using a standard freezing protocol [9]. Heads were then ground into a fine powder with a pre-chilled mortar and pestle. Powder was re-suspended in ice-cold homogenization buffer (10 mM HEPES, pH 7.5, 300 mM sucrose, protease inhibitor (Roche Molecular Biochemicals, Mannheim, Germany)). The homogenate was centrifuged for 10 min at $1000 \times g$ and the supernatant was centrifuged at $50\,000 \times g$ for 30 min at 4°C . Subsequently, the pellet was re-suspended in 4 mL washing buffer (homogenization buffer without sucrose), kept on ice for 30 min and centrifuged at $50\,000 \times g$ for 30 min. A total plasma membrane fraction was separated by equilibrium density centrifugation on a discontinuous sucrose gradient. The pellet, re-suspended in washing buffer, was layered on top of the sucrose cushion (1 M and 1.25 M sucrose solution) followed by centrifugation at $70\,000 \times g$ for 2 h at 4°C . After centrifugation, fractions from the sucrose interface were collected and diluted ten times with washing buffer and subsequently centrifuged for 30 min at $100\,000 \times g$. The pellet was stored at 80°C until use.

Membrane protein extraction was carried out according to a previous study with minor modifications [10]. Protein quantities were estimated by the Pierce 660 protein assay (ThermoFisher Scientific, IL, USA). 300 μg of the sample (1 \times 100 μg each) were digested with trypsin using the filter-aided sample preparation (FASP) as previously described with minor modifications [11, 12]. Samples were mixed in the filter unit with 8 M urea in 100 mM Tris-HCl (pH 8.5) (UA) and centrifuged at $14\,000 \times g$ for 15 min at 22°C . Any remaining detergent was exchanged by urea with 200 μL UA. The proteins were alkylated by the addition of 100 μL of 50 mM iodoacetamide in UA and incubated for 30 min at 22°C . Subsequently, excess reagent was removed by 3 \times 100 μL UA solution followed by 3 \times 100 μL of 50 mM TEAB buffer (Sigma-Aldrich, Hamburg, Germany). Proteins were digested overnight at 37°C with trypsin at an enzyme to protein ratio of 1:100 w/w. Tryptic peptides were recovered from the filter with 40 μL of 50 mM TEAB buffer followed by 50 μL of 0.5 M NaCl. The remaining semi-digested material on the FASP filter was washed with 50 μL 50 mM TEAB buffer and further digested with chymotrypsin (enzyme to protein ratio of 1:10 w/w). Pooled tryptically-digested and se-

quential trypsin–chymotrypsin digested samples were acidified, desalted and concentrated with customised reversed-phase C18 stage tips [13]. Bound peptides were eluted with a buffer containing 90% acetonitrile and 100 mM TEAB and lyophilised in a vacuum concentrator (Eppendorf, Hamburg, Germany). Samples were pooled, peptides were separated by basic reversed-phase liquid chromatography [14] and 50 fractions were collected for trypsin and 20 fractions for the sequential trypsin–chymotrypsin.

Acidified fractions were analysed by LC-MS/MS. Details of the procedure were as described previously with minor modifications [15]. Peptide fractions were separated by nano-LC (Dionex Ultimate 3000) equipped with a μ -precolumn (C18, 5 m, 100 \AA , 5×0.3 mm) and an Acclaim PepMap RSLC nanocolumn (C18, 2 μm , 100 \AA , 150×0.075 mm) (Thermo Fisher Scientific, Vienna, Austria). Eight microliters samples were injected and concentrated on the enrichment column for 2 min using 0.05% TFA as isocratic solvent at a flow rate of 20 $\mu\text{L}/\text{min}$. The column was then switched in the nanoflow circuit, and the sample was loaded on the nanocolumn at a flow rate of 200 nL/min and separated using the following gradient: solvent A: water, 0.05% TFA; solvent B: ACN/water 80/20 v/v, 0.05% TFA; 0–4 min: 4% B; 4–60 min 4–40% B, 60–95 min: 40–95% B, 95–105 min: 95% B, 105–105.1 min: 95–4% B, 105.1–120 min: 4% B. The sample was ionized in the nanospray source equipped with stainless steel emitters (Thermo Fisher Scientific, Vienna, Austria) and analysed in a Thermo Orbitrap Velos Pro mass spectrometer in positive ion mode by alternating full-scan MS (m/z 380–2000) in the ICR cell and MS/MS by CID of the 20 most intense peaks in the ion trap with dynamic exclusion enabled.

LC-MS/MS data were analysed by searching the UniProtKB *Drosophila melanogaster* database (41 965 sequences, download on January 16th, 2015) with Proteome Discoverer 1.4 (Thermo Fisher Scientific, Vienna, Austria) and MAS-COT 2.4 (MatrixScience, London, UK). Detailed search criteria were used as follows; enzyme: trypsin or tryptchymo with a maximum of two missing cleavage sites; fixed modification: carbamidomethylation (C); variable modification: oxidation (M); search mode: MS/MS ion search with decoy database search included; peptide mass tolerance ± 10 ppm; MS/MS mass tolerance ± 0.5 Da. Using the target-decoy search strategy [16], peptide matches with a false discovery rate (FDR) lower than 1% were filtered. All proteins with a minimum of two distinct peptides and ion score > 20 were accepted.

A total of 114 865 redundant and 38 179 distinct peptides were identified and assigned to 4812 proteins, including splicing variants. Of these, 3920 proteins (81, 4%) were identified with more than two distinct peptides (Fig. 1A). On average eight distinct unique peptides were identified per protein. The list of the identified proteins in order of exponentially modified protein abundance index (emPAI) [17, 18] is given in Supporting Information Tables S1 and S2. Identified proteins correspond to 4559 genes which represent 33, 5% of the predicted genes in *Drosophila melanogaster*. The MS proteomics data have been deposited in the ProteomeXchange

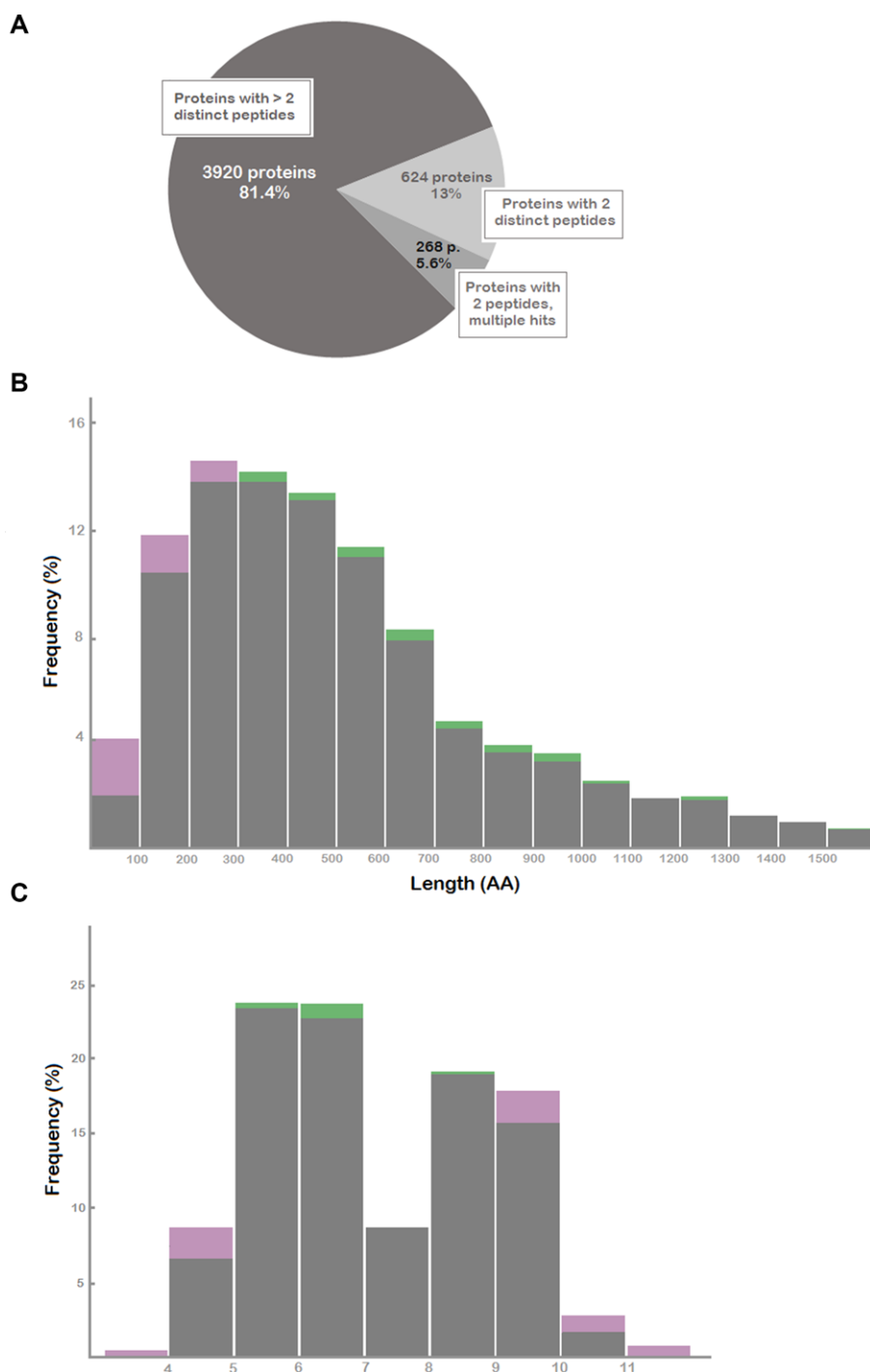


Figure 1. Experimentally identified proteins with respect to the number of distinct peptides (A). Distributions of length (B) and pI (C) of experimentally identified proteins (4812 proteins) in comparison to all proteins (20 328 proteins) in *Drosophila melanogaster* including splice variants. Under-represented areas are shown in pink, over-represented areas in green.

Consortium (<http://proteomecentral.proteomexchange.org>) via the PRIDE partner repository [19] with the dataset identifier PXD001712 and 10.6019/PXD001712. The dataset was classified with respect to several aspects (length, isoelectric point, transmembrane domains and functional annotation). Length and pI distribution of identified proteins are shown in Fig. 1B and C. The TMHMM (www.cbs.dtu.dk/services/TMHMM/) algorithm was used to

predict transmembrane regions from the identified proteins [20]. 35% of proteins (1679 proteins) were identified as membrane proteins containing at least one transmembrane domain and 65% (3133 proteins) had no confirmed or predicted transmembrane domain (Fig. 2).

The identified proteins were categorized to GO terms using the AmiGO2 bioinformatics Tool [21]. 3565 gene models were considered for functional analysis. An enrichment

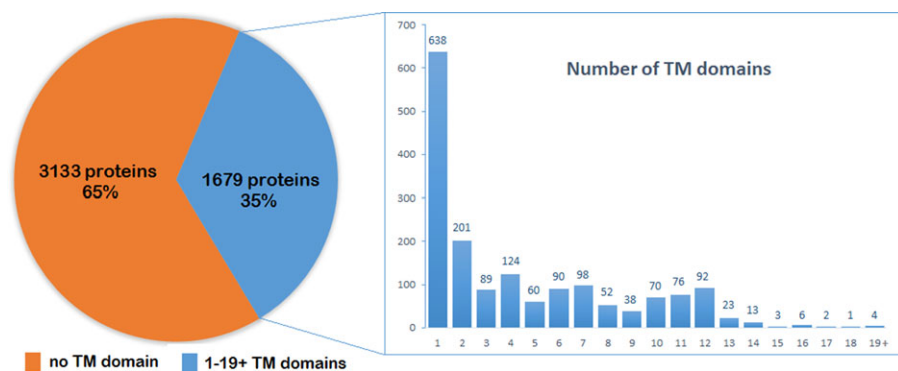


Figure 2. Transmembrane domain distribution among identified proteins.

analysis has been performed against *D. melanogaster* background (13 624 gene models) and GO terms with higher than two-fold enrichment for individual categories are shown in Fig. 3. The list of GO terms, number of associated proteins, fold enrichment and *p*-values for each GO term is given in Supporting Information Table S6. We identified several ionotropic receptors for glutamate and acetylcholine, and metabotropic receptors for acetylcholine, GABA, glutamate, dopamine, octopamine, serotonin and adenosine (Supporting Information Table S3). Based upon the emPAI index we compared the relative abundance of identified receptors. Ionotropic receptors NmdaR1 and NmdaR2, nicotinic

acetylcholine receptor subunits $\alpha 4$ and $\beta 3$; and metabotropic dopamine 1 receptor were the most abundant receptors. The major amino-acid transporters associated with neurotransmitters glutamate, glycine and dopamine were unambiguously identified (Supporting Information Table S4). The most common ion channel families: voltage-gated calcium channels, voltage-gated chloride channels, potassium channels, and voltage-dependent sodium channels were detected and listed in Supporting Information Table S5.

Taken together, a gel-free mass spectrometrical approach using the FASP principle was used to provide a dataset that may be useful for studies on membrane proteins that are

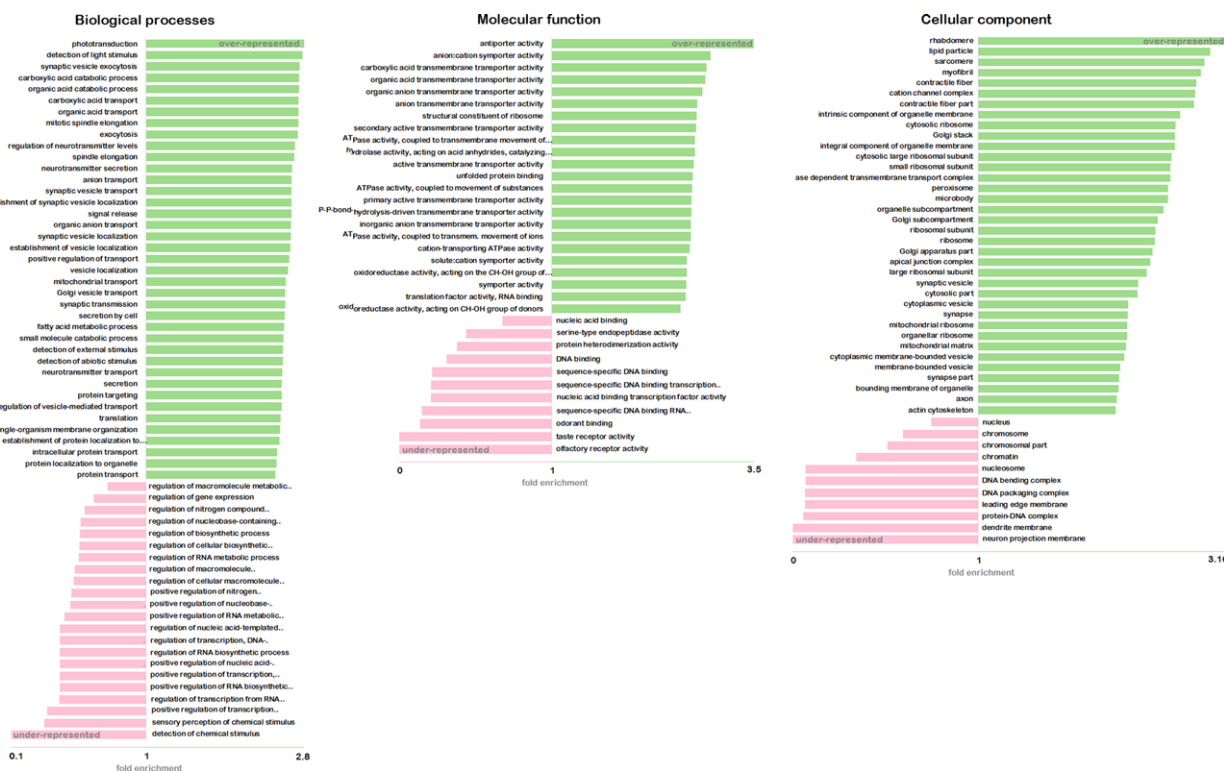


Figure 3. GO term enrichment analysis of experimentally identified proteins. 3565 gene models, considered for functional analysis were compared to 13 624 predicted gene models. The figure shows GO terms that are significantly enriched ($p < 1E-4$) by more than two-fold or are significantly under-represented for all tree functional categories.

mainly representing drug targets and are linked to a large series of human disorders and diseases.

The MS proteomics data in this paper have been deposited in the ProteomeXchange Consortium (<http://proteomecentral.proteomexchange.org>) via the PRIDE partner repository [19]: dataset identifier PXD001712.

The authors have declared no conflict of interest.

References

- [1] Prussing, E., Historical trauma: politics of a conceptual framework. *Transcult. Psychiatry* 2014, 51, 436–458.
- [2] Adams, M. D., Celniker, S. E., Holt, R. A., Evans, C. A. et al., The genome sequence of *Drosophila melanogaster*. *Science* 2000, 287, 2185–2195.
- [3] Bier, E., *Drosophila*, the golden bug, emerges as a tool for human genetics. *Nat. Rev. Genetics* 2005, 6, 9–23.
- [4] Brunner, E., Ahrens, C. H., Mohanty, S., Baetschmann, H. et al., A high-quality catalog of the *Drosophila melanogaster* proteome. *Nat. Biotechnol.* 2007, 25, 576–583.
- [5] Xing, X., Zhang, C., Li, N., Zhai, L. et al., Qualitative and quantitative analysis of the adult *Drosophila melanogaster* proteome. *Proteomics* 2014, 14, 286–290.
- [6] Wasbrough, E. R., Dorus, S., Hester, S., Howard-Murkin, J. et al., The *Drosophila melanogaster* sperm proteome-II (DmSP-II). *J. Proteomics* 2010, 73, 2171–2185.
- [7] Taraszka, J. A., Kurulugama, R., Sowell, R. A., Valentine, S. J. et al., Mapping the proteome of *Drosophila melanogaster*: analysis of embryos and adult heads by LC-IMS-MS methods. *J. Proteome Res.* 2005, 4, 1223–1237.
- [8] Lee, T. R., Huang, S. H., Lee, C. C., Lee, H. Y. et al., Proteome reference map of *Drosophila melanogaster* head. *Proteomics* 2012, 12, 1875–1878.
- [9] Jensen, K., Sanchez-Garcia, J., Williams, C., Khare, S. et al., Purification of transcripts and metabolites from *Drosophila* heads. *J. Visualized Exp.* 2013, e50245.
- [10] Luche, S., Santoni, V., Rabilloud, T., Evaluation of nonionic and zwitterionic detergents as membrane protein solubilizers in two-dimensional electrophoresis. *Proteomics* 2003, 3, 249–253.
- [11] Wisniewski, J. R., Zougman, A., Nagaraj, N., Mann, M., Universal sample preparation method for proteome analysis. *Nat. Methods* 2009, 6, 359–362.
- [12] Manza, L. L., Stamer, S. L., Ham, A. J., Codreanu, S. G., Liebler, D. C., Sample preparation and digestion for proteomic analyses using spin filters. *Proteomics* 2005, 5, 1742–1745.
- [13] Rappsilber, J., Mann, M., Ishihama, Y., Protocol for micro-purification, enrichment, pre-fractionation and storage of peptides for proteomics using StageTips. *Nat. Protoc.* 2007, 2, 1896–1906.
- [14] Gilar, M., Olivova, P., Daly, A. E., Gebler, J. C., Two-dimensional separation of peptides using RP-RP-HPLC system with different pH in first and second separation dimensions. *J. Sep. Sci.* 2005, 28, 1694–1703.
- [15] Heo, S., Spoerk, S., Birner-Gruenberger, R., Lubec, G., Gel-based mass spectrometric analysis of hippocampal transmembrane proteins using high resolution LTQ Orbitrap Velos Pro. *Proteomics* 2014, 14, 2084–2088.
- [16] Elias, J. E., Gygi, S. P., Target-decoy search strategy for increased confidence in large-scale protein identifications by mass spectrometry. *Nat. Methods* 2007, 4, 207–214.
- [17] Ishihama, Y., Oda, Y., Tabata, T., Sato, T. et al., Exponentially modified protein abundance index (emPAI) for estimation of absolute protein amount in proteomics by the number of sequenced peptides per protein. *Mol. Cell. Proteomics* 2005, 4, 1265–1272.
- [18] Shinoda, K., Tomita, M., Ishihama, Y., emPAI Calc—for the estimation of protein abundance from large-scale identification data by liquid chromatography-tandem mass spectrometry. *Bioinformatics* 2010, 26, 576–577.
- [19] Vizcaino, J. A., Deutsch, E. W., Wang, R., Csordas, A. et al., ProteomeXchange provides globally coordinated proteomics data submission and dissemination. *Nat. Biotechnol.* 2014, 32, 223–226.
- [20] Sonnhammer, E. L., vonHeijne, G., Krogh, A., A hidden Markov model for predicting transmembrane helices in protein sequences. *Proc. Int. Conf. Intell. Syst. Mol. Biol.* 1998, 6, 175–182.
- [21] Carbon, S., Ireland, A., Mungall, C. J., Shu, S. et al., AmiGO: online access to ontology and annotation data. *Bioinformatics* 2009, 25, 288–289.

8.2. Table 1. List of proteins identified in 2-DE proteomics study (UP-unique peptide number; PC-protein coverage (%); NM-number of assigned peptide matches; NS-number of protein sequences; PS-protein score)

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 318609972 dbj BAJ61596.1	96	71	97	2700	44575
gi 524888508 ref XP_005100824.1	60	38	78	595	9831
gi 524866478 ref XP_005090060.1	57	86	57	667	11570
gi 305689815 gb ADM64337.1	48	73	48	965	19654
gi 405975785 gb EKC40331.1	45	64	48	342	6259
gi 220067263 gb ACL79834.1	44	76	44	566	12445
gi 556096138 gb ESO84790.1	43	65	46	676	15337
gi 385145402 emb CCG28026.1	42	86	45	2211	30113
gi 127773 sp P24733.1 MYS_ARGIR	41	39	70	660	11244
gi 147905746 ref NP_001080566.1	39	73	47	395	4497
gi 405960426 gb EKC26351.1	39	67	39	228	4756
gi 524903383 ref XP_005107892.1	39	58	39	256	6023
gi 524916025 ref XP_005112795.1	38	69	40	736	16769
gi 73695954 gb AAZ80783.1	38	76	38	2339	16103
gi 126697420 gb ABO26667.1	37	62	37	165	2543
gi 524888358 ref XP_005100750.1	36	62	36	154	3335
gi 524894502 ref XP_005103748.1	36	54	39	307	6303
gi 524896814 ref XP_005104878.1	35	17	35	128	1273
gi 71726729 gb AAZ39528.1	33	61	45	481	7397
gi 524909114 ref XP_005109609.1	32	90	33	791	10634
gi 42559558 sp O97192.1 TPM_HELAS	30	85	40	1540	26058
gi 472824675 dbj BAN09030.1	30	55	30	879	14670
gi 556096092 gb ESO84744.1	30	44	30	217	4565
gi 556110866 gb ESO99517.1	30	27	64	289	5337
gi 829207 emb CAA39415.1	30	61	30	335	6356
gi 296837083 gb ADH59421.1	28	67	42	1397	26938
gi 443692429 gb ELT94023.1	28	71	28	313	6839
gi 524885601 ref XP_005099406.1	28	39	28	180	2923
gi 524884921 ref XP_005099073.1	27	45	27	576	11029
gi 524895745 ref XP_005104354.1	27	55	38	556	11670
gi 524909114 ref XP_005109609.1	26	68	27	458	5086
gi 526117385 ref NP_001267755.1	26	74	26	376	8603
gi 556097340 gb ESO85992.1	26	47	27	343	7708
gi 124233 sp P22488.2 IFEA_HELAS	25	52	30	304	6401
gi 524875601 ref XP_005094526.1	25	48	32	197	4019

List of proteins identified in 2-DE proteomics study

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524887592 ref XP_005100380.1	25	52	25	143	2627
gi 524883288 ref XP_005098277.1	24	42	24	97	1424
gi 524883288 ref XP_005098277.1	24	42	24	97	1424
gi 524895745 ref XP_005104354.1	24	40	40	301	5740
gi 524911954 ref XP_005110834.1	24	43	25	161	2654
gi 524916507 ref XP_005113030.1	24	60	28	195	3926
gi 325296989 ref NP_001191508.1	23	67	23	102	1713
gi 524885520 ref XP_005099366.1	23	80	23	291	2963
gi 524892924 ref XP_005102985.1	23	50	23	106	1768
gi 524896814 ref XP_005104878.1	23	32	23	136	1278
gi 524912440 ref XP_005111068.1	23	67	23	501	7869
gi 556097683 gb ESO86335.1	23	63	23	183	2442
gi 524872922 ref XP_005093216.1	22	42	22	125	3314
gi 524886817 ref XP_005100000.1	22	43	22	108	2123
gi 524895745 ref XP_005104354.1	22	29	24	96	2060
gi 524910869 ref XP_005110308.1	22	53	22	66	1223
gi 524882748 ref XP_005098012.1	21	38	29	90	1701
gi 524900673 ref XP_005106755.1	21	64	21	144	2238
gi 524916507 ref XP_005113030.1	21	64	26	125	2892
gi 556095567 gb ESO84220.1	21	43	22	38	770
gi 524865955 ref XP_005089803.1	20	68	20	142	2804
gi 524885898 ref XP_005099550.1	20	34	20	44	721
gi 524900516 ref XP_005106678.1	20	59	20	404	5064
gi 524911947 ref XP_005110831.1	20	50	20	97	1234
gi 524915789 ref XP_005112685.1	20	35	20	80	1284
gi 526117385 ref NP_001267755.1	20	55	20	194	3066
gi 556110866 gb ESO99517.1	20	15	34	150	1602
gi 218683627 gb ACL00842.1	19	39	19	31	490
gi 524865670 ref XP_005089662.1	19	90	20	397	10161
gi 524883990 ref XP_005098620.1	19	56	19	128	2364
gi 524889526 ref XP_005101318.1	19	33	19	77	1281
gi 524896814 ref XP_005104878.1	19	16	19	61	676
gi 6682319 emb CAB64662.1	19	45	28	223	4006
gi 524872674 ref XP_005093097.1	18	44	18	62	1156
gi 524882968 ref XP_005098119.1	18	36	18	59	1678
gi 524884737 ref XP_005098987.1	18	38	18	79	1549
gi 524893715 ref XP_005103366.1	18	61	18	103	563
gi 524895745 ref XP_005104354.1	18	46	35	249	5457
gi 524899582 ref XP_005106220.1	18	56	18	106	2069
gi 524915771 ref XP_005112676.1	18	48	18	149	1821
gi 42559558 sp O97192.1 TPM_HELAS	17	65	29	574	9620

List of proteins identified in 2-DE proteomics study

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524871885 ref XP_005092713.1	17	43	17	77	575
gi 524895745 ref XP_005104354.1	17	46	21	222	4906
gi 526117405 ref NP_001267760.1	17	89	23	561	7405
gi 556110866 gb ESO99517.1	17	17	26	41	1148
gi 2073142 dbj BAA19861.1	16	71	16	138	1786
gi 346987840 gb AEO51766.1	16	8	17	23	325
gi 524872618 ref XP_005093070.1	16	28	16	30	834
gi 524886261 ref XP_005099727.1	16	18	16	100	1693
gi 524886695 ref XP_005099940.1	16	43	16	91	1539
gi 524888508 ref XP_005100824.1	16	18	41	64	994
gi 524898231 ref XP_005105567.1	16	36	16	42	435
gi 524898324 ref XP_005105612.1	16	32	16	52	1149
gi 524907580 ref XP_005108906.1	16	8	16	26	301
gi 773571 emb CAA60122.1	16	29	17	37	725
gi 325197136 ref NP_001191419.1	15	84	15	137	2192
gi 346987840 gb AEO51766.1	15	22	15	78	1164
gi 51105058 gb AAT97089.1	15	31	16	38	290
gi 524871134 ref XP_005092343.1	15	41	15	171	4821
gi 524875728 ref XP_005094588.1	15	42	15	77	829
gi 524876525 ref XP_005094978.1	15	54	19	144	1905
gi 524881606 ref XP_005097453.1	15	40	15	23	284
gi 524909838 ref XP_005109815.1	15	42	15	72	1527
gi 328933188 gb AEB70965.1	14	49	14	86	1454
gi 405953142 gb EKC20858.1	14	33	14	81	973
gi 524876525 ref XP_005094978.1	14	50	18	134	2002
gi 524878973 ref XP_005096166.1	14	39	14	124	2877
gi 524883622 ref XP_005098438.1	14	32	14	26	576
gi 524885795 ref XP_005099501.1	14	51	22	215	4236
gi 524891584 ref XP_005102325.1	14	36	14	79	1813
gi 524911952 ref XP_005110833.1	14	60	17	151	1575
gi 524911954 ref XP_005110834.1	14	37	16	74	1039
gi 524913044 ref XP_005111356.1	14	41	14	59	899
gi 524916317 ref XP_005112938.1	14	45	14	85	1262
gi 524917241 ref XP_005113381.1	14	36	14	190	3548
gi 556095158 gb ESO83811.1	14	31	14	24	467
gi 564743500 gb AHC02701.1	14	72	34	1115	21582
gi 321472487 gb EFX83457.1	13	21	13	38	516
gi 524865670 ref XP_005089662.1	13	62	16	168	4244
gi 524867613 ref XP_005090615.1	13	14	13	27	392
gi 524871402 ref XP_005092474.1	13	29	13	43	759
gi 524875601 ref XP_005094526.1	13	32	21	70	1251

List of proteins identified in 2-DE proteomics study

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524880840 ref XP_005097078.1	13	40	13	46	1014
gi 524882748 ref XP_005098012.1	13	27	18	67	901
gi 524890599 ref XP_005101843.1	13	30	13	26	341
gi 524903522 ref XP_005107960.1	13	42	13	93	1813
gi 524911950 ref XP_005110832.1	13	32	13	32	501
gi 555688717 gb ESN91949.1	13	62	13	192	2334
gi 325297040 ref NP_001191523.1	12	40	12	43	439
gi 385145402 emb CCG28026.1	12	36	12	63	1157
gi 524867573 ref XP_005090597.1	12	32	12	117	2361
gi 524873169 ref XP_005093336.1	12	38	12	26	625
gi 524876525 ref XP_005094978.1	12	39	14	81	1539
gi 524881811 ref XP_005097555.1	12	35	12	118	2051
gi 524895342 ref XP_005104156.1	12	30	12	23	591
gi 524898259 ref XP_005105581.1	12	48	12	106	1802
gi 524907580 ref XP_005108906.1	12	6	12	27	317
gi 524907822 ref XP_005109023.1	12	43	12	20	313
gi 524913368 ref XP_005111515.1	12	37	12	59	1383
gi 524916601 ref XP_005113074.1	12	10	12	16	216
gi 576693750 gb EUB57355.1	12	51	12	107	1238
gi 74912853 sp Q6QUW1.1 RGP51_LYMST	12	31	16	84	1496
gi 241273677 ref XP_002406620.1	11	26	11	82	1095
gi 325297052 ref NP_001191530.1	11	65	35	109	1751
gi 524868522 ref XP_005091063.1	11	12	11	19	329
gi 524871732 ref XP_005092637.1	11	6	11	52	801
gi 524872616 ref XP_005093069.1	11	17	11	27	741
gi 524876391 ref XP_005094913.1	11	32	11	45	724
gi 524876525 ref XP_005094978.1	11	42	16	83	1709
gi 524876525 ref XP_005094978.1	11	44	17	149	2175
gi 524877261 ref XP_005095336.1	11	23	11	40	708
gi 524910025 ref XP_005109900.1	11	52	11	34	479
gi 524911947 ref XP_005110831.1	11	61	14	88	1049
gi 556104876 gb ESO93528.1	11	16	14	31	461
gi 556105302 gb ESO93954.1	11	43	11	44	836
gi 585719304 ref XP_006826038.1	11	6	11	14	147
gi 325297146 ref NP_001191582.1	10	46	14	28	435
gi 405972388 gb EKC37161.1	10	43	10	11	240
gi 513137512 gb AGO20698.1	10	18	10	23	270
gi 524866272 ref XP_005089958.1	10	35	10	29	588
gi 524867792 ref XP_005090703.1	10	22	10	23	499
gi 524879223 ref XP_005096289.1	10	42	10	46	1026

List of proteins identified in 2-DE proteomics study

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524879388 ref XP_005096370.1	10	33	10	59	1472
gi 524882422 ref XP_005097855.1	10	35	10	75	1587
gi 524884878 ref XP_005099053.1	10	39	10	118	1913
gi 524885821 ref XP_005099514.1	10	33	10	15	276
gi 524887319 ref XP_005100247.1	10	26	10	32	592
gi 524891306 ref XP_005102188.1	10	35	10	40	723
gi 524894818 ref XP_005103900.1	10	48	11	88	1016
gi 524896250 ref XP_005104602.1	10	40	10	60	978
gi 524896457 ref XP_005104703.1	10	37	10	30	451
gi 524902628 ref XP_005107543.1	10	45	10	34	351
gi 524903041 ref XP_005107738.1	10	20	10	21	175
gi 524906745 ref XP_005108595.1	10	37	10	28	516
gi 524915110 ref XP_005112361.1	10	21	10	18	280
gi 556098588 gb ESO87240.1	10	38	10	37	723
gi 576693750 gb EUB57355.1	10	67	10	60	1229
gi 126697356 gb ABO26635.1	9	35	9	58	1175
gi 150251390 gb ABR68007.1	9	23	9	95	1958
gi 325296986 ref NP_001191507.1	9	12	9	34	621
gi 346987844 gb AEO51768.1	9	5	9	35	985
gi 524864482 ref XP_005089076.1	9	25	9	26	612
gi 524866272 ref XP_005089958.1	9	25	9	21	433
gi 524866912 ref XP_005090271.1	9	19	9	48	891
gi 524869485 ref XP_005091535.1	9	16	9	46	642
gi 524869987 ref XP_005091780.1	9	62	9	26	473
gi 524874769 ref XP_005094119.1	9	16	9	19	419
gi 524878388 ref XP_005095881.1	9	17	9	19	364
gi 524881984 ref XP_005097638.1	9	56	9	24	327
gi 524884630 ref XP_005098934.1	9	32	9	47	1055
gi 524885795 ref XP_005099501.1	9	63	12	145	2592
gi 524886540 ref XP_005099863.1	9	31	9	21	620
gi 524886658 ref XP_005099922.1	9	22	9	27	387
gi 524889396 ref XP_005101256.1	9	32	9	22	357
gi 524891426 ref XP_005102246.1	9	29	9	52	1517
gi 524892644 ref XP_005102848.1	9	44	9	124	3499
gi 524893352 ref XP_005103192.1	9	28	9	20	430
gi 524901897 ref XP_005107352.1	9	37	9	36	791
gi 524907533 ref XP_005108883.1	9	32	9	50	1341
gi 524907610 ref XP_005108921.1	9	27	9	15	297
gi 524915397 ref XP_005112499.1	9	22	9	38	484
gi 54020890 ref NP_001005689.1	9	30	9	47	659
gi 556095158 gb ESO83811.1	9	21	9	36	971

List of proteins identified in 2-DE proteomics study

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 556095567 gb ESO84220.1	9	25	9	30	751
gi 556097226 gb ESO85878.1	9	24	9	23	462
gi 556099804 gb ESO88456.1	9	17	9	15	123
gi 556113964 gb ESP02616.1	9	20	9	32	712
gi 994895 gb AAB34494.1	9	34	11	70	1300
gi 156399584 ref XP_001638581.1	8	57	12	67	1619
gi 2073142 dbj BAA19861.1	8	43	8	122	2764
gi 296190822 ref XP_002743358.1	8	13	8	15	242
gi 325297144 ref NP_001191581.1	8	15	8	17	208
gi 351709298 gb EHB12217.1	8	54	8	191	3217
gi 402578741 gb EJW72694.1	8	38	8	108	1422
gi 524872618 ref XP_005093070.1	8	11	8	13	120
gi 524873600 ref XP_005093544.1	8	39	8	23	302
gi 524876525 ref XP_005094978.1	8	29	11	42	765
gi 524877031 ref XP_005095223.1	8	34	8	36	162
gi 524880725 ref XP_005097023.1	8	36	8	28	509
gi 524881984 ref XP_005097638.1	8	64	8	60	1510
gi 524882922 ref XP_005098097.1	8	31	8	28	633
gi 524885854 ref XP_005099530.1	8	36	8	141	2899
gi 524886289 ref XP_005099741.1	8	24	8	18	128
gi 524887834 ref XP_005100500.1	8	50	8	30	563
gi 524887883 ref XP_005100524.1	8	37	8	20	280
gi 524890176 ref XP_005101637.1	8	19	8	11	167
gi 524893673 ref XP_005103345.1	8	15	8	14	308
gi 524896407 ref XP_005104678.1	8	32	8	30	240
gi 524896874 ref XP_005104908.1	8	29	8	52	694
gi 524897999 ref XP_005105454.1	8	26	8	34	678
gi 524900380 ref XP_005106610.1	8	38	8	17	460
gi 524900390 ref XP_005106615.1	8	27	10	41	633
gi 524906491 ref XP_005108543.1	8	10	8	31	478
gi 524907238 ref XP_005108742.1	8	21	8	12	119
gi 524910284 ref XP_005110023.1	8	12	8	20	236
gi 524914230 ref XP_005111933.1	8	18	8	20	362
gi 541039654 gb ERG79357.1	8	21	8	11	79
gi 556103805 gb ESO92457.1	8	27	8	17	423
gi 556108834 gb ESO97486.1	8	19	8	22	503
gi 126697362 gb ABO26638.1	7	46	7	45	567
gi 168252891 gb ACA23131.1	7	53	7	16	302
gi 2073142 dbj BAA19861.1	7	55	7	190	4407
gi 291235073 ref XP_002737469.1	7	24	7	19	253
gi 346987844 gb AEO51768.1	7	4	8	9	190

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 353236678 emb CCA68668.1	7	24	7	13	266
gi 51105064 gb AAT97092.1	7	31	7	28	580
gi 524865020 ref XP_005089342.1	7	17	7	15	340
gi 524866304 ref XP_005089974.1	7	23	7	12	135
gi 524866627 ref XP_005090132.1	7	35	7	18	110
gi 524870833 ref XP_005092197.1	7	22	7	11	154
gi 524873892 ref XP_005093688.1	7	31	7	11	399
gi 524874065 ref XP_005093774.1	7	26	7	18	335
gi 524874374 ref XP_005093923.1	7	52	14	108	1696
gi 524874655 ref XP_005094062.1	7	6	7	31	499
gi 524876098 ref XP_005094768.1	7	17	7	22	531
gi 524876525 ref XP_005094978.1	7	30	8	22	344
gi 524877054 ref XP_005095234.1	7	28	7	9	144
gi 524881743 ref XP_005097521.1	7	34	7	75	1275
gi 524882484 ref XP_005097885.1	7	18	7	12	127
gi 524882717 ref XP_005097997.1	7	22	7	29	523
gi 524883540 ref XP_005098397.1	7	29	7	11	282
gi 524888414 ref XP_005100778.1	7	33	7	20	256
gi 524891844 ref XP_005102454.1	7	23	7	10	104
gi 524892967 ref XP_005103005.1	7	47	7	43	616
gi 524894544 ref XP_005103769.1	7	33	7	8	132
gi 524895123 ref XP_005104049.1	7	40	7	10	250
gi 524895893 ref XP_005104427.1	7	12	7	16	393
gi 524896519 ref XP_005104733.1	7	34	9	24	430
gi 524897999 ref XP_005105454.1	7	26	7	16	230
gi 524906739 ref XP_005108592.1	7	28	7	16	514
gi 524908996 ref XP_005109585.1	7	17	7	19	317
gi 524910895 ref XP_005110321.1	7	20	7	11	272
gi 524911570 ref XP_005110646.1	7	2	7	13	190
gi 524912929 ref XP_005111300.1	7	56	7	34	540
gi 524914829 ref XP_005112224.1	7	16	7	16	311
gi 524916452 ref XP_005113003.1	7	16	7	46	732
gi 524916607 ref XP_005113077.1	7	33	7	59	765
gi 529444582 ref XP_005242003.1	7	43	7	11	175
gi 54020890 ref NP_001005689.1	7	24	7	19	343
gi 546686668 gb ERL95792.1	7	55	7	58	735
gi 556098160 gb ESO86812.1	7	24	7	20	450
gi 556104876 gb ESO93528.1	7	14	13	21	231
gi 556105302 gb ESO93954.1	7	43	7	63	1043
gi 556116273 gb ESP04925.1	7	35	7	34	311
gi 556116376 gb ESP05028.1	7	19	7	20	235

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 557010651 ref XP_006006018.1	7	18	7	8	202
gi 620958561 ref XP_007664363.1	7	29	7	38	461
gi 150251390 gb ABR68007.1	6	12	6	47	406
gi 152032020 gb ABS28869.1	6	22	6	86	888
gi 226478902 emb CAX72946.1	6	21	7	12	198
gi 325296727 ref NP_001191598.1	6	25	6	7	79
gi 346987844 gb AEO51768.1	6	15	7	30	372
gi 405952673 gb EKC20457.1	6	31	7	35	409
gi 405976077 gb EKC40598.1	6	10	6	11	196
gi 421975936 gb AFX73000.1	6	21	6	52	499
gi 443708569 gb ELU03646.1	6	22	6	18	417
gi 524865616 ref XP_005089636.1	6	20	6	13	358
gi 524865670 ref XP_005089662.1	6	52	6	117	2465
gi 524866780 ref XP_005090207.1	6	18	6	14	357
gi 524869199 ref XP_005091396.1	6	21	6	16	265
gi 524871402 ref XP_005092474.1	6	17	6	28	475
gi 524872725 ref XP_005093120.1	6	16	6	13	253
gi 524874372 ref XP_005093922.1	6	14	6	8	108
gi 524876525 ref XP_005094978.1	6	53	8	21	243
gi 524876525 ref XP_005094978.1	6	42	11	76	1207
gi 524879821 ref XP_005096582.1	6	28	6	15	463
gi 524880980 ref XP_005097146.1	6	19	6	18	218
gi 524881119 ref XP_005097215.1	6	13	6	12	140
gi 524881385 ref XP_005097344.1	6	34	6	12	227
gi 524881557 ref XP_005097429.1	6	29	6	33	740
gi 524881651 ref XP_005097475.1	6	12	6	12	230
gi 524882420 ref XP_005097854.1	6	38	7	38	846
gi 524882484 ref XP_005097885.1	6	19	6	20	329
gi 524883534 ref XP_005098394.1	6	41	6	28	409
gi 524883626 ref XP_005098440.1	6	51	6	48	1209
gi 524885581 ref XP_005099396.1	6	30	6	13	139
gi 524886950 ref XP_005100065.1	6	15	6	12	183
gi 524887258 ref XP_005100217.1	6	16	6	11	178
gi 524888067 ref XP_005100610.1	6	6	6	23	241
gi 524889482 ref XP_005101296.1	6	12	6	16	329
gi 524889720 ref XP_005101414.1	6	34	6	16	331
gi 524889992 ref XP_005101546.1	6	34	6	21	776
gi 524892640 ref XP_005102846.1	6	44	6	13	253
gi 524894013 ref XP_005103514.1	6	14	6	15	345
gi 524894778 ref XP_005103881.1	6	13	6	10	174
gi 524899715 ref XP_005106286.1	6	15	6	12	308

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524901536 ref XP_005107174.1	6	14	6	9	95
gi 524901909 ref XP_005107358.1	6	36	6	9	130
gi 524902626 ref XP_005107542.1	6	31	6	10	154
gi 524903367 ref XP_005107884.1	6	19	6	10	213
gi 524907772 ref XP_005108998.1	6	12	6	14	349
gi 524908509 ref XP_005109349.1	6	38	6	21	438
gi 524909258 ref XP_005109678.1	6	20	6	14	307
gi 524911513 ref XP_005110619.1	6	33	6	11	194
gi 524912191 ref XP_005110947.1	6	59	6	12	189
gi 524913881 ref XP_005111763.1	6	11	6	10	173
gi 524914515 ref XP_005112072.1	6	28	6	10	75
gi 524916601 ref XP_005113074.1	6	6	6	33	564
gi 526117405 ref NP_001267760.1	6	68	13	195	1773
gi 555704960 gb ESO08193.1	6	53	6	103	2396
gi 556103805 gb ESO92457.1	6	22	6	7	102
gi 556105072 gb ESO93724.1	6	24	6	12	397
gi 556105072 gb ESO93724.1	6	15	6	29	511
gi 556113526 gb ESP02178.1	6	11	6	10	145
gi 556113946 gb ESP02598.1	6	11	6	9	174
gi 556114460 gb ESP03112.1	6	23	6	12	205
gi 1174755 sp P43689.1 TPM2_BIOGL	5	39	12	63	965
gi 325197162 ref NP_001191425.1	5	29	5	5	101
gi 325296839 ref NP_001191661.1	5	31	5	6	93
gi 405970776 gb EKC35652.1	5	34	5	20	217
gi 42559558 sp O97192.1 TPM_HELAS	5	51	19	277	4877
gi 431831576 gb AGA92562.1	5	39	5	9	193
gi 468860736 gb AGH32328.1	5	8	5	7	139
gi 4929136 gb AAD33872.1 AF141348_1	5	19	5	10	219
gi 524864626 ref XP_005089148.1	5	16	5	8	134
gi 524866300 ref XP_005089972.1	5	19	5	16	269
gi 524866556 ref XP_005090099.1	5	9	5	7	124
gi 524870028 ref XP_005091800.1	5	11	5	5	141
gi 524870886 ref XP_005092223.1	5	48	5	19	609
gi 524873315 ref XP_005093407.1	5	27	5	8	186
gi 524874171 ref XP_005093825.1	5	26	5	11	244
gi 524874376 ref XP_005093924.1	5	72	8	53	1019
gi 524876523 ref XP_005094977.1	5	67	5	15	182
gi 524877040 ref XP_005095227.1	5	29	5	21	544
gi 524879616 ref XP_005096482.1	5	18	5	7	154
gi 524879821 ref XP_005096582.1	5	24	5	14	269
gi 524879933 ref XP_005096636.1	5	19	5	8	109

List of proteins identified in 2-DE proteomics study

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524880339 ref XP_005096834.1	5	16	5	8	163
gi 524880906 ref XP_005097111.1	5	20	5	13	279
gi 524881344 ref XP_005097324.1	5	18	5	10	191
gi 524882120 ref XP_005097705.1	5	16	5	11	176
gi 524882774 ref XP_005098023.1	5	20	5	7	146
gi 524883534 ref XP_005098394.1	5	34	5	43	1018
gi 524885619 ref XP_005099415.1	5	20	5	18	350
gi 524886181 ref XP_005099688.1	5	43	9	52	1391
gi 524886263 ref XP_005099728.1	5	11	5	8	158
gi 524886299 ref XP_005099746.1	5	19	5	23	437
gi 524886483 ref XP_005099835.1	5	32	5	13	196
gi 524886570 ref XP_005099878.1	5	43	5	28	835
gi 524886963 ref XP_005100071.1	5	6	5	6	118
gi 524888229 ref XP_005100690.1	5	8	5	10	297
gi 524889251 ref XP_005101186.1	5	27	5	17	542
gi 524891178 ref XP_005102125.1	5	17	5	16	203
gi 524891634 ref XP_005102350.1	5	16	5	7	115
gi 524892341 ref XP_005102699.1	5	22	7	33	289
gi 524892819 ref XP_005102934.1	5	10	5	6	118
gi 524893098 ref XP_005103069.1	5	31	5	23	251
gi 524893793 ref XP_005103405.1	5	9	5	8	244
gi 524894261 ref XP_005103632.1	5	5	5	6	157
gi 524894818 ref XP_005103900.1	5	25	6	26	126
gi 524895639 ref XP_005104301.1	5	19	5	6	116
gi 524898799 ref XP_005105845.1	5	3	5	5	93
gi 524898960 ref XP_005105922.1	5	10	5	9	162
gi 524902421 ref XP_005107462.1	5	39	5	13	222
gi 524902628 ref XP_005107543.1	5	31	5	7	150
gi 524902929 ref XP_005107684.1	5	17	5	15	191
gi 524903247 ref XP_005107829.1	5	13	5	22	407
gi 524908412 ref XP_005109303.1	5	11	5	6	84
gi 524912853 ref XP_005111263.1	5	11	5	5	86
gi 524912929 ref XP_005111300.1	5	28	5	12	169
gi 524915771 ref XP_005112676.1	5	32	7	37	499
gi 524917285 ref XP_005113402.1	5	15	5	14	230
gi 556100834 gb ESO89486.1	5	23	5	7	131
gi 556101047 gb ESO89699.1	5	4	5	5	94
gi 556113799 gb ESP02451.1	5	16	5	10	220
gi 556114398 gb ESP03050.1	5	16	5	11	93
gi 556114539 gb ESP03191.1	5	12	5	11	224
gi 602626279 ref XP_007420100.1	5	42	6	117	2630

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 608090766 gb EZG30020.1	5	23	5	17	271
gi 632946707 ref XP_007888691.1	5	13	5	16	273
gi 104162064 emb CAK19329.1	4	42	4	67	1304
gi 126697398 gb ABO26656.1	4	17	4	10	235
gi 150251392 gb ABR68008.1	4	8	4	10	202
gi 158997667 gb ABW86957.1	4	35	4	51	1030
gi 325296803 ref NP_001191642.1	4	32	4	8	126
gi 325296995 ref NP_001191510.1	4	38	4	7	102
gi 325297040 ref NP_001191523.1	4	11	4	8	128
gi 325504381 emb CBX41690.1	4	24	4	17	170
gi 405951515 gb EKC19421.1	4	12	4	7	233
gi 405959696 gb EKC25703.1	4	13	4	42	672
gi 405972472 gb EKC37239.1	4	8	4	8	221
gi 405973356 gb EKC38075.1	4	18	4	7	81
gi 4249742 gb AAD13782.1	4	11	4	4	81
gi 425595558 sp O97192.1 TPM_HELAS	4	26	9	19	264
gi 443694956 gb ELT95974.1	4	26	4	25	280
gi 4504279 ref NP_002098.1	4	19	4	28	131
gi 51105058 gb AAT97089.1	4	8	4	6	107
gi 521024335 gb EPQ06123.1	4	15	4	14	182
gi 524863988 ref XP_005088833.1	4	20	4	6	148
gi 524867613 ref XP_005090615.1	4	4	4	8	92
gi 524869393 ref XP_005091491.1	4	17	4	13	178
gi 524870300 ref XP_005091935.1	4	20	4	7	159
gi 524870848 ref XP_005092204.1	4	6	4	6	144
gi 524870945 ref XP_005092251.1	4	32	4	9	157
gi 524871178 ref XP_005092365.1	4	18	4	7	189
gi 524871180 ref XP_005092366.1	4	28	4	9	128
gi 524871774 ref XP_005092658.1	4	23	4	8	94
gi 524872775 ref XP_005093144.1	4	8	4	7	167
gi 524873604 ref XP_005093546.1	4	31	4	11	184
gi 524874374 ref XP_005093923.1	4	40	4	8	156
gi 524874452 ref XP_005093961.1	4	21	4	13	276
gi 524875207 ref XP_005094334.1	4	16	4	6	85
gi 524875325 ref XP_005094393.1	4	33	4	7	122
gi 524876601 ref XP_005095013.1	4	15	4	8	168
gi 524876605 ref XP_005095015.1	4	7	4	6	150
gi 524877129 ref XP_005095271.1	4	24	4	11	123
gi 524877675 ref XP_005095535.1	4	6	4	16	337
gi 524879957 ref XP_005096647.1	4	17	4	10	223
gi 524883640 ref XP_005098447.1	4	11	4	20	423

List of proteins identified in 2-DE proteomics study

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524883762 ref XP_005098507.1	4	13	4	5	88
gi 524884099 ref XP_005098673.1	4	6	4	5	93
gi 524884550 ref XP_005098894.1	4	12	4	6	98
gi 524886285 ref XP_005099739.1	4	42	4	13	120
gi 524886483 ref XP_005099835.1	4	30	4	14	182
gi 524888539 ref XP_005100839.1	4	9	4	6	102
gi 524890683 ref XP_005101885.1	4	3	4	5	78
gi 524892173 ref XP_005102616.1	4	15	4	10	177
gi 524892281 ref XP_005102670.1	4	4	4	14	285
gi 524895031 ref XP_005104004.1	4	9	4	6	109
gi 524895469 ref XP_005104217.1	4	13	4	7	97
gi 524896519 ref XP_005104733.1	4	23	6	15	153
gi 524896868 ref XP_005104905.1	4	18	4	5	151
gi 524899361 ref XP_005106113.1	4	8	4	6	74
gi 524900084 ref XP_005106466.1	4	10	4	5	148
gi 524900809 ref XP_005106822.1	4	14	4	8	146
gi 524900993 ref XP_005106911.1	4	16	4	6	116
gi 524901357 ref XP_005107086.1	4	30	4	5	73
gi 524901394 ref XP_005107104.1	4	23	4	12	567
gi 524906293 ref XP_005108469.1	4	24	4	11	242
gi 524907491 ref XP_005108862.1	4	15	4	11	407
gi 524907576 ref XP_005108904.1	4	24	4	22	366
gi 524910025 ref XP_005109900.1	4	31	4	6	116
gi 524910373 ref XP_005110066.1	4	10	4	14	250
gi 524910660 ref XP_005110205.1	4	9	4	5	92
gi 524910895 ref XP_005110321.1	4	13	4	4	90
gi 524911650 ref XP_005110685.1	4	12	4	11	256
gi 524913419 ref XP_005111540.1	4	3	4	5	91
gi 524913668 ref XP_005111659.1	4	18	4	7	164
gi 524914469 ref XP_005112049.1	4	17	4	7	188
gi 524915677 ref XP_005112631.1	4	23	4	19	312
gi 526117431 ref NP_001267767.1	4	31	4	11	161
gi 556093552 gb ESO82205.1	4	22	4	18	272
gi 556105117 gb ESO93769.1	4	14	5	24	165
gi 556106038 gb ESO94690.1	4	11	4	6	146
gi 556110210 gb ESO98862.1	4	20	4	6	161
gi 556110891 gb ESO99542.1	4	22	4	4	99
gi 556111437 gb ESP00089.1	4	18	4	5	60
gi 577029907 gb AHH81788.1	4	24	4	8	98
gi 625230674 ref XP_007606483.1	4	53	4	10	129
gi 639127132 gb AIA24462.1	4	24	4	5	169

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 1174755 sp P43689.1 TPM2_BIOGL	3	15	3	3	52
gi 126697410 gb ABO26662.1	3	18	3	34	727
gi 133874728 gb ABO40828.1	3	13	3	4	61
gi 150251390 gb ABR68007.1	3	9	3	8	176
gi 150251390 gb ABR68007.1	3	6	3	3	86
gi 150251390 gb ABR68007.1	3	18	3	3	45
gi 210076789 gb ACJ06748.1	3	27	3	10	203
gi 220979902 emb CAQ64775.1	3	14	3	4	86
gi 253317417 gb ACT22633.1	3	47	6	48	407
gi 260821662 ref XP_002606151.1	3	16	5	11	225
gi 260821662 ref XP_002606151.1	3	13	5	31	339
gi 325197162 ref NP_001191425.1	3	13	3	8	133
gi 325296743 ref NP_001191607.1	3	6	3	11	208
gi 346987840 gb AEO51766.1	3	13	3	60	1298
gi 374718481 gb AEZ67032.1	3	6	3	8	92
gi 405965644 gb EKC31006.1	3	14	3	4	89
gi 405965813 gb EKC31167.1	3	14	3	7	79
gi 443705209 gb ELU01864.1	3	22	3	7	163
gi 443729497 gb ELU15363.1	3	7	3	3	87
gi 472824679 dbj BAN09032.1	3	39	4	29	223
gi 524864532 ref XP_005089101.1	3	10	3	6	111
gi 524864665 ref XP_005089167.1	3	18	3	7	92
gi 524865020 ref XP_005089342.1	3	33	3	8	270
gi 524870191 ref XP_005091881.1	3	11	3	14	328
gi 524870815 ref XP_005092188.1	3	11	3	5	117
gi 524871621 ref XP_005092582.1	3	8	4	7	107
gi 524871621 ref XP_005092582.1	3	14	4	4	59
gi 524872832 ref XP_005093172.1	3	8	3	12	294
gi 524872846 ref XP_005093178.1	3	25	3	15	219
gi 524872934 ref XP_005093222.1	3	23	3	6	134
gi 524873892 ref XP_005093688.1	3	33	3	10	383
gi 524874194 ref XP_005093836.1	3	9	3	8	168
gi 524874346 ref XP_005093909.1	3	19	3	7	92
gi 524875029 ref XP_005094246.1	3	20	3	4	132
gi 524875666 ref XP_005094557.1	3	33	3	4	66
gi 524876523 ref XP_005094977.1	3	40	4	5	74
gi 524876525 ref XP_005094978.1	3	33	6	13	221
gi 524877265 ref XP_005095338.1	3	8	3	6	101
gi 524877372 ref XP_005095389.1	3	20	3	15	126
gi 524877853 ref XP_005095623.1	3	23	3	43	726
gi 524879487 ref XP_005096418.1	3	17	3	4	66

List of proteins identified in 2-DE proteomics study

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524879771 ref XP_005096557.1	3	4	3	4	106
gi 524880082 ref XP_005096709.1	3	7	3	7	222
gi 524881557 ref XP_005097429.1	3	11	3	4	85
gi 524881663 ref XP_005097481.1	3	13	3	3	71
gi 524881663 ref XP_005097481.1	3	11	3	4	65
gi 524881729 ref XP_005097514.1	3	7	3	7	155
gi 524881799 ref XP_005097549.1	3	14	3	4	48
gi 524881948 ref XP_005097620.1	3	5	3	8	267
gi 524882305 ref XP_005097797.1	3	22	3	9	266
gi 524882420 ref XP_005097854.1	3	8	4	23	462
gi 524882460 ref XP_005097873.1	3	28	3	16	385
gi 524882748 ref XP_005098012.1	3	7	3	4	110
gi 524884884 ref XP_005099056.1	3	17	3	4	53
gi 524885143 ref XP_005099182.1	3	8	3	4	62
gi 524885272 ref XP_005099245.1	3	28	3	27	904
gi 524885593 ref XP_005099402.1	3	15	3	53	1426
gi 524886568 ref XP_005099877.1	3	25	3	7	79
gi 524888237 ref XP_005100694.1	3	23	3	3	105
gi 524889096 ref XP_005101109.1	3	18	3	9	102
gi 524889992 ref XP_005101546.1	3	43	3	13	405
gi 524891741 ref XP_005102403.1	3	8	3	6	82
gi 524893468 ref XP_005103247.1	3	23	3	4	110
gi 524894608 ref XP_005103800.1	3	5	3	6	139
gi 524895637 ref XP_005104300.1	3	12	3	11	245
gi 524896746 ref XP_005104844.1	3	22	3	6	146
gi 524898165 ref XP_005105535.1	3	7	3	3	75
gi 524898449 ref XP_005105673.1	3	16	3	9	208
gi 524898531 ref XP_005105713.1	3	11	3	8	128
gi 524898670 ref XP_005105781.1	3	10	3	3	96
gi 524898743 ref XP_005105817.1	3	26	3	5	86
gi 524899675 ref XP_005106266.1	3	12	3	3	85
gi 524899675 ref XP_005106266.1	3	11	3	7	125
gi 524900353 ref XP_005106597.1	3	7	3	5	71
gi 524900386 ref XP_005106613.1	3	8	3	3	95
gi 524900516 ref XP_005106678.1	3	9	3	11	245
gi 524903383 ref XP_005107892.1	3	20	3	7	123
gi 524903438 ref XP_005107919.1	3	12	3	3	87
gi 524905999 ref XP_005108354.1	3	18	3	8	147
gi 524906745 ref XP_005108595.1	3	20	4	7	193
gi 524907151 ref XP_005108701.1	3	6	3	4	121
gi 524907406 ref XP_005108821.1	3	11	3	4	80

List of proteins identified in 2-DE proteomics study

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524907727 ref XP_005108978.1	3	11	3	7	204
gi 524907998 ref XP_005109108.1	3	10	3	5	88
gi 524909038 ref XP_005109605.1	3	10	3	9	272
gi 524910167 ref XP_005109966.1	3	11	3	10	197
gi 524910867 ref XP_005110307.1	3	9	3	10	204
gi 524911945 ref XP_005110830.1	3	24	3	5	79
gi 524912291 ref XP_005110996.1	3	31	3	6	150
gi 524912715 ref XP_005111198.1	3	4	3	5	62
gi 524912715 ref XP_005111198.1	3	4	3	3	62
gi 524912919 ref XP_005111295.1	3	6	3	4	67
gi 524913016 ref XP_005111342.1	3	14	3	3	83
gi 524913109 ref XP_005111388.1	3	16	3	3	59
gi 524914862 ref XP_005112240.1	3	13	3	4	70
gi 524915405 ref XP_005112503.1	3	21	3	24	598
gi 524915787 ref XP_005112684.1	3	26	3	3	101
gi 524915990 ref XP_005112779.1	3	11	3	5	96
gi 524916974 ref XP_005113254.1	3	22	3	6	126
gi 54020890 ref NP_001005689.1	3	11	3	12	146
gi 555930987 emb CDJ08105.1	3	35	3	4	82
gi 556094374 gb ESO83027.1	3	8	3	4	72
gi 556098443 gb ESO87095.1	3	8	3	4	78
gi 556101526 gb ESO90178.1	3	9	3	8	149
gi 556106043 gb ESO94695.1	3	9	3	7	111
gi 556106190 gb ESO94842.1	3	20	5	8	119
gi 556109149 gb ESO97801.1	3	16	4	30	460
gi 556110831 gb ESO99482.1	3	23	3	21	264
gi 556113071 gb ESP01723.1	3	9	3	3	51
gi 556114539 gb ESP03191.1	3	3	3	6	88
gi 556116375 gb ESP05027.1	3	8	3	8	110
gi 556116442 gb ESP05094.1	3	32	3	20	958
gi 583994656 ref XP_006792469.1	3	28	3	6	74
gi 585193165 ref XP_006747803.1	3	7	3	8	158
gi 608096069 gb EZG34785.1	3	17	3	5	102
gi 646720608 gb KDR22269.1	3	30	3	6	165
gi 93209532 gb ABF00124.1	3	21	3	29	402
gi 115111533 gb ABI84147.1	2	6	2	4	77
gi 119568248 gb EAW47863.1	2	7	2	2	36
gi 126697398 gb ABO26656.1	2	10	2	2	50
gi 149032103 gb EDL87015.1	2	11	3	10	185
gi 149627356 ref XP_001508693.1	2	14	2	5	115
gi 150251390 gb ABR68007.1	2	13	2	4	66

List of proteins identified in 2-DE proteomics study

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 206597719 gb ACI15835.1	2	22	2	2	67
gi 2073142 dbj BAA19861.1	2	10	2	7	134
gi 2073142 dbj BAA19861.1	2	11	2	4	81
gi 2073146 dbj BAA19863.1	2	27	2	15	502
gi 210076789 gb ACJ06748.1	2	16	2	6	142
gi 218683625 gb ACL00841.1	2	9	2	2	52
gi 225717556 gb ACO14624.1	2	11	2	3	70
gi 260781473 ref XP_002585834.1	2	14	2	3	52
gi 325296803 ref NP_001191642.1	2	18	2	3	73
gi 325296883 ref NP_001191473.1	2	10	2	2	53
gi 325296989 ref NP_001191508.1	2	10	2	3	54
gi 325297146 ref NP_001191582.1	2	17	6	10	209
gi 333449383 gb AEF33378.1	2	17	2	3	104
gi 346987844 gb AEO51768.1	2	8	4	7	161
gi 348577231 ref XP_003474388.1	2	10	2	2	59
gi 38043955 emb CAD83837.1	2	8	2	4	52
gi 38043955 emb CAD83837.1	2	11	2	11	132
gi 390349570 ref XP_782388.3	2	67	9	33	583
gi 402227989 gb AFQ35982.1	2	8	2	2	47
gi 405964935 gb EKC30373.1	2	3	2	2	39
gi 405965644 gb EKC31006.1	2	9	2	4	58
gi 405967885 gb EKC33004.1	2	10	2	2	66
gi 405969794 gb EKC34745.1	2	15	2	5	130
gi 405969853 gb EKC34799.1	2	4	2	2	46
gi 405972489 gb EKC37255.1	2	10	2	2	81
gi 405974628 gb EKC39257.1	2	5	2	9	247
gi 405975533 gb EKC40092.1	2	10	2	2	92
gi 405978690 gb EKC43060.1	2	4	2	2	78
gi 406679464 gb AFS50734.1	2	14	2	5	94
gi 425595558 sp O97192.1 TPM_HELAS	2	19	7	15	259
gi 42560362 gb AAS20336.1	2	31	5	209	4192
gi 46276960 gb AAS86676.1	2	7	2	3	62
gi 524863982 ref XP_005088830.1	2	25	2	3	52
gi 524864066 ref XP_005088871.1	2	14	2	5	59
gi 524864582 ref XP_005089126.1	2	9	2	2	50
gi 524864622 ref XP_005089146.1	2	6	2	8	226
gi 524865813 ref XP_005089733.1	2	10	2	12	155
gi 524865891 ref XP_005089771.1	2	7	2	6	107
gi 524866978 ref XP_005090304.1	2	23	2	2	57
gi 524867549 ref XP_005090585.1	2	10	2	2	60
gi 524868395 ref XP_005091000.1	2	6	2	2	53

List of proteins identified in 2-DE proteomics study

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524868787 ref XP_005091193.1	2	7	2	2	62
gi 524870722 ref XP_005092143.1	2	10	2	4	82
gi 524871066 ref XP_005092309.1	2	5	2	2	44
gi 524872436 ref XP_005092980.1	2	5	2	4	111
gi 524872436 ref XP_005092980.1	2	6	2	2	126
gi 524872672 ref XP_005093096.1	2	19	2	7	100
gi 524872801 ref XP_005093157.1	2	8	2	6	83
gi 524872805 ref XP_005093159.1	2	9	2	2	56
gi 524873087 ref XP_005093297.1	2	6	2	14	203
gi 524873464 ref XP_005093478.1	2	13	2	2	31
gi 524873886 ref XP_005093685.1	2	3	2	3	88
gi 524873894 ref XP_005093689.1	2	28	2	4	71
gi 524874093 ref XP_005093787.1	2	12	2	3	59
gi 524874148 ref XP_005093814.1	2	11	2	2	54
gi 524874148 ref XP_005093814.1	2	7	2	13	125
gi 524874190 ref XP_005093834.1	2	6	2	2	62
gi 524874657 ref XP_005094063.1	2	2	2	8	192
gi 524874940 ref XP_005094202.1	2	12	2	7	85
gi 524875029 ref XP_005094246.1	2	15	2	2	86
gi 524875093 ref XP_005094277.1	2	21	2	4	111
gi 524875173 ref XP_005094317.1	2	4	2	2	60
gi 524875239 ref XP_005094350.1	2	5	2	4	82
gi 524875329 ref XP_005094395.1	2	26	2	4	128
gi 524876118 ref XP_005094778.1	2	10	2	4	111
gi 524876523 ref XP_005094977.1	2	22	2	4	68
gi 524876523 ref XP_005094977.1	2	31	2	11	183
gi 524876525 ref XP_005094978.1	2	14	3	26	467
gi 524876525 ref XP_005094978.1	2	51	3	9	272
gi 524876560 ref XP_005094995.1	2	3	2	2	87
gi 524876618 ref XP_005095021.1	2	8	2	3	95
gi 524876864 ref XP_005095141.1	2	6	2	5	126
gi 524877098 ref XP_005095256.1	2	12	2	4	45
gi 524877853 ref XP_005095623.1	2	10	3	11	291
gi 524878165 ref XP_005095772.1	2	5	2	3	97
gi 524879594 ref XP_005096471.1	2	6	2	3	43
gi 524879801 ref XP_005096572.1	2	6	2	4	75
gi 524881413 ref XP_005097358.1	2	8	2	2	44
gi 524882436 ref XP_005097862.1	2	5	2	9	207
gi 524882460 ref XP_005097873.1	2	19	2	3	72
gi 524882968 ref XP_005098119.1	2	29	2	6	148
gi 524883188 ref XP_005098227.1	2	12	2	3	89

List of proteins identified in 2-DE proteomics study

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524883558 ref XP_005098406.1	2	5	2	2	60
gi 524884444 ref XP_005098842.1	2	10	2	5	79
gi 524884503 ref XP_005098871.1	2	15	2	2	58
gi 524884830 ref XP_005099031.1	2	6	2	2	86
gi 524885197 ref XP_005099209.1	2	8	2	2	44
gi 524885355 ref XP_005099286.1	2	6	2	3	64
gi 524885703 ref XP_005099456.1	2	3	2	6	59
gi 524885955 ref XP_005099578.1	2	5	2	3	55
gi 524886261 ref XP_005099727.1	2	3	2	2	85
gi 524886263 ref XP_005099728.1	2	11	2	5	79
gi 524886662 ref XP_005099924.1	2	6	2	2	48
gi 524886707 ref XP_005099946.1	2	5	2	7	151
gi 524887384 ref XP_005100278.1	2	3	2	2	40
gi 524887465 ref XP_005100318.1	2	30	2	4	103
gi 524887465 ref XP_005100318.1	2	18	2	4	117
gi 524887498 ref XP_005100334.1	2	4	2	2	57
gi 524887580 ref XP_005100374.1	2	5	2	3	72
gi 524889104 ref XP_005101113.1	2	5	2	2	65
gi 524889992 ref XP_005101546.1	2	10	2	2	75
gi 524890376 ref XP_005101735.1	2	7	2	2	54
gi 524890561 ref XP_005101826.1	2	5	2	5	171
gi 524890653 ref XP_005101870.1	2	11	2	3	98
gi 524890724 ref XP_005101905.1	2	60	18	38	463
gi 524890794 ref XP_005101939.1	2	12	2	2	85
gi 524891426 ref XP_005102246.1	2	6	2	2	58
gi 524891574 ref XP_005102320.1	2	12	2	2	77
gi 524892285 ref XP_005102672.1	2	5	2	3	72
gi 524893098 ref XP_005103069.1	2	17	2	8	208
gi 524893405 ref XP_005103218.1	2	8	2	3	101
gi 524893572 ref XP_005103295.1	2	5	2	5	91
gi 524893741 ref XP_005103379.1	2	15	2	2	56
gi 524893900 ref XP_005103458.1	2	4	2	3	62
gi 524893900 ref XP_005103458.1	2	4	2	3	65
gi 524894650 ref XP_005103820.1	2	9	2	3	94
gi 524894780 ref XP_005103882.1	2	4	2	2	55
gi 524895699 ref XP_005104331.1	2	4	2	2	43
gi 524896048 ref XP_005104503.1	2	12	2	3	48
gi 524896189 ref XP_005104572.1	2	5	2	2	49
gi 524896597 ref XP_005104771.1	2	11	2	2	74
gi 524896719 ref XP_005104831.1	2	25	2	5	139
gi 524896719 ref XP_005104831.1	2	15	2	2	44

List of proteins identified in 2-DE proteomics study

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524897390 ref XP_005105156.1	2	23	2	2	40
gi 524898060 ref XP_005105484.1	2	7	2	2	71
gi 524898265 ref XP_005105584.1	2	7	2	2	52
gi 524898815 ref XP_005105852.1	2	34	2	4	182
gi 524899012 ref XP_005105947.1	2	8	2	2	60
gi 524899021 ref XP_005105951.1	2	8	2	4	59
gi 524899091 ref XP_005105986.1	2	18	2	2	44
gi 524899882 ref XP_005106367.1	2	7	2	2	64
gi 524900256 ref XP_005106549.1	2	17	2	4	40
gi 524900270 ref XP_005106556.1	2	15	2	20	169
gi 524900390 ref XP_005106615.1	2	9	4	10	205
gi 524900699 ref XP_005106768.1	2	4	2	7	105
gi 524901519 ref XP_005107166.1	2	11	2	3	63
gi 524901804 ref XP_005107306.1	2	14	2	5	101
gi 524902448 ref XP_005107474.1	2	7	2	3	67
gi 524902931 ref XP_005107685.1	2	7	2	2	46
gi 524903201 ref XP_005107810.1	2	1	2	2	68
gi 524903262 ref XP_005107836.1	2	7	2	2	44
gi 524903293 ref XP_005107850.1	2	8	2	7	154
gi 524906293 ref XP_005108469.1	2	34	2	4	136
gi 524906293 ref XP_005108469.1	2	11	2	7	172
gi 524906745 ref XP_005108595.1	2	14	2	2	45
gi 524906745 ref XP_005108595.1	2	13	2	2	44
gi 524906745 ref XP_005108595.1	2	7	2	2	59
gi 524906932 ref XP_005108623.1	2	2	2	3	65
gi 524907576 ref XP_005108904.1	2	21	2	21	332
gi 524907590 ref XP_005108911.1	2	9	2	2	42
gi 524908282 ref XP_005109244.1	2	18	2	4	160
gi 524908284 ref XP_005109245.1	2	6	2	2	37
gi 524909838 ref XP_005109815.1	2	13	2	3	91
gi 524909848 ref XP_005109820.1	2	7	2	2	52
gi 524909938 ref XP_005109861.1	2	5	2	2	42
gi 524910284 ref XP_005110023.1	2	5	2	3	52
gi 524910373 ref XP_005110066.1	2	3	2	3	54
gi 524910383 ref XP_005110071.1	2	11	2	3	111
gi 524910825 ref XP_005110286.1	2	11	2	8	116
gi 524911477 ref XP_005110601.1	2	31	2	2	59
gi 524911866 ref XP_005110793.1	2	15	2	3	63
gi 524911952 ref XP_005110833.1	2	13	2	2	59
gi 524913773 ref XP_005111711.1	2	6	2	2	65
gi 524913818 ref XP_005111733.1	2	5	2	6	143

List of proteins identified in 2-DE proteomics study

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524914187 ref XP_005111912.1	2	6	2	3	94
gi 524915372 ref XP_005112487.1	2	10	2	2	42
gi 524916029 ref XP_005112797.1	2	7	2	2	89
gi 524916381 ref XP_005112969.1	2	9	2	2	42
gi 524916499 ref XP_005113026.1	2	7	2	9	226
gi 524916593 ref XP_005113070.1	2	9	2	2	44
gi 524917273 ref XP_005113396.1	2	11	2	4	152
gi 526117361 ref NP_001267749.1	2	13	2	137	1145
gi 556093891 gb ESO82544.1	2	9	2	2	42
gi 556095216 gb ESO83869.1	2	8	2	5	180
gi 556097528 gb ESO86180.1	2	7	2	4	90
gi 556098160 gb ESO86812.1	2	7	2	11	226
gi 556100167 gb ESO88819.1	2	17	2	7	84
gi 556101117 gb ESO89769.1	2	9	2	2	49
gi 556106190 gb ESO94842.1	2	8	2	2	74
gi 556106555 gb ESO95207.1	2	3	2	2	64
gi 556107537 gb ESO96189.1	2	4	2	3	111
gi 556108606 gb ESO97258.1	2	17	2	4	82
gi 556111890 gb ESP00542.1	2	12	2	2	56
gi 556113049 gb ESP01701.1	2	5	2	2	56
gi 556114871 gb ESP03523.1	2	23	2	3	73
gi 556115860 gb ESP04512.1	2	4	2	5	64
gi 556116519 gb ESP05171.1	2	11	2	2	81
gi 556729406 ref XP_005960416.1	2	9	2	5	135
gi 568291352 gb ETN79106.1	2	13	2	38	164
gi 589107123 ref XP_006965584.1	2	18	2	2	64
gi 597867153 gb EYC16545.1	2	24	3	26	693
gi 68272049 gb AA Y89300.1	2	12	2	6	108
gi 773571 emb CAA60122.1	2	20	2	3	110
gi 773571 emb CAA60122.1	2	7	3	4	130
gi 85753308 gb AAG49533.3	2	9	2	13	185
gi 994895 gb AAB34494.1	2	10	2	2	81
gi 170596696 ref XP_001902862.1	1	17	2	11	257
gi 241151391 ref XP_002406661.1	1	14	5	8	209
gi 2827905 gb AAB99911.1	1	4	2	5	150
gi 405952673 gb EKC20457.1	1	11	2	3	57
gi 51105058 gb AAT97089.1	1	11	2	2	75
gi 524876525 ref XP_005094978.1	1	13	2	3	58
gi 524885593 ref XP_005099402.1	1	15	3	10	349
gi 524906745 ref XP_005108595.1	1	7	2	3	74
gi 537705579 dbj BAN81895.1	1	30	4	28	503

List of proteins identified in 2-DE proteomics study

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 642099995 emb CDQ77143.1 	1	25	3	3	112

8.3. Table 2. The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524911570 ref XP_005110646.1	150	48	152	489	9983
gi 318609972 dbj BAJ61596.1	112	82	117	3270	88132
gi 524888508 ref XP_005100824.1	92	63	116	847	16625
gi 524907580 ref XP_005108906.1	76	43	77	167	3386
gi 524896814 ref XP_005104878.1	69	50	101	292	6076
gi 346987840 gb AEO51766.1	63	44	93	489	10429
gi 524911094 ref XP_005110416.1	59	40	59	126	2582
gi 127773 sp P24733.1 MYS_ARGIR	50	69	176	1022	23056
gi 524907580 ref XP_005108906.1	49	28	49	157	4164
gi 556096138 gb ESO84790.1	46	74	46	352	7897
gi 524903201 ref XP_005107810.1	45	37	70	164	4119
gi 6682319 emb CAB64662.1	45	67	68	713	17930
gi 124233 sp P22488.2 IFEA_HELAS	43	77	59	847	16905
gi 325297144 ref NP_001191581.1	43	62	46	159	3236
gi 524896814 ref XP_005104878.1	43	38	44	92	1511
gi 524916601 ref XP_005113074.1	40	42	45	118	2572
gi 524866478 ref XP_005090060.1	37	76	37	455	8975
gi 556110866 gb ESO99517.1	37	59	90	456	10538
gi 346987840 gb AEO51766.1	36	19	38	105	2142
gi 468860736 gb AGH32328.1	35	49	35	119	2372
gi 524868522 ref XP_005091063.1	35	42	40	103	2396
gi 556106555 gb ESO95207.1	35	43	52	106	1843
gi 346987840 gb AEO51766.1	33	44	33	183	3319
gi 524875601 ref XP_005094526.1	33	71	43	92	1886
gi 126697420 gb ABO26667.1	32	59	32	93	1806
gi 524867613 ref XP_005090615.1	32	41	32	42	1015
gi 524906491 ref XP_005108543.1	32	51	33	101	1838
gi 220067263 gb ACL79834.1	31	86	31	293	6253
gi 472824675 dbj BAN09030.1	31	62	31	255	4213
gi 524896814 ref XP_005104878.1	31	62	32	118	2214
gi 524894502 ref XP_005103748.1	30	57	30	227	5014
gi 556096092 gb ESO84744.1	30	46	30	136	2794
gi 150251392 gb ABR68008.1	29	51	29	151	3179
gi 218683627 gb ACL00842.1	29	65	29	117	3208
gi 556113526 gb ESP02178.1	29	62	29	75	1397
gi 73695954 gb AAZ80783.1	29	66	29	752	17215
gi 524881119 ref XP_005097215.1	28	56	28	90	1496
gi 556106555 gb ESO95207.1	28	49	43	113	2377

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

gi 127773 sp P24733.1 MYS_ARGIR	27	50	92	659	19586
Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 325296743 ref NP_001191607.1	27	67	34	104	1852
gi 524869485 ref XP_005091535.1	27	57	27	131	2189
gi 524903383 ref XP_005107892.1	27	78	28	362	10496
gi 524916025 ref XP_005112795.1	27	77	28	309	8062
gi 405960426 gb EKC26351.1	26	69	26	121	2932
gi 524888358 ref XP_005100750.1	26	56	27	103	2155
gi 524895745 ref XP_005104354.1	26	63	35	514	13237
gi 524896814 ref XP_005104878.1	26	27	26	73	1752
gi 296837083 gb ADH59421.1	25	72	35	441	9017
gi 305689815 gb ADM64337.1	25	69	25	341	9279
gi 524887592 ref XP_005100380.1	25	72	25	93	1941
gi 556113946 gb ESP02598.1	25	46	25	117	2548
gi 385145402 emb CCG28026.1	24	80	33	2612	54460
gi 42559558 sp O97192.1 TPM_HELAS	24	83	41	319	6065
gi 524886261 ref XP_005099727.1	24	49	43	322	7707
gi 74912853 sp Q6QUW1.1 RGP51_LYMST	24	70	31	199	3590
gi 241273677 ref XP_002406620.1	23	66	26	73	1343
gi 524881811 ref XP_005097555.1	23	73	23	178	4157
gi 556097340 gb ESO85992.1	23	47	23	88	2392
gi 556104876 gb ESO93528.1	23	76	60	255	5495
gi 556106555 gb ESO95207.1	23	35	36	66	1146
gi 585719304 ref XP_006826038.1	23	42	69	165	3915
gi 405950185 gb EKC18187.1	22	9	22	31	518
gi 524881729 ref XP_005097514.1	22	51	22	37	804
gi 524883288 ref XP_005098277.1	22	49	22	60	1107
gi 524885898 ref XP_005099550.1	22	49	22	30	493
gi 524908996 ref XP_005109585.1	22	56	22	86	1619
gi 524913881 ref XP_005111763.1	22	47	26	42	966
gi 150251390 gb ABR68007.1	21	51	21	130	2865
gi 524884737 ref XP_005098987.1	21	57	22	70	1211
gi 524906932 ref XP_005108623.1	21	36	27	37	609
gi 556104876 gb ESO93528.1	21	66	52	235	5405
Unknown	21	73	21	218	4285
gi 150251390 gb ABR68007.1	20	54	20	114	2371
gi 296190822 ref XP_002743358.1	20	39	21	52	1118
gi 321472487 gb EFX83457.1	20	34	20	55	985
gi 524867613 ref XP_005090615.1	20	15	20	35	993
gi 524882748 ref XP_005098012.1	20	70	39	196	3973
gi 524885581 ref XP_005099396.1	20	71	20	106	1541
gi 346987844 gb AEO51768.1	16	36	17	86	1625
gi 524886263 ref XP_005099728.1	20	31	20	57	1002

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524892329 ref XP_005102693.1	20	34	20	24	564
gi 524893765 ref XP_005103391.1	20	42	20	24	403
gi 524894608 ref XP_005103800.1	20	36	20	27	391
gi 524896519 ref XP_005104733.1	20	75	23	95	1222
gi 524896638 ref XP_005104791.1	20	41	20	61	1294
gi 524913881 ref XP_005111763.1	20	47	22	81	2223
gi 556097683 gb ESO86335.1	20	56	20	77	1512
gi 119630315 gb EAX09910.1	19	55	21	93	2079
gi 524867573 ref XP_005090597.1	19	66	19	68	1544
gi 524868093 ref XP_005090853.1	19	28	19	28	612
gi 524870848 ref XP_005092204.1	19	30	19	21	342
gi 524872922 ref XP_005093216.1	19	43	20	75	1573
gi 524890683 ref XP_005101885.1	19	20	19	25	454
gi 524909114 ref XP_005109609.1	19	92	23	592	11227
gi 524916452 ref XP_005113003.1	19	55	19	139	1903
gi 526117385 ref NP_001267755.1	19	73	20	133	3576
gi 556101476 gb ESO90128.1	19	39	26	30	540
gi 773571 emb CAA60122.1	19	45	26	36	744
gi 126697362 gb ABO26638.1	18	83	18	181	3463
gi 325297052 ref NP_001191530.1	18	82	29	351	7647
gi 443692429 gb ELT94023.1	18	64	18	112	2336
gi 524878973 ref XP_005096166.1	18	68	18	94	2172
gi 524884921 ref XP_005099073.1	18	56	18	807	18649
gi 524885520 ref XP_005099366.1	18	82	18	188	2297
gi 524889526 ref XP_005101318.1	18	44	18	48	1113
gi 524892924 ref XP_005102985.1	18	58	18	186	4977
gi 524912440 ref XP_005111068.1	18	64	18	307	6517
gi 524916601 ref XP_005113074.1	18	25	20	85	2449
gi 526117385 ref NP_001267755.1	18	79	19	353	10785
gi 556097528 gb ESO86180.1	18	61	18	49	1345
gi 556105302 gb ESO93954.1	18	79	21	296	5016
gi 524871134 ref XP_005092343.1	17	44	17	58	1385
gi 524880064 ref XP_005096700.1	17	47	17	30	620
gi 524886817 ref XP_005100000.1	17	43	17	80	1717
gi 524898799 ref XP_005105845.1	17	13	17	35	996
gi 524902448 ref XP_005107474.1	17	47	17	124	2385
gi 524912715 ref XP_005111198.1	17	45	30	66	1607
gi 54020890 ref NP_001005689.1	17	66	17	94	1679
gi 241151391 ref XP_002406661.1	16	75	23	122	1778
gi 325297040 ref NP_001191523.1	16	71	18	71	1048
gi 524907576 ref XP_005108904.1	12	64	12	147	3211
gi 524913044 ref XP_005111356.1	12	47	12	39	804

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524894602 ref XP_005103797.1	12	23	12	14	306
gi 524894780 ref XP_005103882.1	12	28	12	27	493
gi 524871930 ref XP_005092735.1	16	22	16	33	692
gi 524872578 ref XP_005093050.1	16	37	32	62	1549
gi 524874065 ref XP_005093774.1	16	69	16	26	674
gi 524877675 ref XP_005095535.1	16	26	16	48	1145
gi 524879388 ref XP_005096370.1	16	56	16	56	1570
gi 524882748 ref XP_005098012.1	16	62	37	152	3098
gi 524885854 ref XP_005099530.1	16	73	16	236	3887
gi 524896250 ref XP_005104602.1	16	54	16	44	567
gi 524907151 ref XP_005108701.1	16	38	16	33	669
gi 524908412 ref XP_005109303.1	16	36	16	20	307
gi 524909189 ref XP_005109645.1	16	26	16	23	369
gi 524911954 ref XP_005110834.1	16	43	19	61	1188
gi 524915110 ref XP_005112361.1	16	39	16	45	698
gi 524916507 ref XP_005113030.1	16	61	19	65	1077
gi 54020890 ref NP_001005689.1	16	59	16	101	2534
gi 555688717 gb ESN91949.1	16	79	20	305	6418
gi 556100505 gb ESO89157.1	16	42	17	42	758
gi 556101476 gb ESO90128.1	16	29	18	46	752
gi 585193165 ref XP_006747803.1	16	46	18	99	1856
gi 147905746 ref NP_001080566.1	15	84	29	182	3912
gi 405953142 gb EKC20858.1	15	47	15	43	711
gi 524866272 ref XP_005089958.1	15	55	15	22	319
gi 524867757 ref XP_005090686.1	15	29	29	53	1203
gi 524868057 ref XP_005090835.1	15	12	15	20	329
gi 524877903 ref XP_005095647.1	15	8	15	31	707
gi 524883990 ref XP_005098620.1	15	67	15	88	2385
gi 524886695 ref XP_005099940.1	15	59	15	39	846
gi 524888067 ref XP_005100610.1	15	20	15	44	764
gi 524891424 ref XP_005102245.1	15	34	15	29	660
gi 524895953 ref XP_005104456.1	15	15	15	22	293
gi 524897671 ref XP_005105294.1	15	33	15	56	1426
gi 524897999 ref XP_005105454.1	15	54	16	63	1580
gi 524900390 ref XP_005106615.1	15	37	15	51	1204
gi 524900673 ref XP_005106755.1	15	66	15	96	1921
gi 524906359 ref XP_005108493.1	15	37	15	26	540
gi 524909114 ref XP_005109609.1	15	89	19	426	7452
gi 526117405 ref NP_001267760.1	15	82	15	216	3211
gi 556101047 gb ESO89699.1	15	17	22	28	568
gi 556110891 gb ESO99542.1	15	68	15	105	1857
gi 1174755 sp P43689.1 TPM2_BIOGL	14	75	28	84	1265

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 150251390 gb ABR68007.1	14	42	15	61	1945
gi 270268071 gb ACZ65578.1	14	45	14	30	654
gi 346987844 gb AEO51768.1	14	14	15	23	405
gi 405978809 gb EKC43171.1	14	32	14	26	634
gi 51105058 gb AAT97089.1	14	62	19	39	841
gi 513137512 gb AGO20698.1	14	40	14	42	1115
gi 524867763 ref XP_005090689.1	14	16	29	34	785
gi 524873169 ref XP_005093336.1	14	39	14	24	475
gi 524873538 ref XP_005093514.1	14	34	14	21	503
gi 524874194 ref XP_005093836.1	14	49	14	29	497
gi 524874655 ref XP_005094062.1	14	14	14	53	1275
gi 524875149 ref XP_005094305.1	14	27	14	24	431
gi 524875601 ref XP_005094526.1	14	42	24	34	770
gi 524876391 ref XP_005094913.1	14	61	14	49	775
gi 524878504 ref XP_005095936.1	14	84	16	45	751
gi 524879388 ref XP_005096370.1	14	50	14	21	435
gi 524879616 ref XP_005096482.1	14	63	14	35	883
gi 524880840 ref XP_005097078.1	14	52	14	24	381
gi 524883140 ref XP_005098203.1	14	31	14	17	408
gi 524883762 ref XP_005098507.1	14	55	14	39	796
gi 524884451 ref XP_005098845.1	14	45	14	27	420
gi 524886299 ref XP_005099746.1	14	64	14	29	520
gi 524892173 ref XP_005102616.1	14	49	14	40	635
gi 524892201 ref XP_005102630.1	14	20	15	16	311
gi 524892341 ref XP_005102699.1	14	72	17	206	3140
gi 524894013 ref XP_005103514.1	14	43	16	50	904
gi 524896519 ref XP_005104733.1	14	63	18	100	1103
gi 524903522 ref XP_005107960.1	14	46	14	75	1603
gi 524907533 ref XP_005108883.1	14	46	14	49	861
gi 524911418 ref XP_005110573.1	14	21	15	24	361
gi 556101117 gb ESO89769.1	14	62	14	43	542
gi 556113049 gb ESP01701.1	14	36	14	27	652
gi 556113964 gb ESP02616.1	14	31	14	33	602
gi 556114539 gb ESP03191.1	14	32	14	29	463
gi 620958561 ref XP_007664363.1	14	76	14	61	1074
gi 2073142 dbj BAA19861.1	13	62	13	278	4823
gi 325297068 ref NP_001191538.1	13	27	14	31	534
gi 325297094 ref NP_001191555.1	13	49	13	28	680
gi 42559558 sp O97192.1 TPM_HELAS	13	67	25	81	1441
gi 464524 sp Q05974.1 RAB1A_LYMST	13	74	13	62	1166
gi 521024335 gb EPQ06123.1	13	50	13	153	2421
gi 524913109 ref XP_005111388.1	12	68	12	62	1080

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524864622 ref XP_005089146.1	13	36	13	36	939
gi 524871621 ref XP_005092582.1	13	53	22	93	1930
gi 524878388 ref XP_005095881.1	13	32	13	27	461
gi 524881606 ref XP_005097453.1	13	51	13	33	740
gi 524882422 ref XP_005097855.1	13	53	13	66	1480
gi 524882436 ref XP_005097862.1	13	30	13	26	394
gi 524885248 ref XP_005099233.1	13	82	13	27	333
gi 524886261 ref XP_005099727.1	13	20	13	23	562
gi 524889482 ref XP_005101296.1	13	36	13	48	912
gi 524890176 ref XP_005101637.1	13	35	13	22	385
gi 524891426 ref XP_005102246.1	13	52	13	34	658
gi 524891679 ref XP_005102372.1	13	47	13	47	1023
gi 524897999 ref XP_005105454.1	13	45	14	43	1484
gi 524898259 ref XP_005105581.1	13	51	13	60	667
gi 524917285 ref XP_005113402.1	13	52	13	42	482
gi 556095158 gb ESO83811.1	13	42	16	47	1133
gi 556098588 gb ESO87240.1	13	53	13	24	472
gi 556106043 gb ESO94695.1	13	38	13	17	314
gi 556106306 gb ESO94958.1	13	33	13	22	510
gi 556116376 gb ESP05028.1	13	44	15	30	628
gi 126697410 gb ABO26662.1	12	56	12	207	5349
gi 32169292 emb CAD89885.1	12	39	12	26	232
gi 325296847 ref NP_001191665.1	12	27	13	18	400
gi 325296855 ref NP_001191459.1	12	27	20	25	638
gi 325296989 ref NP_001191508.1	12	65	14	43	562
gi 405960204 gb EKC26145.1	12	17	12	18	378
gi 405964935 gb EKC30373.1	12	26	12	24	491
gi 443708569 gb ELU03646.1	12	50	12	26	614
gi 524865670 ref XP_005089662.1	12	86	14	259	5953
gi 524865955 ref XP_005089803.1	12	55	13	88	1019
gi 524866304 ref XP_005089974.1	12	55	12	34	897
gi 524866556 ref XP_005090099.1	12	24	12	24	450
gi 524867514 ref XP_005090568.1	12	37	12	25	546
gi 524869199 ref XP_005091396.1	12	50	12	30	883
gi 524870028 ref XP_005091800.1	12	36	12	27	651
gi 524872674 ref XP_005093097.1	12	38	12	17	341
gi 524873087 ref XP_005093297.1	12	49	12	35	626
gi 524873600 ref XP_005093544.1	12	63	12	49	421
gi 524875771 ref XP_005094609.1	12	26	12	30	639
gi 524875985 ref XP_005094714.1	12	16	12	31	430
gi 524876605 ref XP_005095015.1	12	26	12	21	322
gi 524879594 ref XP_005096471.1	12	53	12	28	728

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524880339 ref XP_005096834.1	12	45	12	24	333
gi 524881417 ref XP_005097360.1	12	53	12	38	777
gi 524881980 ref XP_005097636.1	12	40	12	14	285
gi 524882968 ref XP_005098119.1	12	31	12	18	417
gi 524884737 ref XP_005098987.1	12	39	13	33	689
gi 524885197 ref XP_005099209.1	12	48	12	14	233
gi 524885795 ref XP_005099501.1	12	67	37	239	5797
gi 524886263 ref XP_005099728.1	12	34	12	51	2015
gi 524891584 ref XP_005102325.1	12	38	12	17	343
gi 524892179 ref XP_005102619.1	12	27	12	20	291
gi 524892285 ref XP_005102672.1	12	34	13	27	666
gi 524892819 ref XP_005102934.1	12	27	12	32	679
gi 524893900 ref XP_005103458.1	12	33	15	41	703
gi 524917241 ref XP_005113381.1	12	43	12	139	2324
gi 556095158 gb ESO83811.1	12	43	16	38	1143
gi 556095567 gb ESO84220.1	12	43	13	31	735
gi 556105117 gb ESO93769.1	12	66	16	97	1689
gi 556106555 gb ESO95207.1	12	39	35	57	1041
gi 556116427 gb ESP05079.1	12	50	15	58	1157
gi 564743500 gb AHC02701.1	12	81	33	148	2594
gi 773571 emb CAA60122.1	12	43	19	21	313
gi 152003985 gb ABS19816.1	11	32	20	53	1552
gi 152032020 gb ABS28869.1	11	39	11	56	584
gi 168252891 gb ACA23131.1	11	66	11	68	1495
gi 291235073 ref XP_002737469.1	11	41	11	26	327
gi 325296875 ref NP_001191469.1	11	64	11	27	295
gi 325296883 ref NP_001191473.1	11	59	11	26	349
gi 325297040 ref NP_001191523.1	11	36	13	19	302
gi 351709298 gb EHB12217.1	11	53	11	265	4017
gi 405972472 gb EKC37239.1	11	27	12	22	486
gi 425595558 sp O97192.1 TPM_HELAS	11	53	17	41	700
gi 524864482 ref XP_005089076.1	11	32	11	13	266
gi 524865670 ref XP_005089662.1	11	76	15	137	2808
gi 524866934 ref XP_005090282.1	11	12	12	24	452
gi 524867613 ref XP_005090615.1	11	19	11	23	578
gi 524870191 ref XP_005091881.1	11	45	11	48	770
gi 524870300 ref XP_005091935.1	11	56	11	27	433
gi 524870886 ref XP_005092223.1	11	67	11	145	3783
gi 524871621 ref XP_005092582.1	11	51	22	62	1330
gi 524871885 ref XP_005092713.1	11	35	11	20	276
gi 524871930 ref XP_005092735.1	11	23	11	20	479
gi 524874055 ref XP_005093769.1	11	19	11	15	221

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524874414 ref XP_005093942.1	11	25	11	31	543
gi 524878451 ref XP_005095912.1	11	16	11	16	359
gi 524882484 ref XP_005097885.1	11	38	12	28	657
gi 524884293 ref XP_005098768.1	11	20	11	21	322
gi 524884630 ref XP_005098934.1	11	39	11	45	1118
gi 524885193 ref XP_005099207.1	11	22	11	23	489
gi 524887384 ref XP_005100278.1	11	13	11	25	446
gi 524890066 ref XP_005101582.1	11	28	12	23	506
gi 524890120 ref XP_005101609.1	11	39	11	17	201
gi 524893793 ref XP_005103405.1	11	30	11	27	649
gi 524895031 ref XP_005104004.1	11	27	11	33	659
gi 524895893 ref XP_005104427.1	11	22	11	14	284
gi 524896814 ref XP_005104878.1	11	21	21	40	768
gi 524897983 ref XP_005105446.1	11	30	12	24	428
gi 524898231 ref XP_005105567.1	11	31	11	16	291
gi 524898324 ref XP_005105612.1	11	31	11	12	214
gi 524901897 ref XP_005107352.1	11	55	11	28	646
gi 524901909 ref XP_005107358.1	11	54	11	30	679
gi 524906745 ref XP_005108595.1	11	63	11	41	894
gi 524907590 ref XP_005108911.1	11	59	11	21	439
gi 524908062 ref XP_005109139.1	11	59	11	21	559
gi 524909004 ref XP_005109589.1	11	61	14	77	907
gi 524909038 ref XP_005109605.1	11	34	11	32	717
gi 524909838 ref XP_005109815.1	11	34	11	20	479
gi 524910373 ref XP_005110066.1	11	28	13	20	392
gi 524911396 ref XP_005110562.1	11	44	11	27	611
gi 524911954 ref XP_005110834.1	11	41	21	146	2639
gi 524913818 ref XP_005111733.1	11	51	13	38	668
gi 524914469 ref XP_005112049.1	11	46	11	38	490
gi 524914829 ref XP_005112224.1	11	41	11	43	1184
gi 524915789 ref XP_005112685.1	11	28	11	19	408
gi 556096567 gb ESO85219.1	11	20	11	17	215
gi 556116184 gb ESP04836.1	11	54	11	27	729
gi 602626279 ref XP_007420100.1	11	75	11	92	1724
gi 126697356 gb ABO26635.1	10	55	11	39	535
gi 13177628 gb AAK14899.1	10	26	10	17	385
gi 150251390 gb ABR68007.1	10	38	10	59	1260
gi 150251390 gb ABR68007.1	10	25	10	10	270
gi 2073142 dbj BAA19861.1	10	40	10	40	486
gi 328933188 gb AEB70965.1	10	37	10	23	503
gi 346987840 gb AEO51766.1	10	33	10	29	458
gi 346987844 gb AEO51768.1	10	32	12	80	1288

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 383859188 ref XP_003705078.1	10	12	10	15	407
gi 405952673 gb EKC20457.1	10	59	11	58	706
gi 405962978 gb EKC28598.1	10	43	10	14	300
gi 405970776 gb EKC35652.1	10	71	10	84	1224
gi 443718889 gb ELU09307.1	10	21	10	12	124
gi 524864076 ref XP_005088876.1	10	16	10	20	526
gi 524866376 ref XP_005090010.1	10	29	10	26	917
gi 524866912 ref XP_005090271.1	10	36	10	31	493
gi 524866934 ref XP_005090282.1	10	14	11	12	402
gi 524870722 ref XP_005092143.1	10	51	10	40	914
gi 524874657 ref XP_005094063.1	10	11	10	33	853
gi 524874769 ref XP_005094119.1	10	24	10	15	255
gi 524875291 ref XP_005094376.1	10	16	10	16	297
gi 524875728 ref XP_005094588.1	10	47	10	27	259
gi 524876046 ref XP_005094743.1	10	19	10	21	306
gi 524876098 ref XP_005094768.1	10	26	10	20	365
gi 524876712 ref XP_005095067.1	10	18	10	12	261
gi 524878458 ref XP_005095915.1	10	63	10	43	907
gi 524881984 ref XP_005097638.1	10	75	10	83	2209
gi 524882420 ref XP_005097854.1	10	29	14	21	382
gi 524882717 ref XP_005097997.1	10	38	10	31	636
gi 524883534 ref XP_005098394.1	10	57	10	31	476
gi 524884406 ref XP_005098823.1	10	30	10	15	116
gi 524885703 ref XP_005099456.1	10	23	10	17	358
gi 524889104 ref XP_005101113.1	10	30	10	19	348
gi 524892967 ref XP_005103005.1	10	65	10	172	2717
gi 524893098 ref XP_005103069.1	10	83	10	65	1260
gi 524895406 ref XP_005104186.1	10	8	10	17	268
gi 524895699 ref XP_005104331.1	10	22	10	17	166
gi 524895953 ref XP_005104456.1	10	13	10	13	213
gi 524896874 ref XP_005104908.1	10	51	10	29	215
gi 524898060 ref XP_005105484.1	10	27	10	16	376
gi 524898203 ref XP_005105554.1	10	15	10	10	140
gi 524899582 ref XP_005106220.1	10	45	10	110	2701
gi 524900516 ref XP_005106678.1	10	58	10	186	2701
gi 524900699 ref XP_005106768.1	10	36	10	19	413
gi 524901536 ref XP_005107174.1	10	28	12	45	878
gi 524908509 ref XP_005109349.1	10	55	10	53	745
gi 524909848 ref XP_005109820.1	10	49	10	36	455
gi 524910025 ref XP_005109900.1	10	89	10	38	982
gi 524910869 ref XP_005110308.1	10	36	10	14	287
gi 556095567 gb ESO84220.1	10	28	11	28	653

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 556103805 gb ESO92457.1	10	48	10	29	691
gi 556107537 gb ESO96189.1	10	22	10	24	620
gi 556113071 gb ESP01723.1	10	30	10	16	354
gi 556116427 gb ESP05079.1	10	43	13	36	696
gi 556116519 gb ESP05171.1	10	53	10	34	597
gi 560896664 ref XP_006174798.1	10	28	10	20	450
gi 994895 gb AAB34494.1	10	40	12	26	557
gi 150251390 gb ABR68007.1	9	30	10	25	689
gi 2073142 dbj BAA19861.1	9	70	9	126	2816
gi 226478902 emb CAX72946.1	9	75	22	234	5014
gi 260821662 ref XP_002606151.1	9	59	21	110	2702
gi 325297146 ref NP_001191582.1	9	39	9	16	379
gi 346987840 gb AEO51766.1	9	32	12	56	1360
gi 405951515 gb EKC19421.1	9	35	9	15	414
gi 405959696 gb EKC25703.1	9	29	9	14	265
gi 405965813 gb EKC31167.1	9	45	9	22	378
gi 405966469 gb EKC31748.1	9	45	9	13	385
gi 405970946 gb EKC35808.1	9	59	9	17	455
gi 405974628 gb EKC39257.1	9	32	9	52	1144
gi 51105064 gb AAT97092.1	9	33	9	12	242
gi 524865140 ref XP_005089402.1	9	52	9	39	776
gi 524866272 ref XP_005089958.1	9	35	9	21	426
gi 524868977 ref XP_005091286.1	9	38	9	36	453
gi 524869203 ref XP_005091398.1	9	17	9	11	137
gi 524869203 ref XP_005091398.1	9	13	9	10	159
gi 524869207 ref XP_005091400.1	9	10	10	24	358
gi 524871180 ref XP_005092366.1	9	46	9	22	384
gi 524871402 ref XP_005092474.1	9	30	10	27	729
gi 524872186 ref XP_005092857.1	9	16	9	12	362
gi 524872286 ref XP_005092906.1	9	21	9	17	265
gi 524876525 ref XP_005094978.1	9	45	15	43	803
gi 524876525 ref XP_005094978.1	9	60	18	44	1036
gi 524877795 ref XP_005095595.1	9	28	9	20	564
gi 524878165 ref XP_005095772.1	9	27	9	12	222
gi 524878167 ref XP_005095773.1	9	16	9	19	278
gi 524879801 ref XP_005096572.1	9	34	9	16	456
gi 524880507 ref XP_005096916.1	9	60	9	23	403
gi 524881016 ref XP_005097164.1	9	11	9	12	293
gi 524882305 ref XP_005097797.1	9	59	9	86	1955
gi 524882414 ref XP_005097851.1	9	45	10	15	105
gi 524882774 ref XP_005098023.1	9	33	9	15	387
gi 524883140 ref XP_005098203.1	9	24	9	14	288

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524884878 ref XP_005099053.1	9	39	9	21	385
gi 524885619 ref XP_005099415.1	9	51	10	25	526
gi 524885795 ref XP_005099501.1	9	70	16	94	1730
gi 524890066 ref XP_005101582.1	9	12	10	13	166
gi 524890599 ref XP_005101843.1	9	33	9	11	239
gi 524892644 ref XP_005102848.1	9	62	9	52	1179
gi 524893673 ref XP_005103345.1	9	18	9	9	220
gi 524895342 ref XP_005104156.1	9	30	9	13	239
gi 524895745 ref XP_005104354.1	9	73	108	591	13659
gi 524897639 ref XP_005105279.1	9	38	9	15	185
gi 524898449 ref XP_005105673.1	9	49	9	27	313
gi 524900084 ref XP_005106466.1	9	30	9	10	142
gi 524902881 ref XP_005107662.1	9	45	9	10	114
gi 524903367 ref XP_005107884.1	9	31	9	20	364
gi 524903438 ref XP_005107919.1	9	47	9	14	373
gi 524907042 ref XP_005108650.1	9	31	9	9	115
gi 524907491 ref XP_005108862.1	9	33	9	26	559
gi 524907610 ref XP_005108921.1	9	34	9	31	807
gi 524909004 ref XP_005109589.1	9	48	12	70	906
gi 524909258 ref XP_005109678.1	9	43	9	25	503
gi 524910373 ref XP_005110066.1	9	29	10	24	344
gi 524910383 ref XP_005110071.1	9	48	9	22	594
gi 524910825 ref XP_005110286.1	9	47	9	12	150
gi 524911947 ref XP_005110831.1	9	69	16	311	6806
gi 524911947 ref XP_005110831.1	9	64	17	57	801
gi 524912191 ref XP_005110947.1	9	67	9	20	251
gi 524912929 ref XP_005111300.1	9	77	9	105	2318
gi 524912990 ref XP_005111329.1	9	45	9	12	241
gi 524913818 ref XP_005111733.1	9	31	11	14	261
gi 524915771 ref XP_005112676.1	9	43	11	105	1654
gi 524916029 ref XP_005112797.1	9	35	9	46	992
gi 524916507 ref XP_005113030.1	9	45	12	14	230
gi 556093757 gb ESO82410.1	9	31	9	10	139
gi 556093891 gb ESO82544.1	9	47	9	10	168
gi 556097339 gb ESO85991.1	9	11	9	14	259
gi 556100834 gb ESO89486.1	9	42	9	20	468
gi 556101047 gb ESO89699.1	9	10	16	26	338
gi 556101476 gb ESO90128.1	9	32	17	21	400
gi 556103805 gb ESO92457.1	9	36	9	14	231
gi 556107233 gb ESO95885.1	9	27	9	33	905
gi 556109519 gb ESO98171.1	9	7	9	12	237
gi 556110708 gb ESO99359.1	9	35	9	19	297

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 556113321 gb ESP01973.1	9	32	9	18	175
gi 556114539 gb ESP03191.1	9	12	9	13	252
gi 577029907 gb AHH81788.1	9	58	9	19	275
gi 597788484 ref XP_007257657.1	9	35	10	17	112
gi 608090766 gb EZG30020.1	9	74	9	42	1408
gi 118425914 gb ABK90856.1	8	25	8	23	659
gi 150251392 gb ABR68008.1	8	37	8	62	1041
gi 158997667 gb ABW86957.1	8	47	8	196	3488
gi 2073142 dbj BAA19861.1	8	55	8	201	4262
gi 241556129 ref XP_002399605.1	8	51	8	19	286
gi 241835856 ref XP_002415077.1	8	41	8	15	149
gi 260781473 ref XP_002585834.1	8	62	8	37	665
gi 2827905 gb AAB99911.1	8	19	10	22	432
gi 325296853 ref NP_001191458.1	8	23	9	15	293
gi 325296989 ref NP_001191508.1	8	41	10	15	211
gi 326535853 gb ADZ76593.1	8	18	8	9	136
gi 346987840 gb AEO51766.1	8	34	10	107	2726
gi 375073675 gb AFA34396.1	8	58	8	22	355
gi 402227989 gb AFQ35982.1	8	47	8	25	512
gi 405957391 gb EKC23605.1	8	41	8	8	227
gi 405965644 gb EKC31006.1	8	36	9	15	347
gi 405968798 gb EKC33830.1	8	30	8	11	282
gi 405969732 gb EKC34685.1	8	18	8	17	253
gi 405969853 gb EKC34799.1	8	18	8	14	283
gi 405973356 gb EKC38075.1	8	43	9	11	186
gi 421975936 gb AFX73000.1	8	38	8	19	140
gi 42559558 sp O97192.1 TPM_HELAS	8	64	23	77	1135
gi 524863988 ref XP_005088833.1	8	52	8	13	255
gi 524864118 ref XP_005088897.1	8	60	8	11	87
gi 524864532 ref XP_005089101.1	8	26	8	14	231
gi 524866585 ref XP_005090113.1	8	21	8	9	131
gi 524866627 ref XP_005090132.1	8	48	8	13	178
gi 524867048 ref XP_005090337.1	8	62	9	19	260
gi 524867757 ref XP_005090686.1	8	11	19	26	550
gi 524868003 ref XP_005090808.1	8	36	10	44	780
gi 524869549 ref XP_005091567.1	8	24	31	57	1150
gi 524869987 ref XP_005091780.1	8	55	8	14	234
gi 524871178 ref XP_005092365.1	8	37	8	17	207
gi 524871453 ref XP_005092499.1	8	23	8	11	128
gi 524871609 ref XP_005092576.1	8	14	8	10	260
gi 524872616 ref XP_005093069.1	8	21	13	15	250
gi 524876525 ref XP_005094978.1	8	55	15	25	531

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524876974 ref XP_005095195.1	8	32	8	10	202
gi 524876984 ref XP_005095200.1	8	19	8	8	161
gi 524877031 ref XP_005095223.1	8	35	8	18	330
gi 524879910 ref XP_005096625.1	8	22	8	11	191
gi 524879933 ref XP_005096636.1	8	40	8	14	227
gi 524880906 ref XP_005097111.1	8	32	8	9	123
gi 524881047 ref XP_005097179.1	8	46	8	17	236
gi 524881809 ref XP_005097554.1	8	47	8	14	198
gi 524882120 ref XP_005097705.1	8	27	8	22	349
gi 524883534 ref XP_005098394.1	8	59	8	37	537
gi 524883626 ref XP_005098440.1	8	65	8	52	1123
gi 524884884 ref XP_005099056.1	8	55	8	19	185
gi 524885593 ref XP_005099402.1	8	48	9	34	1083
gi 524886540 ref XP_005099863.1	8	33	8	11	306
gi 524886950 ref XP_005100065.1	8	21	8	12	135
gi 524886963 ref XP_005100071.1	8	12	8	11	177
gi 524887258 ref XP_005100217.1	8	22	8	12	173
gi 524888539 ref XP_005100839.1	8	21	8	19	464
gi 524889887 ref XP_005101495.1	8	34	8	12	252
gi 524890204 ref XP_005101650.1	8	44	12	23	398
gi 524890839 ref XP_005101961.1	8	39	8	15	150
gi 524890899 ref XP_005101991.1	8	15	8	30	508
gi 524891306 ref XP_005102188.1	8	27	8	15	369
gi 524891634 ref XP_005102350.1	8	38	8	24	346
gi 524892640 ref XP_005102846.1	8	59	10	37	934
gi 524892640 ref XP_005102846.1	8	80	10	43	929
gi 524894544 ref XP_005103769.1	8	38	8	16	130
gi 524896746 ref XP_005104844.1	8	56	8	21	106
gi 524899180 ref XP_005106028.1	8	29	8	15	371
gi 524900516 ref XP_005106678.1	8	29	8	21	222
gi 524900516 ref XP_005106678.1	8	43	8	25	684
gi 524901804 ref XP_005107306.1	8	74	8	36	814
gi 524901804 ref XP_005107306.1	8	71	8	59	1213
gi 524903262 ref XP_005107836.1	8	31	8	13	120
gi 524903461 ref XP_005107930.1	8	48	8	19	326
gi 524906081 ref XP_005108389.1	8	25	8	13	250
gi 524910284 ref XP_005110023.1	8	16	8	11	243
gi 524910373 ref XP_005110066.1	8	23	10	32	573
gi 524910660 ref XP_005110205.1	8	22	8	19	410
gi 524910816 ref XP_005110282.1	8	27	20	35	694
gi 524911666 ref XP_005110693.1	8	16	8	11	248
gi 524913419 ref XP_005111540.1	8	7	9	43	1232

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524913818 ref XP_005111733.1	8	30	9	19	396
gi 524914230 ref XP_005111933.1	8	19	8	9	127
gi 524915771 ref XP_005112676.1	8	44	9	81	1337
gi 524916607 ref XP_005113077.1	8	41	8	26	345
gi 556094806 gb ESO83459.1	8	27	8	13	227
gi 556104219 gb ESO92871.1	8	11	8	8	176
gi 556105302 gb ESO93954.1	8	55	8	173	3394
gi 655453820 ref WP_028837045.1	8	38	8	16	266
gi 773571 emb CAA60122.1	8	16	8	9	153
gi 104162064 emb CAK19329.1	7	55	7	207	3529
gi 13177628 gb AAK14899.1	7	28	7	11	259
gi 13177628 gb AAK14899.1	7	22	7	14	411
gi 232136 sp P30683.3 GNAO_LYMST	7	19	7	12	101
gi 23394916 gb AAN31640.1 AF484963_1	7	29	7	12	152
gi 297186099 gb ADI24337.1	7	18	7	8	148
gi 325197136 ref NP_001191419.1	7	53	7	29	493
gi 325296995 ref NP_001191510.1	7	43	7	24	434
gi 325297068 ref NP_001191538.1	7	16	9	18	260
gi 348573378 ref XP_003472468.1	7	17	7	18	440
gi 38043955 emb CAD83837.1	7	53	7	19	236
gi 405967712 gb EKC32845.1	7	33	7	7	104
gi 405970416 gb EKC35324.1	7	4	7	11	283
gi 524864066 ref XP_005088871.1	7	44	7	23	306
gi 524864808 ref XP_005089238.1	7	28	7	12	130
gi 524867050 ref XP_005090338.1	7	45	8	19	208
gi 524868522 ref XP_005091063.1	7	16	15	16	253
gi 524868842 ref XP_005091219.1	7	36	7	24	407
gi 524869393 ref XP_005091491.1	7	28	7	10	136
gi 524870815 ref XP_005092188.1	7	39	7	14	226
gi 524871144 ref XP_005092348.1	7	24	7	9	156
gi 524871449 ref XP_005092497.1	7	14	7	10	162
gi 524871513 ref XP_005092529.1	7	28	8	9	181
gi 524871732 ref XP_005092637.1	7	6	7	21	450
gi 524871748 ref XP_005092645.1	7	61	7	24	452
gi 524872186 ref XP_005092857.1	7	21	8	9	242
gi 524872578 ref XP_005093050.1	7	30	23	46	1182
gi 524872801 ref XP_005093157.1	7	61	11	37	808
gi 524872832 ref XP_005093172.1	7	24	7	10	172
gi 524874093 ref XP_005093787.1	7	60	7	20	274
gi 524874093 ref XP_005093787.1	7	48	7	27	390
gi 524874372 ref XP_005093922.1	7	19	7	9	205
gi 524874414 ref XP_005093942.1	7	14	7	11	254

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524875029 ref XP_005094246.1	7	58	7	15	299
gi 524876118 ref XP_005094778.1	7	50	7	18	617
gi 524877040 ref XP_005095227.1	7	43	7	18	508
gi 524877054 ref XP_005095234.1	7	22	7	8	183
gi 524877903 ref XP_005095647.1	7	6	7	8	61
gi 524878669 ref XP_005096017.1	7	40	7	7	147
gi 524878983 ref XP_005096171.1	7	40	10	15	117
gi 524879223 ref XP_005096289.1	7	32	7	26	428
gi 524879541 ref XP_005096445.1	7	24	7	10	184
gi 524880248 ref XP_005096790.1	7	38	7	16	105
gi 524880725 ref XP_005097023.1	7	31	7	16	191
gi 524881413 ref XP_005097358.1	7	31	8	14	93
gi 524881743 ref XP_005097521.1	7	32	7	34	671
gi 524882420 ref XP_005097854.1	7	63	9	29	343
gi 524882956 ref XP_005098113.1	7	20	7	11	235
gi 524883548 ref XP_005098401.1	7	33	7	18	405
gi 524883721 ref XP_005098487.1	7	27	7	11	128
gi 524884503 ref XP_005098871.1	7	46	7	22	200
gi 524885821 ref XP_005099514.1	7	44	7	11	196
gi 524886408 ref XP_005099800.1	7	25	7	12	135
gi 524886658 ref XP_005099922.1	7	20	7	8	195
gi 524886715 ref XP_005099950.1	7	13	8	11	186
gi 524886950 ref XP_005100065.1	7	31	7	8	171
gi 524887715 ref XP_005100441.1	7	17	7	10	128
gi 524887883 ref XP_005100524.1	7	38	7	16	200
gi 524888089 ref XP_005100621.1	7	25	7	13	213
gi 524889772 ref XP_005101439.1	7	28	7	12	297
gi 524890066 ref XP_005101582.1	7	18	7	10	122
gi 524890221 ref XP_005101658.1	7	14	7	8	121
gi 524890263 ref XP_005101679.1	7	14	7	14	255
gi 524890869 ref XP_005101976.1	7	22	7	11	206
gi 524891914 ref XP_005102489.1	7	39	7	12	59
gi 524893468 ref XP_005103247.1	7	68	7	12	198
gi 524893612 ref XP_005103315.1	7	42	7	22	363
gi 524893715 ref XP_005103366.1	7	34	7	23	192
gi 524893900 ref XP_005103458.1	7	29	13	26	355
gi 524894261 ref XP_005103632.1	7	10	9	30	355
gi 524894818 ref XP_005103900.1	7	33	7	33	285
gi 524895745 ref XP_005104354.1	7	57	60	516	14983
gi 524895745 ref XP_005104354.1	7	75	81	648	16063
gi 524895745 ref XP_005104354.1	7	77	81	605	14610
gi 524896457 ref XP_005104703.1	7	35	7	32	567

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524897390 ref XP_005105156.1	7	81	7	23	215
gi 524898535 ref XP_005105715.1	7	35	8	11	134
gi 524898553 ref XP_005105724.1	7	10	7	12	323
gi 524899361 ref XP_005106113.1	7	21	7	10	184
gi 524900516 ref XP_005106678.1	7	28	7	14	139
gi 524900516 ref XP_005106678.1	7	39	8	14	212
gi 524900832 ref XP_005106833.1	7	31	11	18	403
gi 524901536 ref XP_005107174.1	7	22	9	37	797
gi 524903201 ref XP_005107810.1	7	29	47	80	2053
gi 524903293 ref XP_005107850.1	7	23	7	14	227
gi 524906293 ref XP_005108469.1	7	49	7	9	179
gi 524906745 ref XP_005108595.1	7	51	10	28	510
gi 524906932 ref XP_005108623.1	7	15	12	13	263
gi 524906932 ref XP_005108623.1	7	22	13	14	331
gi 524907047 ref XP_005108652.1	7	17	7	12	398
gi 524907590 ref XP_005108911.1	7	42	7	20	286
gi 524907772 ref XP_005108998.1	7	25	7	9	180
gi 524908561 ref XP_005109374.1	7	10	7	13	189
gi 524909434 ref XP_005109764.1	7	32	7	12	116
gi 524911952 ref XP_005110833.1	7	68	16	88	1738
gi 524912848 ref XP_005111261.1	7	29	7	12	150
gi 524913368 ref XP_005111515.1	7	42	7	16	374
gi 524913419 ref XP_005111540.1	7	7	8	33	1012
gi 524915657 ref XP_005112621.1	7	29	7	9	182
gi 524915986 ref XP_005112777.1	7	60	7	38	556
gi 524916381 ref XP_005112969.1	7	29	7	8	98
gi 524916593 ref XP_005113070.1	7	41	8	25	457
gi 556094817 gb ESO83470.1	7	9	7	10	137
gi 556097355 gb ESO86007.1	7	59	7	13	262
gi 556097653 gb ESO86305.1	7	20	7	7	87
gi 556097720 gb ESO86372.1	7	40	7	27	370
gi 556098160 gb ESO86812.1	7	39	10	26	441
gi 556100505 gb ESO89157.1	7	44	7	16	406
gi 556105072 gb ESO93724.1	7	14	7	19	431
gi 556111273 gb ESO99924.1	7	36	7	28	587
gi 556114460 gb ESP03112.1	7	31	7	13	215
gi 556116283 gb ESP04935.1	7	17	8	18	339
gi 568301578 gb ETN87131.1	7	25	7	7	138
gi 617524151 ref XP_007544814.1	7	35	7	15	333
gi 646714599 gb KDR18512.1	7	64	7	19	351
gi 74137592 dbj BAE35828.1	7	30	7	19	481
gi 126697398 gb ABO26656.1	6	29	6	10	148

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 254939725 gb ACT88125.1	6	56	6	10	148
gi 325197162 ref NP_001191425.1	6	29	6	11	232
gi 325296867 ref NP_001191465.1	6	29	6	12	262
gi 348577231 ref XP_003474388.1	6	30	6	27	454
gi 353236678 emb CCA68668.1	6	20	6	7	131
gi 405970946 gb EKC35808.1	6	64	6	30	872
gi 405975785 gb EKC40331.1	6	56	32	156	3232
gi 405976077 gb EKC40598.1	6	12	6	6	124
gi 42559558 sp O97192.1 TPM_HELAS	6	57	18	96	1911
gi 42559558 sp O97192.1 TPM_HELAS	6	73	28	69	876
gi 443733604 gb ELU17896.1	6	11	6	9	130
gi 524865020 ref XP_005089342.1	6	18	6	6	124
gi 524865020 ref XP_005089342.1	6	72	7	11	209
gi 524866113 ref XP_005089881.1	6	22	6	6	91
gi 524866188 ref XP_005089918.1	6	39	6	16	281
gi 524866538 ref XP_005090090.1	6	34	6	16	184
gi 524867549 ref XP_005090585.1	6	37	6	21	436
gi 524867701 ref XP_005090658.1	6	17	6	8	157
gi 524867763 ref XP_005090689.1	6	9	13	21	429
gi 524868003 ref XP_005090808.1	6	30	7	11	262
gi 524868892 ref XP_005091244.1	6	30	6	16	298
gi 524869453 ref XP_005091519.1	6	21	6	8	104
gi 524871619 ref XP_005092581.1	6	10	6	13	311
gi 524871674 ref XP_005092608.1	6	31	6	20	303
gi 524871774 ref XP_005092658.1	6	35	6	10	214
gi 524872460 ref XP_005092992.1	6	32	6	9	247
gi 524873900 ref XP_005093692.1	6	30	6	10	164
gi 524873980 ref XP_005093732.1	6	10	6	6	151
gi 524874093 ref XP_005093787.1	6	43	6	12	198
gi 524874148 ref XP_005093814.1	6	27	6	62	569
gi 524874969 ref XP_005094216.1	6	8	6	7	136
gi 524875029 ref XP_005094246.1	6	47	6	18	147
gi 524876286 ref XP_005094861.1	6	17	6	7	162
gi 524876396 ref XP_005094915.1	6	36	6	12	208
gi 524876525 ref XP_005094978.1	6	32	9	16	356
gi 524876525 ref XP_005094978.1	6	37	11	27	570
gi 524876525 ref XP_005094978.1	6	44	14	31	647
gi 524877265 ref XP_005095338.1	6	26	9	14	321
gi 524878037 ref XP_005095711.1	6	41	6	7	188
gi 524878114 ref XP_005095748.1	6	30	6	9	109
gi 524878247 ref XP_005095812.1	6	12	6	6	128
gi 524878596 ref XP_005095981.1	6	11	6	8	165

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524879065 ref XP_005096212.1	6	18	6	8	173
gi 524879771 ref XP_005096557.1	6	25	13	20	426
gi 524879821 ref XP_005096582.1	6	22	6	9	230
gi 524881344 ref XP_005097324.1	6	23	6	9	178
gi 524881651 ref XP_005097475.1	6	19	6	6	173
gi 524881865 ref XP_005097581.1	6	16	6	7	158
gi 524881948 ref XP_005097620.1	6	8	6	18	550
gi 524881984 ref XP_005097638.1	6	44	6	9	211
gi 524882215 ref XP_005097752.1	6	56	6	13	211
gi 524882332 ref XP_005097810.1	6	27	6	11	135
gi 524882460 ref XP_005097873.1	6	54	6	67	1357
gi 524883129 ref XP_005098198.1	6	38	6	37	548
gi 524883540 ref XP_005098397.1	6	35	6	17	326
gi 524883762 ref XP_005098507.1	6	26	6	8	131
gi 524884099 ref XP_005098673.1	6	12	6	10	152
gi 524884410 ref XP_005098825.1	6	40	6	10	172
gi 524884955 ref XP_005099089.1	6	29	6	14	391
gi 524885029 ref XP_005099126.1	6	32	6	8	45
gi 524885272 ref XP_005099245.1	6	63	6	39	1311
gi 524886028 ref XP_005099614.1	6	49	6	14	219
gi 524886707 ref XP_005099946.1	6	17	6	12	281
gi 524886950 ref XP_005100065.1	6	21	6	18	515
gi 524887138 ref XP_005100158.1	6	33	6	11	177
gi 524887169 ref XP_005100173.1	6	24	6	7	116
gi 524887214 ref XP_005100195.1	6	58	7	16	311
gi 524887319 ref XP_005100247.1	6	26	6	11	274
gi 524887394 ref XP_005100283.1	6	35	6	16	150
gi 524888756 ref XP_005100943.1	6	19	6	8	152
gi 524889720 ref XP_005101414.1	6	41	6	10	151
gi 524889734 ref XP_005101421.1	6	10	6	9	256
gi 524889992 ref XP_005101546.1	6	57	8	21	428
gi 524890794 ref XP_005101939.1	6	33	6	8	144
gi 524891363 ref XP_005102216.1	6	16	6	8	118
gi 524891574 ref XP_005102320.1	6	32	6	9	132
gi 524893281 ref XP_005103158.1	6	14	6	9	116
gi 524893352 ref XP_005103192.1	6	25	6	14	378
gi 524893513 ref XP_005103269.1	6	23	6	8	101
gi 524893741 ref XP_005103379.1	6	47	7	24	292
gi 524895469 ref XP_005104217.1	6	25	6	12	212
gi 524895639 ref XP_005104301.1	6	23	6	14	252
gi 524896057 ref XP_005104507.1	6	38	6	7	117
gi 524896189 ref XP_005104572.1	6	29	7	8	142

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524896407 ref XP_005104678.1	6	36	6	11	220
gi 524898531 ref XP_005105713.1	6	29	7	15	338
gi 524898859 ref XP_005105873.1	6	15	6	12	132
gi 524898960 ref XP_005105922.1	6	18	6	8	207
gi 524899002 ref XP_005105942.1	6	38	12	20	401
gi 524899021 ref XP_005105951.1	6	22	6	13	140
gi 524899091 ref XP_005105986.1	6	53	6	13	151
gi 524899489 ref XP_005106175.1	6	20	6	8	145
gi 524899675 ref XP_005106266.1	6	24	6	13	296
gi 524900390 ref XP_005106615.1	6	19	7	18	375
gi 524900832 ref XP_005106833.1	6	24	10	10	165
gi 524901283 ref XP_005107052.1	6	73	8	10	226
gi 524901464 ref XP_005107139.1	6	38	9	13	173
gi 524902427 ref XP_005107465.1	6	30	7	10	284
gi 524902628 ref XP_005107543.1	6	50	6	13	319
gi 524902755 ref XP_005107604.1	6	25	6	9	179
gi 524903041 ref XP_005107738.1	6	40	12	41	779
gi 524903201 ref XP_005107810.1	6	34	6	17	552
gi 524906862 ref XP_005108596.1	6	74	6	9	133
gi 524906990 ref XP_005108638.1	6	23	6	11	94
gi 524907042 ref XP_005108650.1	6	26	6	9	101
gi 524907370 ref XP_005108803.1	6	26	6	8	169
gi 524908000 ref XP_005109109.1	6	39	6	20	396
gi 524910373 ref XP_005110066.1	6	22	7	8	198
gi 524910744 ref XP_005110246.1	6	16	6	12	286
gi 524910825 ref XP_005110286.1	6	40	6	16	347
gi 524910895 ref XP_005110321.1	6	23	6	14	429
gi 524911513 ref XP_005110619.1	6	33	6	13	218
gi 524911570 ref XP_005110646.1	6	10	8	11	258
gi 524911576 ref XP_005110649.1	6	32	6	16	230
gi 524911939 ref XP_005110827.1	6	26	6	12	288
gi 524911952 ref XP_005110833.1	6	60	15	170	3006
gi 524912806 ref XP_005111241.1	6	17	6	6	129
gi 524912919 ref XP_005111295.1	6	17	6	7	179
gi 524912927 ref XP_005111299.1	6	71	6	27	531
gi 524912929 ref XP_005111300.1	6	54	6	42	713
gi 524913658 ref XP_005111654.1	6	60	6	42	819
gi 524914862 ref XP_005112240.1	6	30	6	16	392
gi 524915397 ref XP_005112499.1	6	20	6	14	356
gi 524916499 ref XP_005113026.1	6	31	6	15	377
gi 541039654 gb ERG79357.1	6	21	6	6	219
gi 555930987 emb CDJ08105.1	6	78	6	27	350

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 556093552 gb ESO82205.1	6	36	6	15	250
gi 556097053 gb ESO85705.1	6	25	6	8	289
gi 556098160 gb ESO86812.1	6	29	7	13	343
gi 556098160 gb ESO86812.1	6	30	7	42	742
gi 556100167 gb ESO88819.1	6	68	6	26	531
gi 556101526 gb ESO90178.1	6	27	6	13	399
gi 556101906 gb ESO90558.1	6	16	6	9	153
gi 556105072 gb ESO93724.1	6	38	6	23	505
gi 556109149 gb ESO97801.1	6	48	7	11	225
gi 556110831 gb ESO99482.1	6	44	6	49	619
gi 556111437 gb ESP00089.1	6	41	10	24	461
gi 556114369 gb ESP03021.1	6	6	6	8	190
gi 576693750 gb EUB57355.1	6	39	6	19	242
gi 72113881 ref XP_787111.1	6	82	6	16	438
gi 91077014 ref XP_966444.1	6	17	6	10	178
gi 126697408 gb ABO26661.1	5	33	5	8	180
gi 147903958 ref NP_001090965.1	5	69	5	25	580
gi 150251390 gb ABR68007.1	5	46	5	41	1156
gi 2073142 dbj BAA19861.1	5	44	5	16	273
gi 239509144 gb ACR81565.1	5	56	5	12	182
gi 325296803 ref NP_001191642.1	5	45	6	19	231
gi 325296881 ref NP_001191472.1	5	28	5	5	72
gi 325296937 ref NP_001191493.1	5	36	5	9	162
gi 325296986 ref NP_001191507.1	5	9	5	6	84
gi 346987840 gb AEO51766.1	5	42	8	22	535
gi 346987844 gb AEO51768.1	5	26	6	28	323
gi 350536823 ref NP_001233149.1	5	24	5	6	125
gi 351714959 gb EHB17878.1	5	17	6	7	193
gi 374093227 ref NP_001243365.1	5	63	5	17	304
gi 380022918 ref XP_003695281.1	5	44	5	21	398
gi 387157924 ref NP_001248345.1	5	23	5	10	71
gi 405959537 gb EKC25563.1	5	27	5	12	308
gi 405960428 gb EKC26353.1	5	18	5	29	618
gi 405961635 gb EKC27403.1	5	5	5	7	150
gi 405967797 gb EKC32924.1	5	46	8	34	1232
gi 405968748 gb EKC33788.1	5	37	5	14	389
gi 405970417 gb EKC35325.1	5	8	5	7	115
gi 405970417 gb EKC35325.1	5	13	5	9	109
gi 405972388 gb EKC37161.1	5	17	5	7	123
gi 405972489 gb EKC37255.1	5	22	5	5	114
gi 431831576 gb AGA92562.1	5	46	5	18	478
gi 443690943 gb ELT92936.1	5	25	5	6	88

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 46277000 gb AAS86696.1	5	32	5	10	213
gi 46277026 gb AAS86709.1	5	11	5	7	96
gi 51105058 gb AAT97089.1	5	49	6	18	482
gi 524864548 ref XP_005089109.1	5	28	5	5	91
gi 524865306 ref XP_005089483.1	5	22	5	8	128
gi 524865670 ref XP_005089662.1	5	71	5	97	1767
gi 524866300 ref XP_005089972.1	5	24	5	16	301
gi 524866733 ref XP_005090185.1	5	40	5	10	170
gi 524866733 ref XP_005090185.1	5	33	5	7	103
gi 524867753 ref XP_005090684.1	5	20	12	16	319
gi 524868037 ref XP_005090825.1	5	23	5	10	97
gi 524868530 ref XP_005091067.1	5	16	5	6	135
gi 524868604 ref XP_005091103.1	5	28	6	8	88
gi 524868651 ref XP_005091126.1	5	18	5	5	101
gi 524868983 ref XP_005091289.1	5	51	5	15	136
gi 524869485 ref XP_005091535.1	5	11	5	5	89
gi 524870008 ref XP_005091790.1	5	30	5	9	100
gi 524870146 ref XP_005091859.1	5	29	6	7	115
gi 524870945 ref XP_005092251.1	5	41	5	13	125
gi 524870955 ref XP_005092256.1	5	20	5	9	247
gi 524871172 ref XP_005092362.1	5	13	5	7	61
gi 524871463 ref XP_005092504.1	5	37	5	12	133
gi 524871621 ref XP_005092582.1	5	21	5	12	165
gi 524872186 ref XP_005092857.1	5	20	5	5	60
gi 524872436 ref XP_005092980.1	5	13	6	6	129
gi 524872618 ref XP_005093070.1	5	16	10	14	204
gi 524872622 ref XP_005093072.1	5	12	6	6	77
gi 524872725 ref XP_005093120.1	5	16	5	10	282
gi 524873045 ref XP_005093276.1	5	21	5	19	307
gi 524873315 ref XP_005093407.1	5	26	5	5	105
gi 524873378 ref XP_005093436.1	5	20	5	12	249
gi 524873738 ref XP_005093612.1	5	35	6	6	74
gi 524873892 ref XP_005093688.1	5	28	5	10	266
gi 524874171 ref XP_005093825.1	5	31	5	10	204
gi 524874182 ref XP_005093830.1	5	49	5	10	227
gi 524874452 ref XP_005093961.1	5	27	5	15	138
gi 524876470 ref XP_005094951.1	5	27	5	10	276
gi 524876525 ref XP_005094978.1	5	57	6	23	400
gi 524876560 ref XP_005094995.1	5	10	5	8	201
gi 524877129 ref XP_005095271.1	5	32	5	20	368
gi 524877853 ref XP_005095623.1	5	29	9	20	410
gi 524878217 ref XP_005095797.1	5	23	5	5	96

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524878910 ref XP_005096136.1	5	21	5	8	214
gi 524879191 ref XP_005096273.1	5	15	5	5	91
gi 524879345 ref XP_005096349.1	5	20	5	7	171
gi 524880082 ref XP_005096709.1	5	28	5	10	114
gi 524881469 ref XP_005097386.1	5	18	5	8	173
gi 524881557 ref XP_005097429.1	5	56	11	57	1482
gi 524882112 ref XP_005097701.1	5	35	5	11	102
gi 524882460 ref XP_005097873.1	5	47	5	26	437
gi 524882484 ref XP_005097885.1	5	22	6	11	238
gi 524883119 ref XP_005098193.1	5	10	5	5	53
gi 524883558 ref XP_005098406.1	5	12	5	9	229
gi 524884059 ref XP_005098653.1	5	19	5	14	465
gi 524884444 ref XP_005098842.1	5	23	5	13	405
gi 524884503 ref XP_005098871.1	5	32	5	16	192
gi 524884697 ref XP_005098967.1	5	15	5	5	99
gi 524885145 ref XP_005099183.1	5	14	5	6	170
gi 524885272 ref XP_005099245.1	5	55	5	31	1067
gi 524886570 ref XP_005099878.1	5	49	5	20	683
gi 524887214 ref XP_005100195.1	5	67	6	13	224
gi 524887250 ref XP_005100213.1	5	20	5	7	163
gi 524887580 ref XP_005100374.1	5	13	6	12	243
gi 524887834 ref XP_005100500.1	5	39	5	15	197
gi 524887886 ref XP_005100525.1	5	23	5	14	305
gi 524887980 ref XP_005100569.1	5	16	5	6	145
gi 524888015 ref XP_005100586.1	5	12	5	5	96
gi 524888025 ref XP_005100590.1	5	52	5	8	210
gi 524888089 ref XP_005100621.1	5	21	5	7	167
gi 524888414 ref XP_005100778.1	5	29	5	11	190
gi 524888480 ref XP_005100811.1	5	24	7	8	118
gi 524888807 ref XP_005100968.1	5	10	5	13	333
gi 524889199 ref XP_005101160.1	5	15	5	9	170
gi 524889332 ref XP_005101225.1	5	5	5	5	77
gi 524889992 ref XP_005101546.1	5	53	5	9	80
gi 524889992 ref XP_005101546.1	5	40	8	14	282
gi 524890204 ref XP_005101650.1	5	42	12	22	434
gi 524890370 ref XP_005101732.1	5	10	5	8	122
gi 524890590 ref XP_005101839.1	5	22	5	5	107
gi 524891178 ref XP_005102125.1	5	26	6	10	168
gi 524891741 ref XP_005102403.1	5	17	5	7	117
gi 524891896 ref XP_005102480.1	5	58	8	31	499
gi 524893405 ref XP_005103218.1	5	19	5	5	106
gi 524893771 ref XP_005103394.1	5	26	5	9	270

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524894650 ref XP_005103820.1	5	18	5	6	176
gi 524894931 ref XP_005103955.1	5	26	5	7	66
gi 524894976 ref XP_005103977.1	5	52	5	9	97
gi 524896213 ref XP_005104584.1	5	4	5	6	208
gi 524896814 ref XP_005104878.1	5	30	5	8	196
gi 524896949 ref XP_005104944.1	5	33	5	5	58
gi 524897983 ref XP_005105446.1	5	26	6	14	254
gi 524898757 ref XP_005105824.1	5	14	5	5	102
gi 524899002 ref XP_005105942.1	5	32	11	20	496
gi 524899023 ref XP_005105952.1	5	26	5	9	182
gi 524900380 ref XP_005106610.1	5	31	5	24	475
gi 524900516 ref XP_005106678.1	5	32	5	9	172
gi 524900809 ref XP_005106822.1	5	16	5	7	122
gi 524900993 ref XP_005106911.1	5	20	5	7	202
gi 524902421 ref XP_005107462.1	5	39	5	56	1035
gi 524902626 ref XP_005107542.1	5	55	10	25	508
gi 524902826 ref XP_005107636.1	5	22	5	15	308
gi 524902881 ref XP_005107662.1	5	33	5	5	85
gi 524903041 ref XP_005107738.1	5	39	15	33	637
gi 524903344 ref XP_005107874.1	5	21	5	11	169
gi 524903676 ref XP_005108033.1	5	19	5	7	138
gi 524906739 ref XP_005108592.1	5	26	6	9	183
gi 524906745 ref XP_005108595.1	5	38	8	12	200
gi 524909120 ref XP_005109612.1	5	20	5	6	129
gi 524910662 ref XP_005110206.1	5	37	5	6	76
gi 524910816 ref XP_005110282.1	5	37	18	32	610
gi 524911952 ref XP_005110833.1	5	54	9	51	818
gi 524913016 ref XP_005111342.1	5	25	5	7	135
gi 524913423 ref XP_005111542.1	5	52	5	24	960
gi 524913881 ref XP_005111763.1	5	10	5	9	94
gi 524914082 ref XP_005111860.1	5	36	6	7	107
gi 524914293 ref XP_005111963.1	5	27	5	14	161
gi 524914515 ref XP_005112072.1	5	26	5	6	56
gi 524914607 ref XP_005112116.1	5	23	5	13	134
gi 524914870 ref XP_005112244.1	5	31	5	5	95
gi 524915427 ref XP_005112513.1	5	11	5	6	190
gi 524915677 ref XP_005112631.1	5	40	5	22	447
gi 524915787 ref XP_005112684.1	5	33	5	12	183
gi 524915990 ref XP_005112779.1	5	21	5	7	135
gi 524916301 ref XP_005112930.1	5	18	5	9	131
gi 524916570 ref XP_005113059.1	5	67	9	39	606
gi 524917273 ref XP_005113396.1	5	22	5	16	288

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 525007750 ref XP_005051527.1	5	26	8	19	359
gi 526117361 ref NP_001267749.1	5	59	5	176	3178
gi 526117405 ref NP_001267760.1	5	65	10	38	188
gi 54020890 ref NP_001005689.1	5	19	5	7	89
gi 546686668 gb ERL95792.1	5	39	5	8	212
gi 555706265 gb ESO09498.1	5	29	5	9	199
gi 556095216 gb ESO83869.1	5	29	5	14	330
gi 556095696 gb ESO84349.1	5	19	5	6	77
gi 556097689 gb ESO86341.1	5	48	5	12	262
gi 556098443 gb ESO87095.1	5	20	5	10	167
gi 556099804 gb ESO88456.1	5	13	5	8	172
gi 556101047 gb ESO89699.1	5	8	8	8	186
gi 556101476 gb ESO90128.1	5	13	8	15	258
gi 556103094 gb ESO91746.1	5	36	5	7	164
gi 556104081 gb ESO92733.1	5	10	5	7	83
gi 556105440 gb ESO94092.1	5	11	5	9	169
gi 556106038 gb ESO94690.1	5	17	5	6	146
gi 556106851 gb ESO95503.1	5	7	5	6	180
gi 556108834 gb ESO97486.1	5	16	5	14	239
gi 556108903 gb ESO97555.1	5	26	5	9	191
gi 556116375 gb ESP05027.1	5	22	7	11	202
gi 576693750 gb EUB57355.1	5	40	5	11	154
gi 632946707 ref XP_007888691.1	5	19	5	8	192
gi 632953883 ref XP_007892665.1	5	11	5	7	282
gi 639127132 gb AIA24462.1	5	39	5	6	164
gi 655453820 ref WP_028837045.1	5	18	5	5	85
gi 66361226 pdb 1YX5 B	5	43	5	22	233
gi 71726729 gb AAZ39528.1	5	76	57	349	6335
gi 773571 emb CAA60122.1	5	47	5	8	203
gi 85362949 gb ABC69854.1	5	45	5	36	931
gi 93209532 gb ABF00124.1	5	48	5	125	3291
gi 109157294 pdb 2CE6 A	4	33	4	40	789
gi 115111533 gb ABI84147.1	4	13	4	5	121
gi 133874728 gb ABO40828.1	4	23	4	4	99
gi 156144968 gb ABU53030.1	4	22	4	5	60
gi 157690722 tpe CAL69088.1	4	41	4	24	311
gi 210076789 gb ACJ06748.1	4	40	4	7	86
gi 210076789 gb ACJ06748.1	4	32	4	9	220
gi 218683625 gb ACL00841.1	4	26	4	17	459
gi 220979902 emb CAQ64775.1	4	27	5	7	94
gi 260821662 ref XP_002606151.1	4	55	19	78	1609
gi 307192533 gb EFN75721.1	4	27	4	12	152

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 325296727 ref NP_001191598.1	4	25	4	34	527
gi 325296743 ref NP_001191607.1	4	26	11	30	621
gi 325296839 ref NP_001191661.1	4	39	4	19	152
gi 325296855 ref NP_001191459.1	4	30	6	6	137
gi 325296855 ref NP_001191459.1	4	45	8	10	287
gi 325296937 ref NP_001191493.1	4	29	4	7	66
gi 325297037 ref NP_001191522.1	4	28	4	21	637
gi 325504381 emb CBX41690.1	4	24	4	68	1934
gi 333449383 gb AEF33378.1	4	35	4	14	398
gi 38043955 emb CAD83837.1	4	50	4	39	793
gi 397511018 ref XP_003825879.1	4	37	12	73	1732
gi 405950185 gb EKC18187.1	4	2	4	4	112
gi 405952673 gb EKC20457.1	4	38	6	15	175
gi 405962076 gb EKC27784.1	4	10	4	4	55
gi 405966102 gb EKC31422.1	4	13	4	4	62
gi 405973516 gb EKC38224.1	4	42	10	13	216
gi 410591636 sp IOCME8.1 HYAL_CONAQ	4	13	4	5	70
gi 443682473 gb ELT87054.1	4	22	4	4	34
gi 443686917 gb ELT90035.1	4	13	4	4	65
gi 443689529 gb ELT91902.1	4	35	4	4	39
gi 443705209 gb ELU01864.1	4	34	4	10	208
gi 443720976 gb ELU10481.1	4	20	4	4	64
gi 46395316 dbj BAD16595.1	4	19	4	4	37
gi 472824679 dbj BAN09032.1	4	67	6	76	1264
gi 524864996 ref XP_005089330.1	4	19	4	6	63
gi 524866780 ref XP_005090207.1	4	15	4	4	129
gi 524866838 ref XP_005090235.1	4	31	4	8	330
gi 524866956 ref XP_005090293.1	4	14	6	10	221
gi 524867567 ref XP_005090594.1	4	19	4	4	48
gi 524867786 ref XP_005090700.1	4	9	4	8	256
gi 524867792 ref XP_005090703.1	4	9	4	4	95
gi 524868297 ref XP_005090952.1	4	49	4	5	127
gi 524868395 ref XP_005091000.1	4	11	4	4	59
gi 524868717 ref XP_005091158.1	4	5	4	4	96
gi 524868787 ref XP_005091193.1	4	14	4	7	143
gi 524868815 ref XP_005091207.1	4	25	4	9	152
gi 524869485 ref XP_005091535.1	4	33	4	10	261
gi 524869549 ref XP_005091567.1	4	23	29	51	1098
gi 524870028 ref XP_005091800.1	4	14	4	10	185
gi 524871130 ref XP_005092341.1	4	8	4	4	108
gi 524871372 ref XP_005092459.1	4	21	4	8	151
gi 524871596 ref XP_005092570.1	4	10	4	16	317

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524871621 ref XP_005092582.1	4	43	4	6	95
gi 524871621 ref XP_005092582.1	4	26	5	12	185
gi 524871930 ref XP_005092735.1	4	24	4	10	202
gi 524872284 ref XP_005092905.1	4	22	4	9	192
gi 524872340 ref XP_005092932.1	4	24	4	4	81
gi 524873738 ref XP_005093612.1	4	35	5	6	47
gi 524873824 ref XP_005093654.1	4	19	5	7	164
gi 524873848 ref XP_005093666.1	4	11	4	4	76
gi 524873900 ref XP_005093692.1	4	18	4	5	58
gi 524874148 ref XP_005093814.1	4	22	4	9	111
gi 524874374 ref XP_005093923.1	4	49	12	31	353
gi 524874572 ref XP_005094021.1	4	51	4	9	214
gi 524874629 ref XP_005094049.1	4	11	4	4	63
gi 524874940 ref XP_005094202.1	4	32	4	7	95
gi 524875329 ref XP_005094395.1	4	45	4	42	770
gi 524875359 ref XP_005094409.1	4	51	4	16	191
gi 524875599 ref XP_005094525.1	4	18	4	10	309
gi 524876286 ref XP_005094861.1	4	26	4	8	106
gi 524876525 ref XP_005094978.1	4	30	8	10	131
gi 524877261 ref XP_005095336.1	4	15	6	13	203
gi 524877602 ref XP_005095500.1	4	16	7	9	120
gi 524877849 ref XP_005095621.1	4	16	4	4	61
gi 524878382 ref XP_005095878.1	4	31	4	5	93
gi 524878669 ref XP_005096017.1	4	20	4	4	61
gi 524879077 ref XP_005096218.1	4	19	4	4	72
gi 524879183 ref XP_005096269.1	4	37	4	6	202
gi 524879769 ref XP_005096556.1	4	38	4	8	164
gi 524880507 ref XP_005096916.1	4	34	4	8	112
gi 524880608 ref XP_005096966.1	4	11	5	5	186
gi 524881385 ref XP_005097344.1	4	30	4	5	72
gi 524881557 ref XP_005097429.1	4	61	11	37	831
gi 524881799 ref XP_005097549.1	4	25	4	6	62
gi 524881948 ref XP_005097620.1	4	6	4	8	135
gi 524882007 ref XP_005097649.1	4	15	4	4	82
gi 524882402 ref XP_005097845.1	4	8	4	4	92
gi 524883562 ref XP_005098408.1	4	15	4	5	144
gi 524883640 ref XP_005098447.1	4	18	4	11	167
gi 524884087 ref XP_005098667.1	4	8	4	6	83
gi 524884624 ref XP_005098931.1	4	36	5	15	367
gi 524884995 ref XP_005099109.1	4	9	4	4	76
gi 524885217 ref XP_005099218.1	4	20	5	7	118
gi 524885248 ref XP_005099233.1	4	24	4	6	86

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524885593 ref XP_005099402.1	4	35	6	35	1019
gi 524885844 ref XP_005099525.1	4	10	4	5	70
gi 524886483 ref XP_005099835.1	4	31	4	6	102
gi 524886483 ref XP_005099835.1	4	30	4	4	36
gi 524886662 ref XP_005099924.1	4	14	4	6	95
gi 524887256 ref XP_005100216.1	4	32	4	4	119
gi 524887260 ref XP_005100218.1	4	15	4	7	109
gi 524887964 ref XP_005100561.1	4	9	4	8	137
gi 524888001 ref XP_005100579.1	4	29	4	10	141
gi 524888480 ref XP_005100811.1	4	19	6	10	243
gi 524888600 ref XP_005100869.1	4	19	4	5	67
gi 524888649 ref XP_005100893.1	4	52	4	6	70
gi 524889085 ref XP_005101104.1	4	28	4	7	141
gi 524889251 ref XP_005101186.1	4	24	4	11	143
gi 524889708 ref XP_005101408.1	4	13	4	8	173
gi 524889734 ref XP_005101421.1	4	12	4	4	57
gi 524889859 ref XP_005101481.1	4	16	4	5	76
gi 524890376 ref XP_005101735.1	4	27	4	11	467
gi 524890575 ref XP_005101832.1	4	13	4	5	53
gi 524891530 ref XP_005102298.1	4	8	4	4	51
gi 524891938 ref XP_005102501.1	4	28	4	5	72
gi 524892590 ref XP_005102822.1	4	19	6	10	160
gi 524892768 ref XP_005102909.1	4	24	4	5	100
gi 524892872 ref XP_005102960.1	4	18	4	9	154
gi 524893572 ref XP_005103295.1	4	10	4	9	147
gi 524893765 ref XP_005103391.1	4	11	4	5	146
gi 524894442 ref XP_005103718.1	4	34	4	6	148
gi 524894915 ref XP_005103947.1	4	18	4	5	104
gi 524895123 ref XP_005104049.1	4	38	9	12	186
gi 524896689 ref XP_005104816.1	4	21	4	4	38
gi 524896814 ref XP_005104878.1	4	15	5	9	171
gi 524897909 ref XP_005105409.1	4	19	4	5	249
gi 524898670 ref XP_005105781.1	4	11	4	5	77
gi 524898725 ref XP_005105808.1	4	23	4	4	56
gi 524898902 ref XP_005105894.1	4	43	4	9	81
gi 524899369 ref XP_005106117.1	4	14	4	5	208
gi 524899735 ref XP_005106295.1	4	15	4	6	133
gi 524901357 ref XP_005107086.1	4	19	4	11	229
gi 524901460 ref XP_005107137.1	4	23	4	8	139
gi 524902240 ref XP_005107439.1	4	11	4	11	270
gi 524902423 ref XP_005107463.1	4	13	4	8	92
gi 524902569 ref XP_005107527.1	4	14	4	7	75

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524902628 ref XP_005107543.1	4	48	9	27	555
gi 524902664 ref XP_005107561.1	4	12	4	4	64
gi 524902931 ref XP_005107685.1	4	64	4	11	261
gi 524903184 ref XP_005107804.1	4	8	4	6	75
gi 524903434 ref XP_005107917.1	4	70	4	5	82
gi 524903799 ref XP_005108092.1	4	9	4	5	111
gi 524905758 ref XP_005108242.1	4	22	4	7	106
gi 524905999 ref XP_005108354.1	4	15	4	7	129
gi 524906293 ref XP_005108469.1	4	25	4	4	68
gi 524906745 ref XP_005108595.1	4	19	4	4	50
gi 524907181 ref XP_005108715.1	4	10	4	7	255
gi 524907228 ref XP_005108737.1	4	16	4	4	102
gi 524907238 ref XP_005108742.1	4	12	4	6	118
gi 524907406 ref XP_005108821.1	4	21	4	6	185
gi 524907580 ref XP_005108906.1	4	58	5	13	233
gi 524907664 ref XP_005108948.1	4	7	4	4	74
gi 524907822 ref XP_005109023.1	4	14	4	6	107
gi 524907998 ref XP_005109108.1	4	17	4	5	129
gi 524908916 ref XP_005109547.1	4	11	4	6	87
gi 524909155 ref XP_005109628.1	4	23	4	7	108
gi 524909236 ref XP_005109667.1	4	29	5	11	236
gi 524909272 ref XP_005109685.1	4	52	4	4	73
gi 524909283 ref XP_005109690.1	4	20	4	8	215
gi 524909958 ref XP_005109871.1	4	16	4	4	88
gi 524910389 ref XP_005110074.1	4	18	4	6	192
gi 524910744 ref XP_005110246.1	4	20	4	4	120
gi 524911947 ref XP_005110831.1	4	57	18	373	6727
gi 524911950 ref XP_005110832.1	4	53	18	214	4236
gi 524913026 ref XP_005111347.1	4	14	5	8	194
gi 524913132 ref XP_005111399.1	4	29	4	7	83
gi 524913755 ref XP_005111702.1	4	31	4	4	125
gi 524913929 ref XP_005111786.1	4	6	4	6	86
gi 524914499 ref XP_005112064.1	4	8	4	4	58
gi 524914928 ref XP_005112273.1	4	16	4	9	326
gi 524915405 ref XP_005112503.1	4	37	4	20	443
gi 529444582 ref XP_005242003.1	4	25	4	6	94
gi 544604255 sp Q3YLA4.2 NCS1_LYMST	4	27	4	6	183
gi 554529929 ref XP_005859764.1	4	47	4	7	98
gi 555704960 gb ESO08193.1	4	46	4	24	469
gi 556094506 gb ESO83159.1	4	21	4	5	74
gi 556094522 gb ESO83175.1	4	9	4	5	100
gi 556097226 gb ESO85878.1	4	12	4	4	64

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 556097653 gb ESO86305.1	4	11	4	5	78
gi 556098530 gb ESO87182.1	4	23	4	4	81
gi 556101476 gb ESO90128.1	4	10	5	6	159
gi 556102412 gb ESO91064.1	4	13	4	15	583
gi 556105643 gb ESO94295.1	4	32	4	6	121
gi 556105791 gb ESO94443.1	4	6	4	6	103
gi 556106190 gb ESO94842.1	4	19	6	16	286
gi 556106190 gb ESO94842.1	4	32	9	21	489
gi 556110210 gb ESO98862.1	4	22	4	4	61
gi 556110324 gb ESO98976.1	4	9	4	4	43
gi 556111239 gb ESO99890.1	4	13	4	7	206
gi 556113679 gb ESP02331.1	4	27	4	6	113
gi 556114019 gb ESP02671.1	4	17	4	4	43
gi 556114028 gb ESP02680.1	4	33	4	4	95
gi 556114329 gb ESP02981.1	4	28	4	8	109
gi 556116442 gb ESP05094.1	4	48	4	11	402
gi 568939963 ref XP_006505278.1	4	42	4	10	259
gi 589107123 ref XP_006965584.1	4	42	4	9	288
gi 619323847 gb AHY00642.1	4	10	4	4	65
gi 646720608 gb KDR22269.1	4	45	4	9	212
gi 68272049 gb AAY89300.1	4	66	10	53	856
gi 71040619 dbj BAE16013.1	4	27	4	16	573
gi 100913266 gb ABF69533.1	3	15	3	6	47
gi 1169742 sp P42577.2 FRIS_LYMST	3	57	3	5	90
gi 126697398 gb ABO26656.1	3	19	3	6	128
gi 148236249 ref NP_001087612.1	3	31	4	4	52
gi 152003983 gb ABS19815.1	3	16	12	19	357
gi 156392725 ref XP_001636198.1	3	10	3	3	44
gi 170053486 ref XP_001862696.1	3	34	3	4	45
gi 184186107 ref NP_001116975.1	3	22	4	6	117
gi 195429214 ref XP_002062659.1	3	26	3	7	96
gi 22024001 ref NP_523708.2	3	24	3	5	110
gi 2493427 sp Q27312.1 NADA_APLKU	3	11	3	4	78
gi 253317417 gb ACT22633.1	3	49	9	208	4145
gi 260784400 ref XP_002587255.1	3	8	3	3	64
gi 260813969 ref XP_002601688.1	3	7	3	3	68
gi 260828603 ref XP_002609252.1	3	63	10	53	1492
gi 281345481 gb EFB21065.1	3	69	4	9	150
gi 29378327 gb AAO83845.1 AF484088_1	3	19	3	4	131
gi 294896502 ref XP_002775589.1	3	43	11	20	344
gi 297186099 gb ADI24337.1	3	9	3	9	187
gi 325197162 ref NP_001191425.1	3	18	3	5	71

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 325209004 ref NP_001191451.1	3	17	3	4	97
gi 325296731 ref NP_001191601.1	3	11	3	4	103
gi 325296853 ref NP_001191458.1	3	26	4	6	134
gi 325297011 ref NP_001191514.1	3	16	4	9	144
gi 38043955 emb CAD83837.1	3	32	4	32	910
gi 405950236 gb EKC18235.1	3	11	3	3	33
gi 405951631 gb EKC19528.1	3	20	3	7	276
gi 405958669 gb EKC24774.1	3	12	4	7	190
gi 405965644 gb EKC31006.1	3	21	6	7	170
gi 405967885 gb EKC33004.1	3	14	3	5	87
gi 405975533 gb EKC40092.1	3	14	3	5	65
gi 405976805 gb EKC41289.1	3	6	3	4	33
gi 405977577 gb EKC42020.1	3	13	3	4	77
gi 405978690 gb EKC43060.1	3	7	3	3	118
gi 42560362 gb AAS20336.1	3	50	7	438	8101
gi 443694956 gb ELT95974.1	3	30	3	5	85
gi 443706726 gb ELU02640.1	3	10	3	4	62
gi 443729373 gb ELU15297.1	3	19	4	13	96
gi 470643071 ref XP_004326123.1	3	82	13	87	2069
gi 47202433 emb CAF87387.1	3	48	3	3	32
gi 50593062 gb AAT79342.1	3	45	3	14	481
gi 524863982 ref XP_005088830.1	3	38	3	13	210
gi 524864475 ref XP_005089073.1	3	16	3	3	56
gi 524864482 ref XP_005089076.1	3	10	3	6	130
gi 524864620 ref XP_005089145.1	3	21	3	7	163
gi 524864626 ref XP_005089148.1	3	12	3	5	124
gi 524864665 ref XP_005089167.1	3	18	3	16	181
gi 524864840 ref XP_005089254.1	3	12	3	3	80
gi 524865955 ref XP_005089803.1	3	28	4	11	124
gi 524866007 ref XP_005089828.1	3	4	3	4	142
gi 524866201 ref XP_005089924.1	3	5	3	3	47
gi 524866665 ref XP_005090151.1	3	31	3	8	171
gi 524866836 ref XP_005090234.1	3	27	3	7	270
gi 524866978 ref XP_005090304.1	3	42	3	10	267
gi 524867383 ref XP_005090503.1	3	9	3	3	50
gi 524867640 ref XP_005090628.1	3	26	3	6	155
gi 524868832 ref XP_005091215.1	3	15	3	4	109
gi 524868942 ref XP_005091269.1	3	42	3	3	66
gi 524868994 ref XP_005091294.1	3	12	3	6	252
gi 524869145 ref XP_005091369.1	3	41	3	9	211
gi 524869207 ref XP_005091400.1	3	25	4	6	132
gi 524869284 ref XP_005091437.1	3	17	3	11	201

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524869323 ref XP_005091456.1	3	9	3	3	70
gi 524869393 ref XP_005091491.1	3	28	3	8	187
gi 524869633 ref XP_005091608.1	3	11	3	4	77
gi 524870217 ref XP_005091894.1	3	19	3	5	111
gi 524870317 ref XP_005091943.1	3	7	3	5	90
gi 524870720 ref XP_005092142.1	3	14	3	5	248
gi 524870819 ref XP_005092190.1	3	6	3	3	53
gi 524870833 ref XP_005092197.1	3	12	3	4	134
gi 524870909 ref XP_005092234.1	3	11	3	3	55
gi 524871469 ref XP_005092507.1	3	7	3	3	41
gi 524871930 ref XP_005092735.1	3	25	3	5	85
gi 524872186 ref XP_005092857.1	3	10	3	3	137
gi 524872446 ref XP_005092985.1	3	16	3	4	77
gi 524872672 ref XP_005093096.1	3	55	3	5	99
gi 524872727 ref XP_005093121.1	3	28	3	3	58
gi 524872775 ref XP_005093144.1	3	6	3	3	62
gi 524872805 ref XP_005093159.1	3	18	3	6	140
gi 524872934 ref XP_005093222.1	3	19	3	3	53
gi 524873075 ref XP_005093291.1	3	21	3	10	184
gi 524873194 ref XP_005093348.1	3	6	3	3	67
gi 524873970 ref XP_005093727.1	3	43	3	5	91
gi 524873980 ref XP_005093732.1	3	9	3	3	58
gi 524874037 ref XP_005093760.1	3	31	3	6	108
gi 524874771 ref XP_005094120.1	3	15	3	3	101
gi 524874903 ref XP_005094185.1	3	4	3	3	66
gi 524875325 ref XP_005094393.1	3	26	3	8	115
gi 524875560 ref XP_005094506.1	3	14	3	8	169
gi 524876140 ref XP_005094789.1	3	14	3	3	62
gi 524876525 ref XP_005094978.1	3	16	3	3	124
gi 524876525 ref XP_005094978.1	3	35	10	19	374
gi 524876601 ref XP_005095013.1	3	14	3	3	72
gi 524876636 ref XP_005095029.1	3	12	3	4	50
gi 524876864 ref XP_005095141.1	3	13	3	11	207
gi 524877098 ref XP_005095256.1	3	32	5	21	197
gi 524877582 ref XP_005095490.1	3	10	3	4	78
gi 524877853 ref XP_005095623.1	3	41	6	17	283
gi 524878217 ref XP_005095797.1	3	14	3	6	114
gi 524878504 ref XP_005095936.1	3	85	6	6	153
gi 524878598 ref XP_005095982.1	3	17	3	5	94
gi 524878653 ref XP_005096009.1	3	37	3	11	188
gi 524878707 ref XP_005096036.1	3	9	4	4	47
gi 524878983 ref XP_005096171.1	3	32	8	10	96

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524879487 ref XP_005096418.1	3	21	3	4	53
gi 524879771 ref XP_005096557.1	3	30	10	11	253
gi 524879943 ref XP_005096641.1	3	24	3	3	52
gi 524879957 ref XP_005096647.1	3	19	3	6	259
gi 524880074 ref XP_005096705.1	3	22	3	3	35
gi 524880570 ref XP_005096947.1	3	30	3	4	62
gi 524880608 ref XP_005096966.1	3	13	4	7	232
gi 524880769 ref XP_005097043.1	3	15	3	5	112
gi 524880783 ref XP_005097050.1	3	20	3	3	47
gi 524880992 ref XP_005097152.1	3	8	3	3	44
gi 524881473 ref XP_005097388.1	3	32	4	7	153
gi 524881555 ref XP_005097428.1	3	25	5	16	557
gi 524881765 ref XP_005097532.1	3	17	3	4	75
gi 524881775 ref XP_005097537.1	3	7	3	3	74
gi 524882193 ref XP_005097741.1	3	28	3	8	180
gi 524882422 ref XP_005097855.1	3	72	6	12	146
gi 524882784 ref XP_005098028.1	3	12	3	3	109
gi 524882922 ref XP_005098097.1	3	12	3	3	115
gi 524883088 ref XP_005098178.1	3	37	9	15	204
gi 524883388 ref XP_005098325.1	3	20	3	12	214
gi 524883534 ref XP_005098394.1	3	37	3	5	56
gi 524883622 ref XP_005098438.1	3	6	3	3	80
gi 524883711 ref XP_005098482.1	3	17	3	5	156
gi 524883861 ref XP_005098556.1	3	34	5	8	88
gi 524883986 ref XP_005098618.1	3	16	3	3	61
gi 524884097 ref XP_005098672.1	3	9	3	4	155
gi 524884410 ref XP_005098825.1	3	15	3	5	97
gi 524884624 ref XP_005098931.1	3	41	4	11	249
gi 524884820 ref XP_005099026.1	3	24	3	10	200
gi 524884828 ref XP_005099030.1	3	9	3	5	119
gi 524885349 ref XP_005099283.1	3	34	5	19	248
gi 524885795 ref XP_005099501.1	3	68	34	165	4075
gi 524886285 ref XP_005099739.1	3	34	3	6	92
gi 524886568 ref XP_005099877.1	3	41	3	8	167
gi 524887166 ref XP_005100172.1	3	27	4	7	81
gi 524887212 ref XP_005100194.1	3	16	3	3	32
gi 524887216 ref XP_005100196.1	3	22	3	3	149
gi 524887218 ref XP_005100197.1	3	28	8	10	251
gi 524887218 ref XP_005100197.1	3	18	9	11	304
gi 524887498 ref XP_005100334.1	3	7	3	4	145
gi 524888025 ref XP_005100590.1	3	28	3	5	79
gi 524888089 ref XP_005100621.1	3	15	3	4	112

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524888237 ref XP_005100694.1	3	23	3	7	214
gi 524888508 ref XP_005100824.1	3	62	3	19	271
gi 524888744 ref XP_005100937.1	3	13	3	3	88
gi 524889396 ref XP_005101256.1	3	10	3	3	39
gi 524889459 ref XP_005101285.1	3	9	3	3	86
gi 524889607 ref XP_005101358.1	3	13	3	5	66
gi 524889661 ref XP_005101385.1	3	10	3	8	117
gi 524889764 ref XP_005101435.1	3	33	3	10	124
gi 524890281 ref XP_005101688.1	3	18	3	4	107
gi 524890561 ref XP_005101826.1	3	7	3	4	68
gi 524890899 ref XP_005101991.1	3	11	3	3	98
gi 524891407 ref XP_005102237.1	3	10	3	4	63
gi 524891741 ref XP_005102403.1	3	14	3	7	121
gi 524891741 ref XP_005102403.1	3	11	3	3	36
gi 524892029 ref XP_005102546.1	3	6	3	3	90
gi 524892053 ref XP_005102558.1	3	29	3	3	78
gi 524892143 ref XP_005102602.1	3	7	3	5	101
gi 524892201 ref XP_005102630.1	3	49	4	4	57
gi 524892590 ref XP_005102822.1	3	14	5	10	201
gi 524893085 ref XP_005103063.1	3	15	3	4	97
gi 524893098 ref XP_005103069.1	3	22	3	20	284
gi 524893513 ref XP_005103269.1	3	5	3	3	73
gi 524893558 ref XP_005103288.1	3	10	3	3	73
gi 524893741 ref XP_005103379.1	3	30	4	9	112
gi 524894028 ref XP_005103521.1	3	10	3	4	100
gi 524894498 ref XP_005103746.1	3	13	3	3	42
gi 524894785 ref XP_005103884.1	3	4	3	3	64
gi 524894862 ref XP_005103922.1	3	13	5	7	106
gi 524895398 ref XP_005104182.1	3	9	3	3	60
gi 524895637 ref XP_005104300.1	3	12	3	8	135
gi 524895763 ref XP_005104363.1	3	16	3	3	58
gi 524896189 ref XP_005104572.1	3	13	4	4	82
gi 524896250 ref XP_005104602.1	3	14	3	4	139
gi 524896597 ref XP_005104771.1	3	20	3	4	80
gi 524896989 ref XP_005104963.1	3	18	3	4	64
gi 524897931 ref XP_005105420.1	3	6	3	3	50
gi 524898101 ref XP_005105504.1	3	10	3	3	80
gi 524898135 ref XP_005105521.1	3	2	3	4	94
gi 524898175 ref XP_005105540.1	3	7	3	6	179
gi 524898265 ref XP_005105584.1	3	19	3	4	112
gi 524898272 ref XP_005105587.1	3	24	3	11	177
gi 524898539 ref XP_005105717.1	3	14	3	5	91

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524898674 ref XP_005105783.1	3	15	3	4	72
gi 524898763 ref XP_005105827.1	3	14	3	3	56
gi 524898815 ref XP_005105852.1	3	21	3	8	102
gi 524899012 ref XP_005105947.1	3	13	3	4	92
gi 524899021 ref XP_005105951.1	3	17	3	8	185
gi 524899023 ref XP_005105952.1	3	22	3	40	1669
gi 524899511 ref XP_005106186.1	3	70	10	10	98
gi 524899675 ref XP_005106266.1	3	12	3	3	38
gi 524899675 ref XP_005106266.1	3	17	3	3	57
gi 524899892 ref XP_005106372.1	3	21	3	3	46
gi 524900236 ref XP_005106539.1	3	17	3	4	119
gi 524900270 ref XP_005106556.1	3	32	3	8	144
gi 524900353 ref XP_005106597.1	3	11	3	4	176
gi 524900538 ref XP_005106689.1	3	7	3	4	121
gi 524901394 ref XP_005107104.1	3	23	4	10	366
gi 524901619 ref XP_005107215.1	3	4	3	3	80
gi 524901901 ref XP_005107354.1	3	18	3	6	139
gi 524902714 ref XP_005107585.1	3	14	3	3	31
gi 524903041 ref XP_005107738.1	3	23	4	13	290
gi 524903247 ref XP_005107829.1	3	12	3	4	87
gi 524903608 ref XP_005108000.1	3	16	3	8	132
gi 524905947 ref XP_005108331.1	3	26	3	6	129
gi 524905999 ref XP_005108354.1	3	18	3	7	165
gi 524906745 ref XP_005108595.1	3	31	5	10	163
gi 524907032 ref XP_005108645.1	3	53	4	6	184
gi 524907580 ref XP_005108906.1	3	3	4	5	104
gi 524907590 ref XP_005108911.1	3	18	3	8	302
gi 524907727 ref XP_005108978.1	3	13	3	3	63
gi 524908048 ref XP_005109132.1	3	5	3	4	69
gi 524908261 ref XP_005109234.1	3	13	3	5	144
gi 524908261 ref XP_005109234.1	3	11	3	6	120
gi 524908282 ref XP_005109244.1	3	35	3	14	376
gi 524908297 ref XP_005109251.1	3	11	3	3	61
gi 524908577 ref XP_005109382.1	3	9	3	3	68
gi 524909270 ref XP_005109684.1	3	13	3	4	129
gi 524909457 ref XP_005109775.1	3	8	3	5	156
gi 524910500 ref XP_005110126.1	3	3	3	3	39
gi 524910662 ref XP_005110206.1	3	22	3	3	45
gi 524910831 ref XP_005110289.1	3	33	3	6	77
gi 524910950 ref XP_005110347.1	3	9	3	4	63
gi 524911110 ref XP_005110424.1	3	18	3	5	115
gi 524911477 ref XP_005110601.1	3	53	3	8	311

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524911650 ref XP_005110685.1	3	11	3	5	173
gi 524912055 ref XP_005110881.1	3	7	3	4	112
gi 524912187 ref XP_005110945.1	3	10	3	3	73
gi 524912355 ref XP_005111027.1	3	15	3	5	84
gi 524912927 ref XP_005111299.1	3	43	3	9	198
gi 524913184 ref XP_005111424.1	3	20	3	6	158
gi 524913273 ref XP_005111468.1	3	26	3	9	286
gi 524913662 ref XP_005111656.1	3	14	3	4	86
gi 524913668 ref XP_005111659.1	3	20	3	6	153
gi 524913879 ref XP_005111762.1	3	7	3	5	173
gi 524913883 ref XP_005111764.1	3	25	3	6	58
gi 524914146 ref XP_005111892.1	3	5	3	4	67
gi 524914287 ref XP_005111960.1	3	28	3	4	112
gi 524914353 ref XP_005111992.1	3	13	3	4	85
gi 524914806 ref XP_005112213.1	3	7	3	5	75
gi 524914928 ref XP_005112273.1	3	10	3	4	79
gi 524915372 ref XP_005112487.1	3	17	3	3	60
gi 524915376 ref XP_005112489.1	3	15	4	4	96
gi 524916570 ref XP_005113059.1	3	62	8	23	262
gi 524916601 ref XP_005113074.1	3	22	4	9	100
gi 524916601 ref XP_005113074.1	3	24	6	11	254
gi 524916639 ref XP_005113093.1	3	20	3	7	159
gi 524916644 ref XP_005113095.1	3	13	3	10	164
gi 524916993 ref XP_005113263.1	3	8	3	3	96
gi 526117431 ref NP_001267767.1	3	31	5	86	1230
gi 556094374 gb ESO83027.1	3	13	3	3	74
gi 556097689 gb ESO86341.1	3	17	3	3	46
gi 556097805 gb ESO86457.1	3	3	3	3	65
gi 556098509 gb ESO87161.1	3	7	3	4	156
gi 556098668 gb ESO87320.1	3	13	3	5	171
gi 556098954 gb ESO87606.1	3	2	3	4	101
gi 556101818 gb ESO90470.1	3	9	3	4	57
gi 556102645 gb ESO91297.1	3	7	3	7	146
gi 556103976 gb ESO92628.1	3	19	3	3	29
gi 556105206 gb ESO93858.1	3	20	3	6	120
gi 556106320 gb ESO94972.1	3	19	3	5	85
gi 556106462 gb ESO95114.1	3	5	3	3	146
gi 556107134 gb ESO95786.1	3	10	3	4	76
gi 556107378 gb ESO96030.1	3	17	3	4	93
gi 556108215 gb ESO96867.1	3	8	3	3	74
gi 556108856 gb ESO97508.1	3	8	3	3	56
gi 556108856 gb ESO97508.1	3	8	3	4	54

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 556108929 gb ESO97581.1	3	34	3	3	38
gi 556109606 gb ESO98258.1	3	11	3	8	152
gi 556110532 gb ESO99183.1	3	12	3	6	137
gi 556110866 gb ESO99517.1	3	53	40	177	4066
gi 556113064 gb ESP01716.1	3	12	3	4	127
gi 556113650 gb ESP02302.1	3	37	3	3	109
gi 556114587 gb ESP03239.1	3	8	3	3	30
gi 556116273 gb ESP04925.1	3	24	3	3	72
gi 577029965 gb AHH81817.1	3	40	3	5	61
gi 583994656 ref XP_006792469.1	3	28	3	32	436
gi 602642710 ref XP_007428140.1	3	49	3	6	153
gi 608096069 gb EZG34785.1	3	17	3	13	302
gi 632935059 ref XP_007887607.1	3	14	3	6	124
gi 635065029 ref XP_008001282.1	3	62	4	27	325
gi 6492342 gb AAF14287.1 AF182339_1	3	7	3	3	63
gi 67005832 gb AAN31641.3 AF486290_1	3	10	3	3	86
gi 68533593 gb AAH98579.1	3	6	3	3	88
gi 72084055 ref XP_790644.1	3	11	3	3	92
gi 93209532 gb ABF00124.1	3	27	3	79	1692
gi 109157294 pdb 2CE6 A	2	29	2	2	34
gi 110764763 ref XP_001123191.1	2	44	2	8	239
gi 1174755 sp P43689.1 TPM2_BIOGL	2	58	23	33	515
gi 118401762 ref XP_001033201.1	2	18	2	4	81
gi 126697272 gb ABO26593.1	2	21	3	5	81
gi 126697410 gb ABO26662.1	2	16	2	4	102
gi 149032103 gb EDL87015.1	2	41	9	53	1271
gi 150251392 gb ABR68008.1	2	35	5	9	178
gi 156373963 ref XP_001629579.1	2	8	2	2	27
gi 198285569 gb ACH85323.1	2	7	2	2	49
gi 198430879 ref XP_002123222.1	2	38	2	4	188
gi 2073146 dbj BAA19863.1	2	16	2	2	57
gi 220979902 emb CAQ64775.1	2	44	7	9	186
gi 260807417 ref XP_002598505.1	2	17	2	2	36
gi 260811516 ref XP_002600468.1	2	16	2	3	73
gi 27763677 gb AAO20109.1	2	9	2	2	40
gi 291234885 ref XP_002737379.1	2	29	2	5	100
gi 291242935 ref XP_002741349.1	2	20	2	2	30
gi 291245044 ref XP_002742403.1	2	8	2	3	65
gi 293595779 gb ADE45333.1	2	14	2	3	128
gi 29378329 gb AAO83846.1 AF484089_1	2	15	2	4	80
gi 29378333 gb AAO83848.1 AF484091_1	2	27	4	23	845
gi 29378335 gb AAO83849.1 AF484092_1	2	4	2	2	55

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 297186099 gb ADI24337.1	2	15	2	2	50
gi 323134763 gb ADX31293.1	2	14	2	4	84
gi 325197143 ref NP_001191422.1	2	17	2	4	81
gi 325296789 ref NP_001191633.1	2	5	2	2	48
gi 325296853 ref NP_001191458.1	2	24	5	5	114
gi 325296855 ref NP_001191459.1	2	24	4	5	114
gi 325297015 ref NP_001191515.1	2	15	5	5	101
gi 325297068 ref NP_001191538.1	2	8	2	2	62
gi 328905044 gb AEB54797.1	2	11	2	2	30
gi 340381720 ref XP_003389369.1	2	16	2	11	168
gi 346721859 gb AEO50742.1	2	14	3	7	118
gi 346987840 gb AEO51766.1	2	13	2	8	185
gi 346987844 gb AEO51768.1	2	38	65	120	2074
gi 374718481 gb AEZ67032.1	2	5	2	4	89
gi 395484809 gb AFN66649.1	2	5	2	2	41
gi 397776464 gb AFO64930.1	2	27	3	5	110
gi 403310251 emb CCJ09600.1	2	6	2	3	70
gi 405952171 gb EKC20013.1	2	6	2	3	114
gi 405952830 gb EKC20593.1	2	5	2	2	82
gi 405955268 gb EKC22447.1	2	3	2	2	48
gi 405957390 gb EKC23604.1	2	18	2	3	68
gi 405957680 gb EKC23874.1	2	6	2	2	36
gi 405958669 gb EKC24774.1	2	8	2	3	60
gi 405960075 gb EKC26025.1	2	5	2	5	124
gi 405960381 gb EKC26308.1	2	35	3	8	224
gi 405960381 gb EKC26308.1	2	35	3	8	160
gi 405960599 gb EKC26510.1	2	24	2	2	28
gi 405960672 gb EKC26570.1	2	26	2	6	118
gi 405962100 gb EKC27804.1	2	4	2	2	34
gi 405970416 gb EKC35324.1	2	3	2	3	81
gi 405970417 gb EKC35325.1	2	4	2	2	32
gi 405970417 gb EKC35325.1	2	6	2	2	47
gi 405972817 gb EKC37565.1	2	54	2	6	117
gi 405972881 gb EKC37628.1	2	10	2	5	82
gi 405974177 gb EKC38843.1	2	12	2	2	58
gi 405977265 gb EKC41724.1	2	5	2	2	45
gi 405977876 gb EKC42303.1	2	12	2	2	98
gi 42559558 sp O97192.1 TPM_HELAS	2	24	8	9	130
gi 443720976 gb ELU10481.1	2	9	2	2	50
gi 4504279 ref NP_002098.1	2	15	3	12	95
gi 459180352 ref XP_004226627.1	2	64	18	99	2063
gi 46277000 gb AAS86696.1	2	6	2	2	75

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 46277106 gb AAS86749.1	2	22	2	3	27
gi 4929136 gb AAD33872.1 AF141348_1	2	23	4	5	114
gi 51105030 gb AAT97075.1	2	17	2	5	77
gi 51105058 gb AAT97089.1	2	45	12	19	518
gi 511880705 ref XP_004760254.1	2	27	3	3	33
gi 524864102 ref XP_005088889.1	2	20	2	2	32
gi 524864130 ref XP_005088903.1	2	14	2	6	211
gi 524864189 ref XP_005088932.1	2	13	2	3	77
gi 524864189 ref XP_005088932.1	2	18	2	2	58
gi 524864582 ref XP_005089126.1	2	10	2	3	64
gi 524864630 ref XP_005089150.1	2	32	2	7	111
gi 524864630 ref XP_005089150.1	2	11	2	2	33
gi 524864893 ref XP_005089280.1	2	11	2	3	112
gi 524865136 ref XP_005089400.1	2	11	2	3	67
gi 524865144 ref XP_005089404.1	2	12	2	2	35
gi 524865614 ref XP_005089635.1	2	9	2	2	85
gi 524865813 ref XP_005089733.1	2	23	4	7	109
gi 524865869 ref XP_005089760.1	2	12	2	2	45
gi 524866201 ref XP_005089924.1	2	13	2	3	54
gi 524866264 ref XP_005089955.1	2	8	2	2	39
gi 524866798 ref XP_005090215.1	2	14	2	2	75
gi 524866978 ref XP_005090304.1	2	45	3	8	183
gi 524866999 ref XP_005090313.1	2	8	2	2	51
gi 524867173 ref XP_005090399.1	2	8	2	2	36
gi 524867407 ref XP_005090515.1	2	8	2	2	40
gi 524867567 ref XP_005090594.1	2	10	2	2	75
gi 524867755 ref XP_005090685.1	2	9	6	6	183
gi 524867763 ref XP_005090689.1	2	9	5	5	182
gi 524867882 ref XP_005090748.1	2	27	2	7	159
gi 524868005 ref XP_005090809.1	2	47	3	8	130
gi 524868127 ref XP_005090870.1	2	9	2	2	74
gi 524868280 ref XP_005090944.1	2	4	2	2	55
gi 524868395 ref XP_005091000.1	2	6	2	2	92
gi 524868850 ref XP_005091223.1	2	10	2	3	88
gi 524868888 ref XP_005091242.1	2	52	3	5	152
gi 524868962 ref XP_005091279.1	2	6	2	2	22
gi 524868983 ref XP_005091289.1	2	14	3	3	80
gi 524869296 ref XP_005091443.1	2	11	2	3	87
gi 524869517 ref XP_005091551.1	2	8	2	4	55
gi 524869549 ref XP_005091567.1	2	2	2	2	34
gi 524869798 ref XP_005091688.1	2	8	2	3	77
gi 524870116 ref XP_005091844.1	2	19	2	5	124

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524870146 ref XP_005091859.1	2	28	3	3	67
gi 524870213 ref XP_005091892.1	2	35	2	19	365
gi 524870306 ref XP_005091938.1	2	4	2	2	30
gi 524870519 ref XP_005092044.1	2	5	2	2	42
gi 524870720 ref XP_005092142.1	2	9	2	2	69
gi 524870754 ref XP_005092159.1	2	19	2	2	42
gi 524870758 ref XP_005092161.1	2	8	2	2	56
gi 524871250 ref XP_005092401.1	2	27	2	4	96
gi 524871258 ref XP_005092404.1	2	5	2	3	87
gi 524871394 ref XP_005092470.1	2	14	2	4	87
gi 524871451 ref XP_005092498.1	2	6	2	2	60
gi 524871596 ref XP_005092570.1	2	2	2	3	133
gi 524871599 ref XP_005092571.1	2	7	2	2	66
gi 524871635 ref XP_005092589.1	2	9	2	3	99
gi 524872452 ref XP_005092988.1	2	18	2	4	78
gi 524872578 ref XP_005093050.1	2	4	2	3	107
gi 524872604 ref XP_005093063.1	2	6	2	3	60
gi 524872801 ref XP_005093157.1	2	51	9	20	345
gi 524872910 ref XP_005093210.1	2	11	2	3	116
gi 524873152 ref XP_005093328.1	2	4	2	2	45
gi 524873501 ref XP_005093496.1	2	3	2	2	50
gi 524873892 ref XP_005093688.1	2	37	2	7	311
gi 524873954 ref XP_005093719.1	2	6	2	2	67
gi 524874182 ref XP_005093830.1	2	14	2	2	47
gi 524874186 ref XP_005093832.1	2	10	2	2	38
gi 524874190 ref XP_005093834.1	2	7	2	2	45
gi 524874352 ref XP_005093912.1	2	3	2	2	70
gi 524874629 ref XP_005094049.1	2	12	2	2	59
gi 524874903 ref XP_005094185.1	2	5	2	4	57
gi 524875141 ref XP_005094301.1	2	16	2	3	70
gi 524875173 ref XP_005094317.1	2	6	2	2	51
gi 524875207 ref XP_005094334.1	2	12	2	3	121
gi 524875757 ref XP_005094602.1	2	3	2	2	48
gi 524876116 ref XP_005094777.1	2	15	2	2	49
gi 524876138 ref XP_005094788.1	2	5	2	2	40
gi 524876523 ref XP_005094977.1	2	47	4	4	84
gi 524876525 ref XP_005094978.1	2	32	4	7	187
gi 524876618 ref XP_005095021.1	2	8	2	3	51
gi 524876668 ref XP_005095045.1	2	13	2	3	79
gi 524876816 ref XP_005095117.1	2	6	2	2	74
gi 524876998 ref XP_005095207.1	2	4	2	2	37
gi 524877372 ref XP_005095389.1	2	30	2	3	48

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524877614 ref XP_005095506.1	2	8	4	7	129
gi 524877675 ref XP_005095535.1	2	3	2	2	70
gi 524877793 ref XP_005095594.1	2	14	2	3	50
gi 524877803 ref XP_005095598.1	2	8	2	2	33
gi 524878219 ref XP_005095798.1	2	19	2	3	65
gi 524878295 ref XP_005095836.1	2	3	2	5	169
gi 524878368 ref XP_005095871.1	2	8	2	2	49
gi 524878410 ref XP_005095892.1	2	5	2	2	37
gi 524878707 ref XP_005096036.1	2	18	3	3	72
gi 524878727 ref XP_005096045.1	2	8	2	2	65
gi 524878727 ref XP_005096045.1	2	8	2	4	136
gi 524878917 ref XP_005096139.1	2	11	2	9	222
gi 524879087 ref XP_005096223.1	2	7	2	2	22
gi 524879355 ref XP_005096354.1	2	15	2	2	38
gi 524879751 ref XP_005096547.1	2	4	2	3	77
gi 524879821 ref XP_005096582.1	2	9	2	3	52
gi 524880074 ref XP_005096705.1	2	17	2	11	171
gi 524880525 ref XP_005096925.1	2	6	2	3	45
gi 524880698 ref XP_005097010.1	2	5	2	2	55
gi 524880806 ref XP_005097061.1	2	7	2	5	88
gi 524880862 ref XP_005097089.1	2	12	2	3	50
gi 524880971 ref XP_005097142.1	2	7	2	2	24
gi 524880980 ref XP_005097146.1	2	6	2	3	59
gi 524881358 ref XP_005097331.1	2	24	2	3	46
gi 524881413 ref XP_005097358.1	2	17	3	7	204
gi 524881487 ref XP_005097395.1	2	7	2	2	52
gi 524881587 ref XP_005097444.1	2	4	2	5	69
gi 524881805 ref XP_005097552.1	2	12	2	5	69
gi 524881845 ref XP_005097572.1	2	18	2	2	78
gi 524882014 ref XP_005097652.1	2	7	2	2	41
gi 524882249 ref XP_005097769.1	2	10	2	2	51
gi 524882640 ref XP_005097961.1	2	4	2	2	43
gi 524882665 ref XP_005097972.1	2	13	2	2	40
gi 524882721 ref XP_005097999.1	2	11	2	3	170
gi 524882733 ref XP_005098005.1	2	23	2	4	90
gi 524882750 ref XP_005098013.1	2	5	2	2	46
gi 524882784 ref XP_005098028.1	2	11	2	5	151
gi 524882812 ref XP_005098042.1	2	8	2	2	66
gi 524882956 ref XP_005098113.1	2	7	2	2	73
gi 524882968 ref XP_005098119.1	2	15	2	2	55
gi 524883074 ref XP_005098171.1	2	4	2	2	40
gi 524883090 ref XP_005098179.1	2	28	8	12	155

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524883672 ref XP_005098463.1	2	5	2	3	78
gi 524883770 ref XP_005098511.1	2	4	2	2	95
gi 524883861 ref XP_005098556.1	2	34	5	5	46
gi 524883909 ref XP_005098580.1	2	21	2	5	44
gi 524884059 ref XP_005098653.1	2	6	2	5	168
gi 524884099 ref XP_005098673.1	2	13	2	2	65
gi 524884550 ref XP_005098894.1	2	6	2	2	51
gi 524884737 ref XP_005098987.1	2	10	2	9	249
gi 524884830 ref XP_005099031.1	2	6	2	3	33
gi 524884921 ref XP_005099073.1	2	5	2	2	57
gi 524885143 ref XP_005099182.1	2	9	3	3	93
gi 524885349 ref XP_005099283.1	2	51	4	10	78
gi 524885451 ref XP_005099333.1	2	7	2	2	36
gi 524885518 ref XP_005099365.1	2	11	2	2	60
gi 524885623 ref XP_005099417.1	2	34	4	5	48
gi 524885625 ref XP_005099418.1	2	63	3	5	87
gi 524885654 ref XP_005099432.1	2	9	2	2	35
gi 524885681 ref XP_005099445.1	2	6	2	2	27
gi 524885955 ref XP_005099578.1	2	6	2	2	66
gi 524886190 ref XP_005099692.1	2	3	2	3	72
gi 524886289 ref XP_005099741.1	2	11	2	2	30
gi 524886313 ref XP_005099753.1	2	10	2	6	141
gi 524886402 ref XP_005099797.1	2	4	2	3	70
gi 524886408 ref XP_005099800.1	2	12	2	4	42
gi 524886760 ref XP_005099972.1	2	19	2	3	42
gi 524887166 ref XP_005100172.1	2	18	3	4	54
gi 524887278 ref XP_005100227.1	2	6	2	2	56
gi 524887465 ref XP_005100318.1	2	30	2	2	42
gi 524887733 ref XP_005100450.1	2	21	2	2	28
gi 524888476 ref XP_005100809.1	2	24	2	3	131
gi 524888516 ref XP_005100828.1	2	4	2	2	59
gi 524888693 ref XP_005100913.1	2	5	2	3	24
gi 524888709 ref XP_005100920.1	2	9	4	4	46
gi 524888713 ref XP_005100922.1	2	5	2	2	89
gi 524888919 ref XP_005101023.1	2	9	2	2	40
gi 524889160 ref XP_005101141.1	2	18	2	4	108
gi 524889160 ref XP_005101141.1	2	32	2	3	72
gi 524889486 ref XP_005101298.1	2	16	2	2	48
gi 524889574 ref XP_005101342.1	2	8	2	2	53
gi 524889734 ref XP_005101421.1	2	18	2	2	50
gi 524890096 ref XP_005101597.1	2	23	2	4	141
gi 524890208 ref XP_005101652.1	2	43	11	16	271

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524890233 ref XP_005101664.1	2	3	2	2	54
gi 524890263 ref XP_005101679.1	2	4	2	2	37
gi 524890384 ref XP_005101739.1	2	18	2	2	53
gi 524890516 ref XP_005101804.1	2	12	2	4	63
gi 524890817 ref XP_005101950.1	2	13	2	3	59
gi 524890881 ref XP_005101982.1	2	12	2	3	83
gi 524890899 ref XP_005101991.1	2	19	2	2	53
gi 524891164 ref XP_005102118.1	2	17	2	3	116
gi 524891320 ref XP_005102195.1	2	54	2	3	72
gi 524891426 ref XP_005102246.1	2	10	2	2	26
gi 524891440 ref XP_005102253.1	2	14	2	2	101
gi 524891478 ref XP_005102272.1	2	10	2	5	88
gi 524891582 ref XP_005102324.1	2	23	2	6	66
gi 524892070 ref XP_005102566.1	2	27	2	2	97
gi 524892281 ref XP_005102670.1	2	4	4	10	360
gi 524892866 ref XP_005102957.1	2	11	2	2	40
gi 524892979 ref XP_005103011.1	2	34	2	2	34
gi 524893065 ref XP_005103053.1	2	16	2	2	80
gi 524893160 ref XP_005103100.1	2	17	3	7	57
gi 524893168 ref XP_005103104.1	2	33	2	3	154
gi 524893177 ref XP_005103108.1	2	4	2	5	123
gi 524893771 ref XP_005103394.1	2	7	2	2	42
gi 524893797 ref XP_005103407.1	2	3	2	2	37
gi 524893965 ref XP_005103490.1	2	12	2	4	204
gi 524894013 ref XP_005103514.1	2	21	6	9	137
gi 524894026 ref XP_005103520.1	2	6	2	3	114
gi 524894126 ref XP_005103566.1	2	10	2	2	114
gi 524894456 ref XP_005103725.1	2	12	2	2	44
gi 524894778 ref XP_005103881.1	2	4	2	2	39
gi 524894895 ref XP_005103937.1	2	24	3	8	154
gi 524894895 ref XP_005103937.1	2	32	3	12	314
gi 524894939 ref XP_005103959.1	2	13	2	3	63
gi 524895123 ref XP_005104049.1	2	27	6	9	164
gi 524895689 ref XP_005104326.1	2	15	2	4	37
gi 524895775 ref XP_005104369.1	2	9	2	3	72
gi 524895899 ref XP_005104430.1	2	17	2	7	115
gi 524896095 ref XP_005104525.1	2	13	2	3	57
gi 524896371 ref XP_005104660.1	2	9	2	3	62
gi 524896476 ref XP_005104712.1	2	16	2	4	61
gi 524896554 ref XP_005104750.1	2	16	2	5	118
gi 524896814 ref XP_005104878.1	2	15	3	4	77
gi 524897050 ref XP_005104992.1	2	8	2	3	147

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524897076 ref XP_005105005.1	2	26	2	3	113
gi 524897447 ref XP_005105184.1	2	9	2	2	48
gi 524897481 ref XP_005105201.1	2	11	2	2	55
gi 524897756 ref XP_005105335.1	2	13	2	2	45
gi 524898422 ref XP_005105660.1	2	2	2	2	55
gi 524898581 ref XP_005105738.1	2	10	2	2	61
gi 524898743 ref XP_005105817.1	2	18	2	2	54
gi 524898799 ref XP_005105845.1	2	2	2	2	51
gi 524898815 ref XP_005105852.1	2	34	2	2	49
gi 524899018 ref XP_005105950.1	2	11	2	2	65
gi 524899677 ref XP_005106267.1	2	31	2	9	140
gi 524899715 ref XP_005106286.1	2	5	2	2	30
gi 524899892 ref XP_005106372.1	2	13	2	6	150
gi 524900256 ref XP_005106549.1	2	16	2	3	40
gi 524900316 ref XP_005106579.1	2	3	2	2	81
gi 524900332 ref XP_005106587.1	2	18	2	3	48
gi 524900386 ref XP_005106613.1	2	5	2	2	46
gi 524900555 ref XP_005106697.1	2	7	2	2	44
gi 524900577 ref XP_005106708.1	2	14	2	2	45
gi 524900723 ref XP_005106780.1	2	6	2	3	82
gi 524900911 ref XP_005106871.1	2	20	2	3	51
gi 524901283 ref XP_005107052.1	2	46	5	7	129
gi 524901394 ref XP_005107104.1	2	31	5	7	265
gi 524901519 ref XP_005107166.1	2	13	2	2	40
gi 524901565 ref XP_005107188.1	2	13	2	5	119
gi 524901877 ref XP_005107342.1	2	16	2	2	51
gi 524902425 ref XP_005107464.1	2	15	3	3	82
gi 524902702 ref XP_005107579.1	2	9	2	3	99
gi 524902931 ref XP_005107685.1	2	11	2	3	125
gi 524903184 ref XP_005107804.1	2	4	2	2	43
gi 524903262 ref XP_005107836.1	2	8	2	3	63
gi 524903740 ref XP_005108064.1	2	5	2	3	76
gi 524905586 ref XP_005108170.1	2	15	2	4	96
gi 524905668 ref XP_005108199.1	2	8	2	4	194
gi 524905977 ref XP_005108344.1	2	11	2	2	48
gi 524906238 ref XP_005108452.1	2	6	2	2	63
gi 524906293 ref XP_005108469.1	2	34	2	5	86
gi 524906398 ref XP_005108509.1	2	6	2	2	54
gi 524906745 ref XP_005108595.1	2	27	5	5	56
gi 524906862 ref XP_005108596.1	2	26	2	3	69
gi 524906990 ref XP_005108638.1	2	8	2	3	47
gi 524907177 ref XP_005108713.1	2	12	2	4	63

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524907372 ref XP_005108804.1	2	9	2	3	69
gi 524907491 ref XP_005108862.1	2	34	2	2	40
gi 524907580 ref XP_005108906.1	2	7	2	3	97
gi 524907580 ref XP_005108906.1	2	7	3	3	80
gi 524907699 ref XP_005108965.1	2	14	2	3	50
gi 524907836 ref XP_005109029.1	2	37	2	4	132
gi 524908062 ref XP_005109139.1	2	25	2	3	79
gi 524908579 ref XP_005109383.1	2	4	2	2	70
gi 524908585 ref XP_005109386.1	2	13	2	2	44
gi 524908782 ref XP_005109481.1	2	6	2	2	26
gi 524908892 ref XP_005109535.1	2	6	2	2	43
gi 524909938 ref XP_005109861.1	2	7	2	2	37
gi 524909980 ref XP_005109881.1	2	18	2	3	106
gi 524910025 ref XP_005109900.1	2	18	2	4	102
gi 524910197 ref XP_005109980.1	2	8	2	3	174
gi 524910546 ref XP_005110149.1	2	16	2	7	213
gi 524911076 ref XP_005110408.1	2	2	2	2	59
gi 524911122 ref XP_005110430.1	2	14	2	5	112
gi 524911542 ref XP_005110633.1	2	5	2	5	200
gi 524911570 ref XP_005110646.1	2	6	2	2	73
gi 524911866 ref XP_005110793.1	2	15	2	9	186
gi 524912187 ref XP_005110945.1	2	13	2	3	91
gi 524912191 ref XP_005110947.1	2	42	2	2	63
gi 524912369 ref XP_005111034.1	2	8	2	3	168
gi 524912387 ref XP_005111042.1	2	8	2	2	36
gi 524912628 ref XP_005111158.1	2	6	2	4	82
gi 524912696 ref XP_005111190.1	2	16	2	4	42
gi 524913352 ref XP_005111507.1	2	17	2	4	57
gi 524913662 ref XP_005111656.1	2	11	2	3	57
gi 524913685 ref XP_005111667.1	2	9	2	3	97
gi 524913834 ref XP_005111741.1	2	9	2	2	74
gi 524913836 ref XP_005111742.1	2	2	2	2	41
gi 524914201 ref XP_005111919.1	2	15	2	2	46
gi 524914370 ref XP_005112000.1	2	13	2	3	44
gi 524914697 ref XP_005112160.1	2	11	2	2	38
gi 524914776 ref XP_005112198.1	2	7	2	2	25
gi 524914926 ref XP_005112272.1	2	8	2	4	151
gi 524915376 ref XP_005112489.1	2	16	3	4	96
gi 524915538 ref XP_005112566.1	2	23	2	4	87
gi 524916317 ref XP_005112938.1	2	24	2	4	194
gi 524916317 ref XP_005112938.1	2	16	2	6	86
gi 524916381 ref XP_005112969.1	2	17	2	3	39

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524916381 ref XP_005112969.1	2	16	2	2	52
gi 524916593 ref XP_005113070.1	2	42	10	20	434
gi 524916601 ref XP_005113074.1	2	34	3	12	259
gi 524916925 ref XP_005113230.1	2	5	2	3	46
gi 524917226 ref XP_005113374.1	2	7	2	2	26
gi 526117417 ref NP_001267763.1	2	4	2	3	53
gi 526117443 ref NP_001267770.1	2	9	2	6	125
gi 541950160 ref XP_005432023.1	2	40	7	28	557
gi 546679302 gb ERL89789.1	2	24	2	2	64
gi 555706265 gb ESO09498.1	2	13	2	6	113
gi 556093608 gb ESO82261.1	2	10	2	2	50
gi 556093971 gb ESO82624.1	2	16	2	5	95
gi 556095217 gb ESO83870.1	2	13	2	4	107
gi 556095970 gb ESO84622.1	2	11	2	4	140
gi 556097226 gb ESO85878.1	2	5	2	2	39
gi 556097346 gb ESO85998.1	2	20	2	2	63
gi 556098140 gb ESO86792.1	2	38	2	8	193
gi 556098590 gb ESO87242.1	2	24	2	3	150
gi 556101476 gb ESO90128.1	2	5	3	5	150
gi 556101533 gb ESO90185.1	2	17	2	3	59
gi 556101906 gb ESO90558.1	2	6	2	6	116
gi 556102306 gb ESO90958.1	2	14	2	2	41
gi 556105877 gb ESO94529.1	2	25	2	6	172
gi 556106462 gb ESO95114.1	2	22	2	3	97
gi 556106462 gb ESO95114.1	2	9	2	3	61
gi 556107128 gb ESO95780.1	2	20	2	2	44
gi 556108350 gb ESO97002.1	2	3	2	3	70
gi 556109892 gb ESO98544.1	2	12	2	2	44
gi 556111890 gb ESP00542.1	2	13	2	2	58
gi 556112256 gb ESP00908.1	2	8	2	2	53
gi 556112262 gb ESP00914.1	2	42	2	2	39
gi 556113060 gb ESP01712.1	2	2	2	2	32
gi 556114398 gb ESP03050.1	2	14	2	4	198
gi 556114716 gb ESP03368.1	2	4	2	2	40
gi 556115870 gb ESP04522.1	2	7	2	4	115
gi 556116273 gb ESP04925.1	2	9	2	2	44
gi 577029901 gb AHH81785.1	2	74	3	9	279
gi 585188 sp P38411.1 GNAQ_LYMST	2	25	2	2	49
gi 632935059 ref XP_007887607.1	2	30	2	2	32
gi 632968605 ref XP_007900618.1	2	18	2	2	58
gi 637321743 ref XP_008112910.1	2	28	2	3	84
gi 67989656 gb AAY84715.1	2	7	2	7	293

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 71726733 gb AAZ39530.1	2	45	2	13	340
gi 77628181 gb ABB00056.1	2	71	3	36	688
gi 85362949 gb ABC69854.1	2	28	2	9	206
gi 91079426 ref XP_967808.1	2	13	2	5	70
gi 1174755 sp P43689.1 TPM2_BIOGL	1	14	3	4	34
gi 148724316 gb ABR08403.1	1	16	2	7	284
gi 170596696 ref XP_001902862.1	1	30	3	21	589
gi 189308122 gb ACD86945.1	1	70	7	273	4574
gi 21388656 dbj BAC00784.1	1	35	34	37	701
gi 29378333 gb AAO83848.1 AF484091_1	1	30	4	15	499
gi 294896502 ref XP_002775589.1	1	71	5	10	204
gi 302139997 gb ADK94900.1	1	86	6	32	523
gi 325296803 ref NP_001191642.1	1	27	2	2	55
gi 339247869 ref XP_003375568.1	1	19	2	2	60
gi 339247869 ref XP_003375568.1	1	62	9	29	525
gi 390349570 ref XP_782388.3	1	69	5	21	382
gi 393908216 gb EFO23038.2	1	6	2	4	58
gi 399932045 gb AFP57558.1	1	35	7	22	540
gi 405970776 gb EKC35652.1	1	12	2	2	39
gi 405975785 gb EKC40331.1	1	56	32	116	2523
gi 42559558 sp O97192.1 TPM_HELAS	1	11	4	8	140
gi 42560362 gb AAS20336.1	1	54	3	95	2345
gi 524865813 ref XP_005089733.1	1	23	4	6	119
gi 524868888 ref XP_005091242.1	1	29	2	4	166
gi 524871402 ref XP_005092474.1	1	28	9	10	299
gi 524871513 ref XP_005092529.1	1	8	2	2	38
gi 524873824 ref XP_005093654.1	1	10	2	3	88
gi 524874374 ref XP_005093923.1	1	46	5	8	117
gi 524874376 ref XP_005093924.1	1	64	7	14	219
gi 524876525 ref XP_005094978.1	1	33	5	6	89
gi 524877853 ref XP_005095623.1	1	13	2	4	86
gi 524879771 ref XP_005096557.1	1	10	3	4	107
gi 524880608 ref XP_005096966.1	1	22	2	3	115
gi 524881407 ref XP_005097355.1	1	9	2	2	101
gi 524881473 ref XP_005097388.1	1	23	2	3	93
gi 524881817 ref XP_005097558.1	1	20	9	11	208
gi 524881819 ref XP_005097559.1	1	20	9	11	208
gi 524882112 ref XP_005097701.1	1	69	13	39	716
gi 524882279 ref XP_005097784.1	1	5	2	2	39
gi 524883774 ref XP_005098513.1	1	6	2	2	26
gi 524885219 ref XP_005099219.1	1	16	3	3	87
gi 524886261 ref XP_005099727.1	1	47	41	130	2703

The list of proteins obtained from 1D-SDS-PAGE of *A. vulgaris mantel*

Hit in the NCBI nr database	UP	PC (%)	NM	NS	PS
gi 524887580 ref XP_005100374.1	1	10	4	5	69
gi 524888358 ref XP_005100750.1	1	4	2	6	83
gi 524890724 ref XP_005101905.1	1	84	18	58	1117
gi 524890869 ref XP_005101976.1	1	6	2	2	54
gi 524890899 ref XP_005101991.1	1	16	2	2	31
gi 524891896 ref XP_005102480.1	1	45	6	10	164
gi 524894261 ref XP_005103632.1	1	3	3	6	41
gi 524894818 ref XP_005103900.1	1	22	2	8	71
gi 524894858 ref XP_005103920.1	1	7	3	3	61
gi 524896814 ref XP_005104878.1	1	40	78	88	1633
gi 524899004 ref XP_005105943.1	1	15	3	13	446
gi 524901033 ref XP_005106931.1	1	48	2	7	173
gi 524901281 ref XP_005107051.1	1	20	2	2	85
gi 524901464 ref XP_005107139.1	1	23	5	6	48
gi 524903383 ref XP_005107892.1	1	62	4	7	174
gi 524903383 ref XP_005107892.1	1	32	5	18	416
gi 524905758 ref XP_005108242.1	1	38	2	2	66
gi 524906745 ref XP_005108595.1	1	12	3	3	33
gi 524906745 ref XP_005108595.1	1	28	3	5	112
gi 524906862 ref XP_005108596.1	1	31	2	7	153
gi 524907032 ref XP_005108645.1	1	21	2	6	148
gi 524907032 ref XP_005108645.1	1	28	4	6	88
gi 524911570 ref XP_005110646.1	1	14	3	4	118
gi 524911945 ref XP_005110830.1	1	63	5	15	360
gi 524912496 ref XP_005111095.1	1	9	3	3	57
gi 524912496 ref XP_005111095.1	1	8	3	3	66
gi 524912621 ref XP_005111155.1	1	28	3	3	51
gi 524912715 ref XP_005111198.1	1	41	27	41	982
gi 524913881 ref XP_005111763.1	1	3	2	5	67
gi 537705579 dbj BAN81895.1	1	30	4	8	189
gi 556097681 gb ESO86333.1	1	3	2	3	52
gi 556102658 gb ESO91310.1	1	10	2	22	457
gi 556729406 ref XP_005960416.1	1	61	11	25	747
gi 56792845 gb AAW30622.1	1	42	4	5	137
gi 585719304 ref XP_006826038.1	1	39	64	121	2420
gi 597867153 gb EYC16545.1	1	17	2	5	79
gi 625230674 ref XP_007606483.1	1	56	4	15	248
gi 773571 emb CAA60122.1	1	7	2	2	109
gi 829207 emb CAA39415.1	1	76	39	53	737
gi 85753308 gb AAG49533.3	1	20	4	19	264

8.4. Table 3. Lectin-like proteins detected in transcriptomics level (status: I-incomplete, C-complete)

prot_hit	prot_hit_desc	status	evidence	protein
gi 524881819 ref XP_005097559.1	PREDICTED: calnexin-like isoform X2 [Aplysia californica]	I	T	calnexin
gi 524881819 ref XP_005097559.1	PREDICTED: calnexin-like isoform X2 [Aplysia californica]	I	T	calnexin
gi 2073142 dbj BAA19861.1	Incilarin A [Meghimatium fruhstorferi]	C	T	C-type lectin
gi 2073142 dbj BAA19861.1	Incilarin A [Meghimatium fruhstorferi]	C	T	C-type lectin
gi 2073146 dbj BAA19863.1	Incilarin C [Meghimatium fruhstorferi]	C	T	C-type lectin
gi 405959962 gb EKC25931.1	hypothetical protein CGI_10009560 [Crassostrea gigas]	C	T	C-type lectin
gi 524894115 ref XP_005103561.1	PREDICTED: uncharacterized protein LOC101845181 [Aplysia californica]	C	T	C-type lectin
gi 524906450 ref XP_005108528.1	PREDICTED: secretory phospholipase A2 receptor-like [Aplysia californica]	C	T	C-type lectin
gi 524907181 ref XP_005108715.1	PREDICTED: uncharacterized protein LOC101848467 [Aplysia californica]	C	T	C-type lectin
gi 556103033 gb ESO91685.1	hypothetical protein LOTGIDRAFT_163414 [Lottia gigantea]	C	T	C-type lectin
gi 556111065 gb ESO99716.1	hypothetical protein LOTGIDRAFT_238723 [Lottia gigantea]	C	T	C-type lectin
gi 126697272 gb ABO26593.1	putative perlucin 4 [Haliotis discus discus]	I	T	C-type lectin
gi 126697272 gb ABO26593.1	putative perlucin 4 [Haliotis discus discus]	I	T	C-type lectin
gi 157694790 gb ABV66064.1	type II antifreeze protein [Lates calcarifer]	I	T	C-type lectin
gi 2073142 dbj BAA19861.1	Incilarin A [Meghimatium fruhstorferi]	I	T	C-type lectin
gi 405959962 gb EKC25931.1	hypothetical protein CGI_10009560 [Crassostrea gigas]	I	T	C-type lectin
gi 405972881 gb EKC37628.1	Low affinity immunoglobulin epsilon Fc receptor [Crassostrea gigas]	I	T	C-type lectin

prot_hit	prot_hit_desc	status	evidence	protein
gi 524889950 ref XP_005101525.1	PREDICTED: FRAS1-related extracellular matrix protein 1-like [Aplysia californica]	I	T	C-type lectin
gi 524889950 ref XP_005101525.1	PREDICTED: FRAS1-related extracellular matrix protein 1-like [Aplysia californica]	I	T	C-type lectin
gi 524890590 ref XP_005101839.1	PREDICTED: cell wall protein DAN4-like [Aplysia californica]	I	T	C-type lectin
gi 524894115 ref XP_005103561.1	PREDICTED: uncharacterized protein LOC101845181 [Aplysia californica]	I	T	C-type lectin
gi 524895689 ref XP_005104326.1	PREDICTED: uncharacterized protein LOC101845349 [Aplysia californica]	I	T	C-type lectin
gi 524907181 ref XP_005108715.1	PREDICTED: uncharacterized protein LOC101848467 [Aplysia californica]	I	T	C-type lectin
gi 546241983 gb AGW83938.1	vitelline coat lysin M3, partial [Mytilus trossulus]	I	T	C-type lectin
		I	T	C-type lectin
gi 226731904 gb ACO82046.1	C-type lectin 13 [Perca flavescens]	I	T	C-type lectin
gi 617468332 ref XP_007574500.1	PREDICTED: galactose-specific lectin natterctin-like, partial [Poecilia formosa]	I	T	C-type lectin
gi 524864072 ref XP_005088874.1	PREDICTED: uncharacterized protein LOC101856539 [Aplysia californica]	I	T	C-type lectin
gi 524907181 ref XP_005108715.1	PREDICTED: uncharacterized protein LOC101848467 [Aplysia californica]	I	T	C-type lectin
gi 556100655 gb ESO89307.1	hypothetical protein LOTGIDRAFT_229175 [Lottia gigantea]	C	T	C-type lectin
gi 556097543 gb ESO86195.1	hypothetical protein LOTGIDRAFT_235549 [Lottia gigantea]	C	T	C-type lectin
gi 556097543 gb ESO86195.1	hypothetical protein LOTGIDRAFT_235549 [Lottia gigantea]	C	T	C-type lectin
gi 524870399 ref XP_005091984.1	PREDICTED: macrophage mannose receptor 1-like [Aplysia californica]	I	T	C-type lectin
gi 556103033 gb ESO91685.1	hypothetical protein LOTGIDRAFT_163414 [Lottia gigantea]	C	T	C-type lectin
gi 152032020 gb ABS28869.1	tandem repeat galectin [Biomphalaria glabrata]	C	T	galectin
gi 524891001 ref XP_005102040.1	PREDICTED: galectin-4-like isoform X3 [Aplysia californica]	C	T	galectin

prot_hit	prot_hit_desc	status	evidence	protein
gi 524891001 ref XP_005102040.1	PREDICTED: galectin-4-like isoform X3 [Aplysia californica]	C	T	galectin
gi 524891001 ref XP_005102040.1	PREDICTED: galectin-4-like isoform X3 [Aplysia californica]	C	T	galectin
gi 402580802 gb EJW74751.1	galectin, partial [Wuchereria bancrofti]	I	T	galectin
gi 560121553 emb CDJ93815.1	Galectin domain containing protein, partial [Haemonchus contortus]	I	T	galectin
gi 152032020 gb ABS28869.1	tandem repeat galectin [Biomphalaria glabrata]	I	T	galectin
gi 568300466 gb ETN86155.1	galactoside-binding lectin [Necator americanus]	I	T	galectin
gi 405976166 gb EKC40683.1	Cubilin [Crassostrea gigas]	I	T	galectin
gi 85362949 gb ABC69854.1	agglutinin [Helix pomatia]	C	T	H-type lectin
gi 109157294 pdb 2CE6 A	Chain A, Structure Of Helix Pomatia Agglutinin With No Ligands	I	T	H-type lectin
gi 85362949 gb ABC69854.1	agglutinin [Helix pomatia]	I	T	H-type lectin
		I	T	H-type lectin
gi 405965843 gb EKC31192.1	Protein ERGIC-53 [Crassostrea gigas]	C	T	L-type lectin
gi 405965843 gb EKC31192.1	Protein ERGIC-53 [Crassostrea gigas]	C	T	L-type lectin
gi 556113252 gb ESP01904.1	hypothetical protein LOTGIDRAFT_138892, partial [Lottia gigantea]	I	T	R-type lectin
gi 524895404 ref XP_005104185.1	PREDICTED: polypeptide N-acetylgalactosaminyltransferase 5-like [Aplysia californica]	I	T	R-type lectin
gi 524896314 ref XP_005104633.1	PREDICTED: polypeptide N-acetylgalactosaminyltransferase 2-like [Aplysia californica]	C	T	R-type lectin
gi 556102066 gb ESO90718.1	hypothetical protein LOTGIDRAFT_123129, partial [Lottia gigantea]	I	T	R-type lectin
gi 524888837 ref XP_005100983.1	PREDICTED: N-acetylgalactosaminyltransferase 7-like isoform X1 [Aplysia californica]	I	T	R-type lectin
gi 524865257 ref XP_005089459.1	PREDICTED: polypeptide N-acetylgalactosaminyltransferase 11-like isoform X2 [Aplysia californica]	I	T	R-type lectin

prot_hit	prot_hit_desc	status	evidence	protein
gi 321455342 gb EFX66478.1	hypothetical protein DAPPUDRAFT_302681 [Daphnia pulex]	C	T	R-type lectin
gi 405966386 gb EKC31679.1	Polypeptide N-acetylgalactosaminyltransferase 5 [Crassostrea gigas]	I	T	R-type lectin
gi 524884259 ref XP_005098751.1	PREDICTED: putative polypeptide N-acetylgalactosaminyltransferase 10-like, partial [Aplysia californica]	I	T	R-type lectin
gi 556113252 gb ESP01904.1	hypothetical protein LOTGIDRAFT_138892, partial [Lottia gigantea]	I	T	R-type lectin
gi 556113252 gb ESP01904.1	hypothetical protein LOTGIDRAFT_138892, partial [Lottia gigantea]	I	T	R-type lectin
gi 556113252 gb ESP01904.1	hypothetical protein LOTGIDRAFT_138892, partial [Lottia gigantea]	I	T	R-type lectin
gi 556113252 gb ESP01904.1	hypothetical protein LOTGIDRAFT_138892, partial [Lottia gigantea]	I	T	R-type lectin
gi 556113252 gb ESP01904.1	hypothetical protein LOTGIDRAFT_138892, partial [Lottia gigantea]	I	T	R-type lectin
gi 556113252 gb ESP01904.1	hypothetical protein LOTGIDRAFT_138892, partial [Lottia gigantea]	I	T	R-type lectin
gi 556113252 gb ESP01904.1	hypothetical protein LOTGIDRAFT_138892, partial [Lottia gigantea]	I	T	R-type lectin
gi 556113252 gb ESP01904.1	hypothetical protein LOTGIDRAFT_138892, partial [Lottia gigantea]	I	T	R-type lectin
gi 556113252 gb ESP01904.1	hypothetical protein LOTGIDRAFT_138892, partial [Lottia gigantea]	I	T	R-type lectin
gi 556113252 gb ESP01904.1	hypothetical protein LOTGIDRAFT_138892, partial [Lottia gigantea]	I	T	R-type lectin
gi 556113252 gb ESP01904.1	hypothetical protein LOTGIDRAFT_138892, partial [Lottia gigantea]	I	T	R-type lectin
gi 556113252 gb ESP01904.1	hypothetical protein LOTGIDRAFT_138892, partial [Lottia gigantea]	I	T	R-type lectin
gi 556113252 gb ESP01904.1	hypothetical protein LOTGIDRAFT_138892, partial [Lottia gigantea]	I	T	R-type lectin

prot_hit	prot_hit_desc	status	evidence	protein
gi 556113252 gb ESP01904.1	hypothetical protein LOTGIDRAFT_138892, partial [Lottia gigantea]	C	T	R-type lectin
gi 556113252 gb ESP01904.1	hypothetical protein LOTGIDRAFT_138892, partial [Lottia gigantea]	C	T	R-type lectin
gi 556113252 gb ESP01904.1	hypothetical protein LOTGIDRAFT_138892, partial [Lottia gigantea]	C	T	R-type lectin
gi 556093434 gb ESO82087.1	hypothetical protein LOTGIDRAFT_198292 [Lottia gigantea]	I	T	R-type lectin
gi 556093434 gb ESO82087.1	hypothetical protein LOTGIDRAFT_198292 [Lottia gigantea]	I	T	R-type lectin
gi 524864445 ref XP_005089059.1	PREDICTED: polypeptide N-acetylgalactosaminyltransferase 1-like isoform X2 [Aplysia californica]	I	T	R-type lectin
gi 524864445 ref XP_005089059.1	PREDICTED: polypeptide N-acetylgalactosaminyltransferase 1-like isoform X2 [Aplysia californica]	I	T	R-type lectin
gi 524864445 ref XP_005089059.1	PREDICTED: polypeptide N-acetylgalactosaminyltransferase 1-like isoform X2 [Aplysia californica]	I	T	R-type lectin
gi 524864445 ref XP_005089059.1	PREDICTED: polypeptide N-acetylgalactosaminyltransferase 1-like isoform X2 [Aplysia californica]	I	T	R-type lectin
gi 556102066 gb ESO90718.1	hypothetical protein LOTGIDRAFT_123129, partial [Lottia gigantea]	I	T	R-type lectin
gi 524890796 ref XP_005101940.1	PREDICTED: polypeptide N-acetylgalactosaminyltransferase 5-like [Aplysia californica]	I	T	R-type lectin
gi 556112926 gb ESP01578.1	hypothetical protein LOTGIDRAFT_91617, partial [Lottia gigantea]	C	T	R-type lectin
gi 524884463 ref XP_005098851.1	PREDICTED: techylectin-5A-like [Aplysia californica]	C	T	techylectin
gi 524914011 ref XP_005111826.1	PREDICTED: techylectin-5B-like [Aplysia californica]	I	T	techylectin
gi 524914011 ref XP_005111826.1	PREDICTED: techylectin-5B-like [Aplysia californica]	I	T	techylectin
gi 3347852 gb AAC27743.1	sialic acid-binding lectin 2 [Limax flavus]	I	T	lectin
gi 3347852 gb AAC27743.1	sialic acid-binding lectin 2 [Limax flavus]	I	T	lectin

prot_hit	prot_hit_desc	status	evidence	protein
gi 524900094 ref XP_005106471.1	PREDICTED: endoplasmic reticulum lectin 1-like [Aplysia californica]	I	T	lectin
gi 524875712 ref XP_005094580.1	PREDICTED: EMILIN-2-like [Aplysia californica]	C	T	C1qDC
gi 583980443 ref XP_006785537.1	PREDICTED: complement C1q-like protein 2-like [Neolamprologus brichardi]	C	T	C1qDC
		C	T	C1qDC
gi 325504319 emb CBX41659.1	putative C1q domain containing protein MgC1q10 [Mytilus galloprovincialis]	I	T	C1qDC
gi 325504319 emb CBX41659.1	putative C1q domain containing protein MgC1q10 [Mytilus galloprovincialis]	I	T	C1qDC
gi 325504319 emb CBX41659.1	putative C1q domain containing protein MgC1q10 [Mytilus galloprovincialis]	I	T	C1qDC
gi 325504327 emb CBX41663.1	putative C1q domain containing protein MgC1q14 [Mytilus galloprovincialis]	I	T	C1qDC
gi 325504381 emb CBX41690.1	putative C1q domain containing protein MgC1q41 [Mytilus galloprovincialis]	I	T	C1qDC
gi 325504381 emb CBX41690.1	putative C1q domain containing protein MgC1q41 [Mytilus galloprovincialis]	I	T	C1qDC
gi 325504405 emb CBX41702.1	putative C1q domain containing protein MgC1q53 [Mytilus galloprovincialis]	I	T	C1qDC
gi 325504405 emb CBX41702.1	putative C1q domain containing protein MgC1q53 [Mytilus galloprovincialis]	I	T	C1qDC
gi 325504493 emb CBX41746.1	putative C1q domain containing protein MgC1q97 [Mytilus galloprovincialis]	I	T	C1qDC
gi 405955760 gb EKC22740.1	Complement C1q subcomponent subunit B [Crassostrea gigas]	I	T	C1qDC
gi 405963338 gb EKC28921.1	Collagen alpha-2(VIII) chain [Crassostrea gigas]	I	T	C1qDC
gi 405972817 gb EKC37565.1	Caprin-2 [Crassostrea gigas]	I	T	C1qDC
gi 472825914 gb AGI44588.1	C1q domain containing protein [Azumapecten farreri]	I	T	C1qDC

prot_hit	prot_hit_desc	status	evidence	protein
gi 472825914 gb AGI44588.1	C1q domain containing protein [Azumapecten farreri]	I	T	C1qDC
gi 524875712 ref XP_005094580.1	PREDICTED: EMILIN-2-like [Aplysia californica]	I	T	C1qDC
		I	T	C1qDC
gi 126697410 gb ABO26662.1	sialic acid binding lectin [Haliotis discus discus]	I	T	C1qDC
gi 38043955 emb CAD83837.1	sialic acid binding lectin [Cepaea hortensis]	I	T	C1qDC
gi 38043955 emb CAD83837.1	sialic acid binding lectin [Cepaea hortensis]	I	T	C1qDC
gi 93209532 gb ABF00124.1	sialic acid binding lectin [Helix pomatia]	I	T	C1qDC
gi 93209532 gb ABF00124.1	sialic acid binding lectin [Helix pomatia]	I	T	C1qDC
gi 40218026 gb AAR82936.1	macrophage expressed protein [Haliotis rufescens]	C	T	pore-forming membrane attack
gi 524916402 ref XP_005112978.1	PREDICTED: uncharacterized protein LOC101845147 [Aplysia californica]	C	T	pore-forming membrane attack
gi 119112553 ref XP_001237961.1	AGAP007821-PA [Anopheles gambiae str. PEST]	C	T	toxin-like
gi 193685923 ref XP_001950560.1	PREDICTED: uncharacterized protein LOC100163563 isoform X1 [Acyrtosiphon pisum]	C	T	toxin-like

8.5. Figure 1. Alignment of amino acid sequences of H-type lectins from *A. vulgaris* with *Helix pomatia* and *Cepaea hortensis*

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HACG01016007 ----MNTFTAGLVLLCLLSLSTAL---LQEGIIQCGTDDWKKTYDDRIQLNERQSANNVKFPPTAHRSLPGVVVDVSQLDSELSQNLRYQLRISAITKTGFSLSCVTWPKTHIFSLNVRWVFFDRKTC--
HACG01031738 ITDRMDSFMKGLVLLCLVSLSAAV---IQDGIIVACGADSSWKPTGAKTSRAVE-RQSANSIQFKKAYKRAPKVLVSVIQLDSEQSQNLRYEIRLNSVANIGFKFSCVTWVHYTKIYSLTVRWVSVDE-----
HACG01031741 ITDRMDSFMKGLVLLCLVSLSAAH---SLHGTVKCGADSSWKPTGAKTSRAAE-RQSTNSIQFKKPYESIPPTVVLVSVIQLDSEQSENLRYEIRLHSVTKTGFKFSCVTWVHYTVIYSLNVHWVSVDE-----
HACG01039153 ----MNSIMKGFVCLLPLTSAL---IQEGIIKCGNDHHWRLTGYDPTIIRLNERESVNNIKFPIPYTSTPAVTVSVHQLDTRFQNLRYELRLTSVITGFKLSCVTWHHTGIYSLNVRWVSLSDSAQQID
HACG01039154 ----MNSIMKGFVCLLPLTSAL---IQEGIIKCGNDHHWRLTGYDPTIIRLNERESVNNIKFPIPYTSTPAVTVSVHQLDTRFQNLRYELRLTSVITGFKLSCVTWHHTGIYSLNVRWVSLSDSAQQID
HACG01041396 ----MSSLISGLILAVMLSFCAAK---VQIGNFTCVNSETWVPTGDFKVKIVD---V---VPEPEYKIVPKVHLSIVHLDIDKIKTLRVKTYVSGITNIGFKAIFTIWDVSVVKTAIVSNLSIES-----
HACG01043414 ----MSSLINGLILAILISFCARESEMEEGKFRCGDSGFRSIPNATEKISTHN---V---TFTTFNEVPKVYLFHFKGS-G---DSRMWILDSVSTIGFNVTCHMSENSGYSRIVTDYFAINEYF----
HACG01005310 -----MLSFCAAN---VQTGSFTCENSATWANVGGLPCTCKK---FVFDVFPYTSAPNVILITLDW---DIQNSKNLQIVTNVTEVSKTGFATAAFAWGKIVNKKVIVSWLSTDAVSWSN
Cepaea hortensis ----MQAFTAGLLLCVLSFAAAQ--CVQTGTIECGNDDSNPSVPSDDPSKVNTRELAKTITFSSSYCRSPVVLLSITQLDVEKQNLRVISRLYIASPTGFKASCYTWNTKVRMTESWIAIEN-----
Helix pomatia ----MQAFYSGLLLCVFAAAQ--RVQSGKIDCGNDAGNAKVPSSDDPGSDNTRRELAKNITFASPYCRFPVVLLSITQLDVEQSQNLRVIAARLYSVSPGFKASCYTWNTKVVYSMSISWISIEYNY-----

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